

Barnes, P.W., Robson, Matthew ORCID: <https://orcid.org/0000-0002-8631-796X> , Zepp, R.G., Bornman, J.F., Jansen, M.A.K., Ossola, R., Wang, Q-W., Robinson, S.A., Foereid, B., Klekociuk, A.R., Martinez-Abaigar, J., Hou, W-C., Mackenzie, R. and Paul, N.D. (2023) Interactive effects of changes in UV radiation and climate on terrestrial ecosystems, biogeochemical cycles, and feedbacks to the climate system. *Photochemical & Photobiological Sciences*, 22 (5). pp. 1049-1091.

Downloaded from: <http://insight.cumbria.ac.uk/id/eprint/6879/>

***Usage of any items from the University of Cumbria's institutional repository 'Insight' must conform to the following fair usage guidelines.***

Any item and its associated metadata held in the University of Cumbria's institutional repository Insight (unless stated otherwise on the metadata record) may be copied, displayed or performed, and stored in line with the JISC fair dealing guidelines (available [here](#)) for educational and not-for-profit activities

**provided that**

- the authors, title and full bibliographic details of the item are cited clearly when any part of the work is referred to verbally or in the written form
- a hyperlink/URL to the original Insight record of that item is included in any citations of the work
- the content is not changed in any way
- all files required for usage of the item are kept together with the main item file.

**You may not**

- sell any part of an item
- refer to any part of an item without citation
- amend any item or contextualise it in a way that will impugn the creator's reputation
- remove or alter the copyright statement on an item.

The full policy can be found [here](#).

Alternatively contact the University of Cumbria Repository Editor by emailing [insight@cumbria.ac.uk](mailto:insight@cumbria.ac.uk).



# Interactive effects of changes in UV radiation and climate on terrestrial ecosystems, biogeochemical cycles, and feedbacks to the climate system

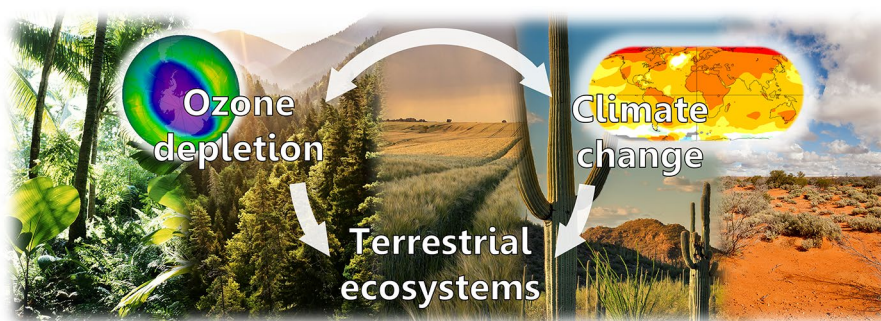
P. W. Barnes<sup>1</sup> · T. M. Robson<sup>2,3</sup> · R. G. Zepp<sup>4</sup> · J. F. Bornman<sup>5</sup> · M. A. K. Jansen<sup>6</sup> · R. Ossola<sup>7</sup> · Q.-W. Wang<sup>8</sup> · S. A. Robinson<sup>9</sup> · B. Foereid<sup>10</sup> · A. R. Klekociuk<sup>11</sup> · J. Martinez-Abaigar<sup>12</sup> · W.-C. Hou<sup>13</sup> · R. Mackenzie<sup>14,15</sup> · N. D. Paul<sup>16</sup>

Received: 22 December 2022 / Accepted: 13 January 2023  
© The Author(s) 2023, corrected publication 2023

## Abstract

Terrestrial organisms and ecosystems are being exposed to new and rapidly changing combinations of solar UV radiation and other environmental factors because of ongoing changes in stratospheric ozone and climate. In this Quadrennial Assessment, we examine the interactive effects of changes in stratospheric ozone, UV radiation and climate on terrestrial ecosystems and biogeochemical cycles in the context of the Montreal Protocol. We specifically assess effects on terrestrial organisms, agriculture and food supply, biodiversity, ecosystem services and feedbacks to the climate system. Emphasis is placed on the role of extreme climate events in altering the exposure to UV radiation of organisms and ecosystems and the potential effects on biodiversity. We also address the responses of plants to increased temporal variability in solar UV radiation, the interactive effects of UV radiation and other climate change factors (e.g. drought, temperature) on crops, and the role of UV radiation in driving the breakdown of organic matter from dead plant material (i.e. litter) and biocides (pesticides and herbicides). Our assessment indicates that UV radiation and climate interact in various ways to affect the structure and function of terrestrial ecosystems, and that by protecting the ozone layer, the Montreal Protocol continues to play a vital role in maintaining healthy, diverse ecosystems on land that sustain life on Earth. Furthermore, the Montreal Protocol and its Kigali Amendment are mitigating some of the negative environmental consequences of climate change by limiting the emissions of greenhouse gases and protecting the carbon sequestration potential of vegetation and the terrestrial carbon pool.

