

Stevenson-Holt, Claire D. ORCID: <https://orcid.org/0000-0003-3635-5404> ,
Ramsey, Andrew D. ORCID: <https://orcid.org/0000-0002-5550-9977> , Nevin,
Owen ORCID: <https://orcid.org/0000-0003-3513-8053> and Sinclair, Billy (2012)
Assessing grey squirrel dispersal patterns within the landscape using sequence
variation. *Landscape Research*, 38 (4). pp. 553-559.

Downloaded from: <http://insight.cumbria.ac.uk/id/eprint/1984/>

Usage of any items from the University of Cumbria's institutional repository 'Insight' must conform to the following fair usage guidelines.

Any item and its associated metadata held in the University of Cumbria's institutional repository Insight (unless stated otherwise on the metadata record) may be copied, displayed or performed, and stored in line with the JISC fair dealing guidelines (available [here](#)) for educational and not-for-profit activities

provided that

- the authors, title and full bibliographic details of the item are cited clearly when any part of the work is referred to verbally or in the written form
 - a hyperlink/URL to the original Insight record of that item is included in any citations of the work
- the content is not changed in any way
- all files required for usage of the item are kept together with the main item file.

You may not

- sell any part of an item
- refer to any part of an item without citation
- amend any item or contextualise it in a way that will impugn the creator's reputation
- remove or alter the copyright statement on an item.

The full policy can be found [here](#).

Alternatively contact the University of Cumbria Repository Editor by emailing insight@cumbria.ac.uk.



Assessing grey squirrel dispersal patterns within the landscape using DNA sequence variation.

Journal:	<i>Landscape Research</i>
Manuscript ID:	Draft
Manuscript Type:	Short Communication
Keywords:	least cost, mtDNA, grey squirrel, <i>Sciurus</i> , dispersal

SCHOLARONE™
Manuscripts

1
2
3 1 Assessing grey squirrel dispersal patterns within the landscape using sequence variation.
4
5
6 2

7
8 3 ABSTRACT
9

10 4 The grey squirrel *Sciurus carolinensis* is thought to have contributed to the decline of
11
12 5 red squirrel *S. vulgaris* populations in the UK through resource competition and disease
13
14 6 spread. This study used mtDNA sequencing to assess patterns of grey squirrel dispersal in the
15
16 7 UK. Patterns of genetic variation within the dloop sequence were characterised for seven grey
17
18 8 squirrel populations. Infiltration directions and potential barriers to dispersal are identified
19
20 9 and discussed, with a focus on Cumbria, a county at the forefront of grey squirrel expansion.
21
22 10 Understanding the dynamics of grey squirrel dispersal will aid their management at a
23
24 11 landscape scale and enhance the conservation of red squirrels.
25
26
27
28
29
30
31
32

33
34 13 INTRODUCTION
35

36 14 The Eastern grey squirrel *Sciurus carolinensis* was first introduced to Britain between
37
38 15 1876 and 1929 (Middleton 1931; Shorten 1954; [Lloyd 1983](#)). Subsequent successful
39
40 16 introductions and translocations occurred within the UK and by the 1930's populations were
41
42 17 established in southeast England and rapidly spreading through the country (Shorten 1954).
43
44 18 This expansion occurred simultaneously with the decline in native red squirrel *Sciurus*
45
46 19 *vulgaris* populations ([Lloyd 1962](#); [Lloyd 1983](#)). Studies suggest that interspecific resource
47
48 20 competition occurs ([Wauters et al. 2000](#); [Gurnell et al. 2004](#)) and that this, along with the
49
50 21 effects of a particularly virulent squirrelpox virus (SQPV), carried by the grey squirrel, has
51
52 22 caused the decline and extirpation of many red squirrel populations ([Tompkins et al. 2003](#);
53
54 23 [Gurnell et al. 2004](#); [Carroll et al. 2009](#)).

55
56
57 24 Cumbria in North West England has been found to hold genetically unique
58
59 25 populations of red squirrel which contain high levels of genetic diversity ([Hale et al. 2004](#)).

1
2
3 26 Concerns have been raised over the impact of expanding grey squirrel populations are having
4
5 27 on red squirrel populations. Interspecific competition and SQPV transmission are thought to
6
7
8 28 be highest at times of grey squirrel dispersal (Sainsbury et al. 2008). An understanding of the
9
10 29 dispersal ecology and directional movements of the grey squirrel will aid grey squirrel
11
12 30 management decisions and red squirrel conservation.