## Graphical abstract



This Perspective is part of the topical collection: Environmental effects of stratospheric ozone depletion, UV radiation, and interactions with climate change: UNEP Environmental Effects Assessment Panel, 2022 Quadrennial Assessment.

Extended author information available on the last page of the article

## 1 Introduction

The Montreal Protocol and its Amendments have been highly effective in protecting the Earth's stratospheric ozone layer and preventing global-scale increases in solar

ultraviolet-B radiation (UV-B; wavelengths between 280 and 315 nm) at the Earth's surface [1]. Consequently, this multilateral treaty, ratified by all 198 United Nations member states, has prevented large-scale detrimental effects of elevated UV-B radiation on agricultural productivity, terrestrial organisms and ecosystems [2–4]. Moreover, because many of the ozone-depleting compounds regulated by the Montreal Protocol are also potent greenhouse gases, this treaty and its Kigali Amendment are playing an important role in mitigating global warming and other environmental effects of climate change [5, 6].

Changes in stratospheric ozone and climate are not independent of one another [7, 8] and both can affect surface ultraviolet radiation (UV; 280–400 nm), especially UV-B radiation [9–11]. According to current projections, which assume full compliance with the Montreal Protocol, future changes in UV radiation reaching the Earth's surface are likely to be due primarily to changes in climate (i.e. mainly cloud cover, aerosols and surface reflectivity) rather than changes in stratospheric ozone [10, 12, 13]. However, future changes in UV radiation at the Earth's surface are uncertain: a new study projects an increase in the UV Index of 3–8% over the tropics and mid-latitudes, respectively, by 2100 depending on the greenhouse gas (GHG) scenario used in the model simulations, cloud cover, and aerosol concentrations [10, 14]. Changes in the exposure of organisms and ecosystems to UV radiation also results from increased incidence and extent of wildfires, which generate aerosols (also causing further damage to the ozone layer), and from alterations in vegetation cover from land-use practices (e.g. deforestation), melting of snow and ice, and shifting distribution ranges of species responding to climate change [10, 12, 15–18] (Summarised in Table 1). In this assessment, we address how the expected, rather small changes in UV irradiation interact with the ongoing changes in climate to affect food security, biodiversity, biogeochemical cycles and feedbacks to the climate system.

Since our last Quadrennial Assessment [12, 19], the Earth's climate has continued to change and the frequency and intensity of extreme climate events (e.g. heat waves, droughts, and storms), and those events resulting from a combination of weather extremes and other drivers (e.g. wildfires), have increased [20, 21]. As global warming and its consequences continue to increase, there is renewed interest in possible technological interventions to reduce the warming. Stratospheric Aerosol Injection (SAI), an intervention that involves Solar Radiation Management (SRM), has received the most attention due to its potential feasibility. SAI would involve injecting reflective aerosols, such as sulphate, into the stratosphere to reflect incoming solar radiation away from the Earth's surface [22]. There are many uncertainties associated with this intervention, including risks to the stratospheric ozone layer that could increase ground-level UV irradiance [23–25]. In addition to the risks associated with the initiation of SAI, once adopted,

**Table 1** Summary of the effects of various climate change-driven factors on the potential exposure of terrestrial plants and animals to UV radiation

| Climate change effect                         | Effect on exposure to UV radiation |
|---|------------------------------------|
| Migration or range shift to higher elevations | +                                  |
| Migration or range shift to higher latitudes  | - -                                |
| Altered phenology (seasonal development)      | -/+                                |
| Deforestation (wet regions)                   | ++ +                               |
| Shrub encroachment (dry regions)              | - - -                              |
| Altered cloud cover                           | - - -/+ + +                        |
| Change in aerosols                            | - -/+ + +                          |
| Decreased snow/ice cover                      | - /+ +                             |

Effects show direction (i.e. decreases (–) or increases (+)) in exposure to UV radiation with the relative magnitude of these changes indicated by the number of negative and positive signs. In some cases (e.g. altered phenology), changes may either increase or decrease UV exposure depending on the circumstances and species. Changes in exposure to UV radiation resulting from modifications in land cover (i.e. deforestation and shrub encroachment) refer to effects on ground-dwelling, understory organisms. The effects on exposure to UV radiation shown here do not include changes in stratospheric ozone. Additional information and relevant references are provided in the text that follows

any subsequent termination of this climate intervention would lead to a rapid increase in temperature and extreme deleterious effects on ecosystems [26, 27]. This, and other SRM interventions, would likely expose the Earth's ecosystems to new and potentially rapidly changing combinations of UV radiation and other biotic and abiotic environmental factors [28].

In this Quadrennial Assessment, we evaluate the current state of the science on the changes in stratospheric ozone, solar UV radiation and their interactions with climate change as they affect terrestrial ecosystems and biogeochemical cycles in the context of the Montreal Protocol [29]. We also address key gaps in knowledge and how these interacting effects and the Montreal Protocol will have a bearing on the targets of the United Nations Sustainable Development Goals (SDGs) and their targets.

## 2 Effects of stratospheric ozone depletion on climate and extreme climate events on exposure to UV radiation

While both stratospheric ozone depletion and climate change can modify the amount of UV radiation reaching terrestrial ecosystems [8, 10], ozone depletion itself can also contribute to climate change by modifying atmospheric circulation patterns and altering regional patterns of wind, precipitation and temperature [30–32]. The impacts of these changes in climate on terrestrial ecosystems have been most

pronounced in Antarctica and in the high latitudes of the Southern Hemisphere, although there is evidence of ozone-driven climate change in Arctic regions as well [33]. In addition to the effects of climate change on UV irradiation outlined above (Table 1), extreme events linked to climate change (e.g. droughts, floods, heat waves, fires) may abruptly change UV radiation conditions for many organisms. Below, we assess recent findings on the effects of ozone-driven climate change on polar ecosystems and the potential effects of extreme events on the exposure of terrestrial ecosystems in general to UV radiation.

## 2.1 Recent stratospheric ozone depletion and climate change effects on polar ecosystems

The impact of stratospheric ozone depletion on polar ecosystems is a complex interplay between the consequences of changing surface UV radiation, and effects caused by shifts in the weather and climate due to the associated cooling of the lower stratosphere [12, 29, 34]. The increased UV irradiance in the polar regions as a direct result of ozone depletion has been documented since the late 1970s (ozone hole era) [8]. This has particularly been the case in the Antarctic region, where measurements show that the UV Index at the surface in late spring and early summer has, at times, been similar to that at mid- and subtropical latitudes [10, 35]. In the past, it was assumed that snow and ice cover would provide plants and surface organisms some protection from the high UV irradiances that occur during the peak of ozone depletion, but with climate warming accelerating the melting of snow and sea ice, Antarctic organisms are increasingly being exposed to this elevated UV radiation. How these high UV irradiances in late springtime impact the resident plants and animals is not entirely clear; studies conducted at the end of the twentieth century found relatively small effects on plants exposed to the elevated UV radiation experienced at that time. This was likely due to the inherent adaptations, UV-protective mechanisms and acclimation responses of these species in order to survive extreme environments [36–38]. Without the Montreal Protocol, the maximum UV Index would have potentially increased from pre-ozone depletion levels of 6–20, exposing coastal Antarctic organisms to UV Indices at the end of this century that would be greater than those experienced today in the tropics [10]. These extreme UV radiation conditions would likely have exceeded the UV-tolerances of many Antarctic organisms. In the Arctic, surface UV-B irradiance has also been elevated in recent years (e.g. 2019/2020) when episodic large stratospheric ozone depletion has followed anomalously cold stratospheric winters [10]. However, unlike in the Antarctic, these events occur during early spring when most organisms are still protected by sea ice or snow cover.