13
14
15 31 Recent work (Stevenson et al., in review) has indicated that the Cumbrian Mountain
16
17 32 range is acting as a barrier to dispersal. Grey squirrel populations within Cumbria have been
18
19 33 derived from the infiltration of individuals from two directions; to south Cumbria from
20
21 34 Lancashire and to north Cumbria from Northumberland/ Scottish Borders. Hale et al. (2001)
22
23 35 and Trizio et al. (2005) both suggest genetic analysis has enabled the identification of land
24
25 36 cover types which either facilitate dispersal or provide barriers to red squirrels over large
26
27 37 geographic scales. In this paper we report on variation within Cytochrome b (Cytb) and Dloop
28
29 38 DNA sequences found within grey squirrel mtDNA, the first such report to date.
30
31
32
33

34 39 35 36 40 METHODOLOGY 37

38 41 Two grey squirrel accessions were collected from each of four known introduction
39
40 42 points (see Middleton, 1931); Balloch near Loch Long; Dalkeith in Edinburgh; Henbury in
41
42 43 Cheshire, and from Alice Holt Forest in Surrey. In addition, samples were collected from
43
44 44 three established/ emerging populations within the UK; Doune, in Stirlingshire; Millom, in
45
46 45 south Cumbria and Brampton, in north Cumbria (Figure 1).
47
48
49

50 46 DNA was extracted from the leg muscle tissue of 14 individuals using the QIAGEN
51
52 47 DNeasy blood and tissue extraction kit and following the manufactures instructions
53
54 48 (QIAGEN Ltd). Fragments of the two mtDNA sequence encoding Cytb and Dloop were
55
56 49 amplified by polymerase chain reaction (PCR). Primers for *Sciurus carolinensis* Cytb were
57
58 50 available from Meece et al. (2005), BM1 (5'-CCCCTCAGAATGATATTTGTCCTCA) and
59
60

1
2
3 51 BM2 (5'-CCATCCAACATCTCAGCATGATGAAA). Each PCR reaction had a reaction
4
5 52 volume of 25 µl and containing 1 µl 1:10 mtDNA, 12.5 µl AmpliTaq Gold PCR master mix
6
7
8 53 (Roche, USA), 1 µl 1:10 BM1 primer to distilled water, 1 µl 1:10 BM2, 9.5 µl distilled water.
9
10 54 PCR amplification followed the protocol of Meece et al. (2005): denaturation of 3.5 min at
11
12 55 95°C followed by 36 cycles of 30s at 95°C, annealing 50s at 60 °C, extension 40s at 72°C,
13
14 56 final extension of 5min at 72 °C.

15
16
17 57 Primers for *Sciurus carolinensis* Dloop were taken from the published sequence of the
18
19 58 Dloop (GenBank Accession no. AF111027; Barrett et al. 1999), Dloop forward primer 5'-
20
21 59 GCCACCCCAAGTTAAATGG-3' and Dloop reverse primer 5'-
22
23 60 ATTCGTGCATTAATGCACTATCC-3'. Each PCR reaction for Dloop contained the same
24
25 61 relative quantities of components as outlined above for cytb amplification apart from 5 µl of
26
27 62 each primer was used. PCR amplification for Dloop sequence followed the protocol of Trizio
28
29 63 et al. (2005), denaturation of 1 min at 94°C followed by 40 cycles of 30s at 94°C, annealing
30
31 64 30s at 52 °C, extension 1min at 72°C, final extension of 5min at 72 °C.

32
33
34 65 Electrophoresis on 1% agarose gels were used to check the success of the PCR
35
36 66 reactions for each sample. All PCR products were then cleaned following the methodology of
37
38 67 the QIAquick PCR purification kit (QIAGEN Ltd). Sequencing of PCR products was carried
39
40 68 out (DBS Genomics, Durham, UK).