Changes in the stratosphere driven by ozone depletion have also been clearly shown to cause seasonally dependent shifts in near-surface patterns of wind, temperature and precipitation [39–42]. Knock-on effects on warming of oceans and melting sea ice cover have been investigated [40–49], but many uncertainties persist [50] as the effects of ozone depletion on weather patterns are occurring against a backdrop of climate change. Collectively, these changes have led to increased variability of weather and climate, which is most pronounced in the polar regions [51]. As documented in our previous assessments and elsewhere, these shifts in weather and climate have had pronounced impacts on many Antarctic organisms, from tiny moss and cushion plants to wandering albatross [12, 34, 52, 53].

Since our last Quadrennial Assessment, extremes have occurred in both ozone depletion and climatic events that have led to observed or potential effects on plants and animals in polar regions (Table 2). Specific findings include:

- During spring 2019, the Antarctic stratosphere was strongly disturbed by meteorological influences from upward-propagating atmospheric waves [54–56] resulting in a small ozone hole. These stratospheric conditions played a role in enhancing prolonged drought over the 2019/2020 austral summer that exacerbated the unprecedented wildfires in eastern Australia [57–64]. Effects on stratospheric chemistry following the wildfires led to wider changes in both the chemical composition and temperature of the stratosphere across southern mid-latitudes [18, 65–73]. Strong vertical and horizontal gradients in the ozone concentration of the Antarctic upper troposphere during the austral spring potentially delayed the subsequent effects on surface climate [32]. The role of ozone depletion in modulating the dynamical coupling between the polar stratosphere and the surface at lower latitudes for this particular season is still under investigation. Nevertheless, it appears likely that the combined effects of climate change and ozone depletion could have impacted both the timing and magnitude of these wildfires with considerable consequences for ecosystems in this region.
- In contrast to 2019, a strong and persistent Antarctic ozone hole occurred in 2020 and 2021 [54, 74–76] and this led to record surface UV irradiances at several sites across East Antarctica during early summer. It has been suggested that the Australian wildfires that occurred during the previous summer contributed to this strong ozone loss [18, 54–56, 71, 72, 74]. There is evidence that increased ozone depletion has tended to delay the annual breakdown of the Antarctic stratospheric vortex [77]. Modelling suggests that increasing greenhouse gas concentrations also favour a more persistent vortex [78], as well as reducing the likelihood of a weaker vortex

**Table 2** Summary of environmental effects of stratospheric ozone changes and concurrent climate extremes from 2018 to 2022.

| Ozone effects   | Climate extremes and associated effects   | Plant responses  | Animal responses, including humans   |
|---|---|--|--|
| <b>Antarctica and Southern Hemisphere (September 2019–February 2020)</b>            |   |  |  |
| Anomalously small ozone hole  | Wildfires in Australia produced aerosols that caused ozone depletion and black carbon particles that accelerated snow melt [76] | Widespread loss of plant biomass in Australia [97]   | Loss of human life and adverse health effects; loss of domestic animals and wildlife [98–101]  |
|   | Heatwaves in Antarctica [102]   | Additional snow melt that caused temporary greening of some previously moribund moss beds in East Antarctica [53, 103]<br>Vascular plants on the Antarctic Peninsula appear to be faring better than mosses under global warming and the grass <i>Deschampsia antarctica</i> appears to be quite tolerant of <i>in-vitro</i> high temperature shock treatments [104]<br>Extreme summer marine heatwaves increased chlorophyll <i>a</i> (an indicator of the abundance of phytoplankton) in the Southern Ocean [105]<br>Hotter and longer heatwaves increased the mortality and decreased post-heatwave growth rates in the Southern Ocean diatom <i>Actinocyclus actinocylus</i> relative to milder, shorter heatwaves [106] | Functional thermal limits for the Antarctic sea urchin ( <i>Sterechinus neumayeri</i> ) were determined under simulated marine heatwaves.<br>Key biological functions vary in their thermal sensitivity and in their responses to different rates of warming [107] |
| <b>Arctic (January – April 2020)</b>  |   |  |  |
| Anomalously large ozone depletion [8]   | Heatwave in the Siberian Arctic; accelerated loss of sea ice [96]   | Permafrost warming and thaw lead to landscape changes (retrogressive thaw slumps) and increased greenhouse gas emissions [108]   | Negative impacts on organisms that depend on sea ice; positive impacts on animals that thrive in open oceans [95, 109, 110]  |
| <b>Antarctica (November – December 2020 and November – December 2021)</b>           |   |  |  |
| Persistent ozone hole producing anomalous surface UV irradiance [76]. See also [10] |   | High potential for excessive exposure to UV radiation as plants emerge from under winter snow [82]. Earlier snow melt, meaning more exposure, which coincided with extreme maximum UV Index  | High potential for excessive exposure to UV radiation as animals return to breeding sites in spring and early summer [111]. Reductions in Antarctic sea ice [21] will result in higher exposure to UV radiation in the water column                                |