41
42
43 69 Sequences for cytb and Dloop from each individual were aligned manually using
44
45 70 BioEdit_R v7.0.4.1 Freeware (Ibis Therapeutics, California,USA) and Sequence Analysis
46
47 71 5.2.0 (Applied Biosystems) using the default settings within the programmes. A sequence
48
49 72 from *S. vulgaris* (GenBank Accession no. AJ238588) was also included as an outgroup
50
51 73 accession in the analysis. The sequences were entered into Paup 4.0 Beta 10 (Swofford,
52
53 74 Illinois Natural History Survey, Illinois, USA) for phylogeny reconstruction and a 50%

1
2
3 75 majority rule consensus tree was created with 1000 bootstrap and jackknife replicates (Harrison
4
5 76 and Langdale 2006).
6
7
8 77
9

10 78 RESULTS

11
12 79 MtDNA was successfully extracted from 14 grey squirrel individuals from selected
13
14 80 locations around the UK. Cytb sequence data was analysed for all 14 accessions with no
15
16 81 sequence variation observed. The 325bp sequence generated, demonstrated a 99% similarity
17
18 82 match for grey squirrel cytochrome b gene in the Genbank BLAST search tool (Accession no.
19
20 83 AY509680). Similarly, the 329bp Dloop sequence generated showed a 98% similarity match
21
22 84 to that of grey squirrel (Accession no. AF111027). Dloop sequences were aligned in BioEdit
23
24 85 and discrete points of variation were detected at 16 unique sites, representing 4.9 % of the
25
26 86 overall sequence.
27
28
29
30

31 87 Parsimony analysis generated a 50% majority rule consensus tree from the grey
32
33 88 squirrel dloop sequence data which was rooted against red squirrel (GenBank Accession no.
34
35 89 AJ238588). The consensus tree (Figure 2) indicated that the grey squirrel samples are
36
37 90 separated into four distinct clades; Clade I; Henbury, Balloch and accession S8 from Alice
38
39 91 Holt; Clade II; Doune and accession S7 from Alice Holt; Clade III; Dalkeith and Brampton
40
41 92 (north Cumbria); Clade IV; Millom (south Cumbria). The branching of samples from
42
43 93 Dalkeith and Brampton is strongly supported with bootstrap values of 73% and jackknife
44
45 94 values of 56%.
46
47
48
49
50

51 95

52 96 DISCUSSION

53
54
55 97 This study examines DNA sequence variation and its utility in assessing patterns of
56
57 98 grey squirrel dispersal in the UK. Cytb sequences generated in this study demonstrated no
58
59 99 detectable differences across the range of populations sampled. This may be due to the low
60

1
2
3 100 mutation rate within the cytb coding sequence and the relatively short time frame of
4
5 101 introduction and dispersal within the UK. Conversely, sequence variation was detected
6
7
8 102 within the dloop sequence for these same accessions. Dloop sequence analysis showed
9
10 103 significant statistical support for the distinct separation of north and south Cumbria grey
11
12 104 squirrel populations.

13
14
15 105 Accessions from north Cumbria grouped with accessions from Dalkeith in Edinburgh.
16
17 106 Individual accessions within north Cumbria may not necessarily have been derived from
18
19 107 individuals dispersing south from Edinburgh. However, as they are both within the same
20
21 108 clade they may have been derived from the same progenitor individuals from another location
22
23 109 such as northeast England (Stevenson et al., in review). Similar patterns of dispersal have
24
25 110 been seen in red squirrels, mediated by the afforestation of woodland between Cumbria and
26
27 111 the Northeast (Hale & Lurz 2003).

28
29
30
31 112 The two samples taken from each population were generally grouped together within
32
33 113 the phylogenetic tree, however, samples from Alice Holt Forest did not follow this pattern and
34
35 114 were grouped separately with samples from elsewhere. David-Gray et al. (1998) study found
36
37 115 high levels of genetic diversity within grey squirrel populations from Alice Holt Forest and
38
39 116 attributed this to numerous introduction sites and translocations from different source
40
41 117 populations, which could explain the results demonstrated in Figure 2 in this study.

42
43
44
45 118 The initial findings presented here demonstrate support for the hypotheses that the
46
47 119 Cumbrian Mountain range is acting as a barrier to dispersal and that invasion into the area is
48
49 120 coming from multiple directions. If grey squirrels colonised Cumbria with a northerly
50
51 121 advance as suggested (Lowe 1993; Skelcher 1997) accessions from north and south Cumbria
52
53 122 should be grouped together within a phylogenetic reconstruction. However, this is not the
54
55 123 case, and the two populations separate out in the tree. This points towards separate
56
57 124 populations and the effectiveness of the Cumbrian Mountain range as a barrier. Whilst it is

1
2
3 125 acknowledged that a small number of individuals may try to disperse over this mountain
4
5 126 barrier, the majority of individuals will choose a lower cost dispersal route. Further validation
6
7 127 of prospective incursion directions will require an increase in the sample size for each of the
8
9 128 seven locations and additional accessions from within Cumbria and adjoining areas.