The factors that affect the size of the Antarctic and Arctic ozone holes each spring bring widespread climate impacts that can extend far beyond the polar regions. For example, Antarctic ozone depletion varies with the phase of the southern annular mode (SAM). The SAM phase has been linked to the black summer (2019/2020) bushfires in Australia, which produced aerosols that contributed to further ozone depletion and smoke particles, which accelerated snow melt in New Zealand and Antarctica. Note that the assessment of the environmental impacts of heatwaves and anomalous ozone dynamics has been extremely limited due to the COVID-19 pandemic that prevented many planned scientific visits to remote polar regions



[58]. While concentrations of ozone-depleting substances (ODSs) remain elevated, later seasonal persistence of the Antarctic vortex could expose organisms to higher UV irradiances at times of year when young animals are born/hatch and when plants are actively growing. The loss of protective snow cover could exacerbate these effects [34].

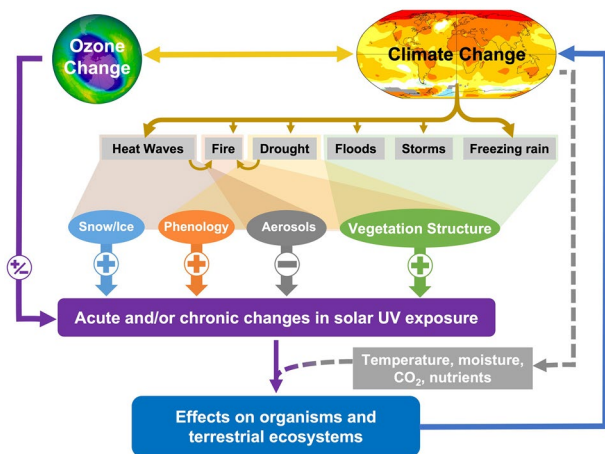
- Since our last assessment there have been two widespread heatwave events in Antarctica, the first in summer 2019/2020 when heat records were broken around the continent [63]. In March 2022 (autumn) extreme temperatures, almost 40 °C higher than normal, were reported as an atmospheric river, or plume of warm, moist air, moved onto the Antarctic plateau. Heatwaves such as these accelerate melting of icebanks [79], potentially exposing vegetation to high springtime UV-B radiation from which they have previously been protected [36]. The impacts of these heatwaves and the subsequent ice melt have been poorly studied in Antarctica in part due to the lack of environmental monitoring with networks of sensors tracking temperature and climate variables at appropriate scales. This lack of data is well illustrated by the recently published global maps of soil temperature [80], which exclude Antarctica. Warming temperatures on the Antarctic Peninsula are opening up ice free areas [79] causing the expansion of vascular plants [81] and increasing the possibility of new plant and animal species invading the continent [21, 82]. As in the Arctic, there are examples of both plant expansion (i.e. “greening”, [81, 83]) and death of plants by drought (i.e. “browning”, [84–88]). Heatwaves may also be particularly detrimental to mosses, as they survive by creating warm microclimates in Antarctica’s cold environments but this may become a disadvantage as air temperatures increase [89].
- In the Arctic, unprecedented low total column ozone values occurred in the 2020 boreal spring [90, 91] due to strong stratospheric ozone depletion, and this resulted in record-breaking high solar UV-B irradiances [92, 93]. These conditions were promoted by weak tropospheric wave activity [90, 94], associated with anomalous sea surface temperature in the North Pacific [33], which caused the stratospheric vortex to become large and stable. Heatwave conditions that occurred in the Siberian Arctic in early 2020 [95] appear to have been aided by atmospheric circulation patterns that were affected by the strong ozone depletion [33, 94]. Ozone depletion in March 2020 may also have aided the prevailing reduction of sea ice in the Arctic Ocean bordering Siberia [96]. As indicated above, most Arctic organisms are currently protected by snow and sea ice at the time of maximum ozone depletion and high UV radiation conditions at this time of year (i.e. early March to mid-April 2020), but

changes in snow and ice cover resulting from climate change could increase exposure to UV radiation.