10
11
12 129 This study has provided the first evidence of Dloop sequence variation within UK
13
14 130 grey squirrel populations. Despite the small sample size, it supports the suggestion that the
15
16 131 Cumbrian Mountains provide a barrier to grey squirrel dispersal and infers both northerly and
17
18 132 southerly grey squirrel incursion routes into the county. Ultimately, further knowledge of
19
20 133 dispersal and colonisation of grey squirrels will inform conservation policy and can be used to
21
22 134 target resources for grey squirrel control and enable better protection for vulnerable red
23
24 135 squirrel populations. We have shown here, that landscape genetics can provide evidence of
25
26 136 population origin and genetic differences. Although this study has focused on grey squirrels in
27
28 137 Cumbria, the techniques are equally applicable to other landscapes, validating and
29
30 138 highlighting dispersal routes of invasive species and species of conservation concern.

31
32
33
34
35
36 139

37 38 140 REFERENCES

- 39
40
41 141
42 142 [Carroll, B., P. Russell, J. Gurnell, P. Nettleton, and A. W. Sainsbury. 2009. Epidemics of](#)
43
44 143 [squirrelpox virus disease in red squirrels \(*Sciurus vulgaris*\): Temporal and serological](#)
45
46 144 [findings. *Epidemiology and Infection* **137**:257-265.](#)
- 47
48
49 145 David-Gray, Z. K., J. Gurnell, and D. M. Hunt. 1998. DNA fingerprinting reveals high levels
50
51 146 of genetic diversity within British populations of the introduced non-native grey squirrel
52
53 147 (*Sciurus carolinensis*). *Journal of zoology* **246**(4):443-445.
- 54
55
56 148 Felsenstein, J. 1985. Confidence limits on phylogenies: An approach using the bootstrap.
57
58 149 *Evolution* **39**(4):783-791.
- 59
60

- 1
2
3 150 Gurnell, J., L. A. Wauters, P. W. W. Lurz, and G. Tosi. 2004. Alien species and interspecific
4
5 151 competition: Effects of introduced eastern grey squirrels on red squirrel population dynamics.
6
7 152 *Journal of Animal Ecology* **73**(1):26-35.
8
9
10 153 Hale, M. L., P. W. W. Lurz, and K. Wolff. 2004. Patterns of genetic diversity in the red
11
12 154 squirrel (*Sciurus vulgaris* L.): Footprints of biogeographic history and artificial introductions.
13
14 155 *Conservation Genetics* **5**:167-179.
15
16
17 156 Hale, M. L., and P. W. W. Lurz. 2003. Morphological changes in a British mammal as a result
18
19 157 of introductions and changes in landscape management: The red squirrel (*Sciurus vulgaris*).
20
21 158 *Journal of zoology* **260**(2):159-167.
22
23
24 159 Hale, M. L., P. W. W. Lurz, M. D. F. Shirley, S. Rushton, R. Fuller, and K. Wolff. 2001.
25
26 160 Impact of landscape management on the genetic structure of red squirrel populations.
27
28
29 161 Harrison, C. J., and J. A. Langdale. 2006. A step by step guide to phylogeny reconstruction.
30
31 162 *The Plant Journal* **45**:561-572.
32
33
34 163 Hillis, D. M., and J. J. Bull. 1993. An empirical test of bootstrapping as a method for
35
36 164 assessing confidence in phylogenetic analysis. *Systematic Biology* **42**(2):182.
37
38
39 165 Lloyd, H. G. 1962. The distribution of squirrels in England and Wales, 1959. *Journal of*
40
41 166 *Animal Ecology* **31**:157-166.
42
43
44 167 Lloyd, H. G. 1983. Past and present distribution of red and grey squirrels. *Mammal Review*
45
46 168 **13**(2-4):69-80.
47
48
49 169 Lowe, V. P. W. 1993. The spread of the grey squirrel (*Sciurus carolinensis*) into Cumbria
50
51 170 since 1960 and its present distribution. *Journal of Zoology* **231**:663-667.
52
53 171 Meece, J. K., C. E. Reynolds, P. J. Stockwell, T. A. Jenson, J. E. Christensen, and K. D. Reed.
54
55 172 2005. Identification of mosquito bloodmeal source by terminal restriction fragment length
56
57 173 polymorphism profile analysis of the cytochrome B gene. *Journal of Medical Entomology*
58
59 174 **42**(4):657-667.
60

- 1
2
3 175 Middleton, A. D. 1931. The grey squirrel. The introduction and spread of the American grey
4
5 176 squirrel in the British Isles, its habits, food, and relations with the native fauna of the country.
6
7
8 177 Sidgwick and Jackson, London.
9
10 178 Sainsbury, A. W., R. Deaville, B. Lawson, W. A. Cooley, S. S. J. Farely, M. J. Stack, P.
11
12 179 Duff, C. J. McInnes, J. Gurnell, P. H. Russell, S. P. Rushton, D. U. Pfeiffer, P. Nettleton, and
13
14 180 P. W. W. Lurz. 2008. Poxviral disease in red squirrels *Sciurus vulgaris* in the UK: Spatial and
15
16 181 temporal trends of an emerging threat. *EcoHealth* 5(3):305-316.
17
18 182 Shorten, M. 1954. Squirrels. Collins, London.
19
20 183 Skelcher, G. 1997. The ecological replacement of red by grey squirrels. Pages 67-78 in J.
21
22 184 Gurnell and P. Lurz, editors. The conservation of red squirrels, *Sciurus vulgaris* L. People's
23
24 185 Trust for Endangered Species, London.
25
26 186 Stevenson, C. D., Watts, K., Nevin, O.T., Ramsey, A. D. and S. Bailey. In review. Validation
27
28 187 and creation of a 'best fit' resistance set within a spatially explicit behaviourally-informed
29
30 188 landscape model using historical observations of species invasion and information theory.
31
32 189 Tompkins, D. M., A. R. White, and M. Boots. 2003. Ecological replacement of native red
33
34 190 squirrels by invasive greys driven by disease. *Ecology Letters* 6(3):189-196.
35
36 191 Trizio, I., B. Crestanello, P. Galbusera, L. A. Wauters, G. Tosi, E. Matthysen, N., and H. C.
37
38 192 Hauffe. 2005. Geographical distance and physical barriers shape the genetic structure of
39
40 193 Eurasian red squirrels (*Sciurus vulgaris*) in the Italian Alps. *Molecular ecology* 14(2):469-
41
42 194 481.
43
44 195 Wauters, L. A., P. W. W. Lurz, and J. Gurnell. 2000. Interspecific effects of grey squirrels
45
46 196 (*Sciurus carolinensis*) on the space use and population demography of red squirrels (*Sciurus*
47
48 197 *vulgaris*) in conifer plantations. *Ecological Research* 15(3):271-284.
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



©Crown Copyright/database right 2010. An Ordnance Survey/EDINA supplied service

Figure 1. Geographical location of grey squirrel sample sites within the UK.

186x273mm (300 x 300 DPI)

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

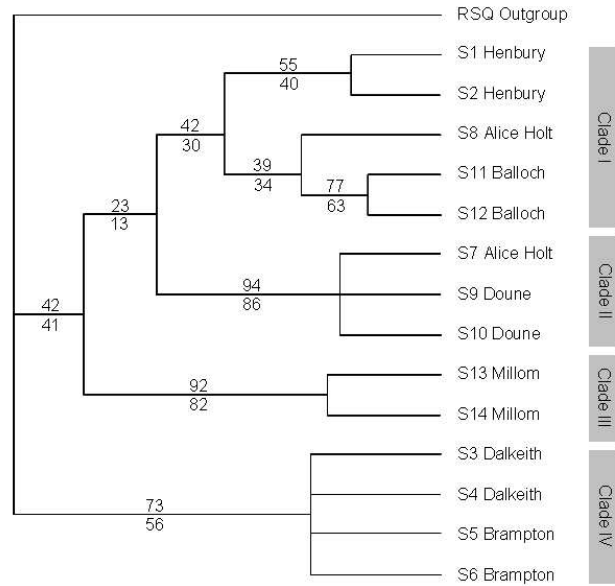


Figure 2. Phylogenetic tree showing Bootstrap and Jackknife analysis results above and below each group separation.
254x190mm (96 x 96 DPI)