## 2.2 Interactive effects of extreme climate events and UV radiation extending beyond polar ecosystems

Globally, extreme climate events (ECEs<sup>1</sup>) are increasing in frequency and severity with climate change and are projected to become even more prevalent in the future as the climate continues to change [20]. Examples of ECEs include stronger storms and tropical cyclones, catastrophic floods, protracted droughts, anomalous heat waves and freezes, and more intense wildfires [113–118]. ECEs cause long-term disruption to ecosystem structure and function [119–122] and occur against a backdrop of more gradual changes in the environment (e.g. rising surface temperatures and atmospheric carbon dioxide (CO<sub>2</sub>) concentrations). These disruptions to ecosystem function can exacerbate the deleterious effects of ECEs on plants and animals [123]. Extreme climate events also alter the amount of UV radiation reaching terrestrial ecosystems (Fig. 1). These changes in UV radiation can occur over short or long timeframes, which can then lead to acute or chronic effects on ecosystems, respectively. The changes in solar UV radiation together with other environmental factors (e.g. temperature, availability of moisture) may affect biodiversity, productivity, emissions of greenhouse gases [124–126], and ecosystem carbon storage [123]. For example, fires, floods, and tropical cyclones (hurricanes) all create openings in forest canopies [127, 128], driving subsequent adjustment in the understorey vegetation to an acute or chronic increase in incident solar radiation; these increases in solar radiation are often accompanied by increases in temperature and decreases in soil moisture [129–131]. There is also an associated increase in the amplitude of fluctuations in these abiotic factors. Some plant species (e.g. shade-adapted specialists) may not be able to adjust to this new environment and will go locally extinct. However, other plant species can respond quickly to these environmental changes [132–134] and may increase in

<sup>1</sup> An extreme climate event has been defined as “an episode or occurrence in which a statistically rare or unusual climatic period alters ecosystem structure and/or function well outside the bounds of what is considered typical or normal variability” [112]; or similarly, according to the IPCC, “if the value of a variable exceeds (or lies below) a threshold” that is exceeded [20]. Compound extreme events are the “combination of multiple drivers and/or hazards that contribute to societal or environmental risk.” An example of a compound extreme event would be fire weather conditions which are the combination of hot, dry, and windy conditions [20].



**Fig. 1** Pathways by which extreme climate events (ECEs) driven by changes in stratospheric ozone and climate can affect exposure of terrestrial organisms and ecosystems to UV radiation. Changes in stratospheric ozone and climate interact to influence the frequency and intensity of a number of ECEs (upper-most grey rectangles). These ECEs in turn affect atmospheric and surface intermediaries (multi-coloured ovals connected with ECEs by overlapping shaded regions), which can increase (+) or decrease (-) the solar UV radiation reaching terrestrial organisms and ecosystems. Solid arrows show direct mediation by climate, ozone and UV radiation on ECEs and potential interactive and feedback effects. Dashed arrows show chronic effects of climate change factors

abundance. With respect to UV radiation, some plant species can respond rapidly to increases in amounts and variability in solar UV radiation through the production and accumulation of UV-protective pigments [131, 135] (Sect. 3), and these attributes may allow these species to be successful in the changing conditions. From an ecosystem perspective, fires and hurricanes are among the most disruptive examples of ECEs as they can cause the loss of productivity and biodiversity, and increase the emissions of GHGs [136–138], which can be enhanced by UV radiation (Sect. 6).

The disruptive nature of ECEs also opens up the remaining ecological communities to invasive species, which can further destabilise these systems [12]. For example, certain invasive species that can tolerate high solar radiation and colonise open habitats may displace some native, specialised or endemic species [139]. To what extent differences among plant species in their tolerances to UV radiation influences species invasions into high UV environments remains unclear [140, 141]. Recovery of ecosystems from these ECEs will largely depend on the species that colonise the more open habitats created, and their biodiversity value and traits that support ecosystem function [142].

Wildfires and droughts affect the amount of UV and photosynthetically active radiation (PAR; 400–700 nm) reaching terrestrial ecosystems due to increasing aerosols from smoke and dust, and volatile organic compounds released by plants [143]. These atmospheric changes not only reduce PAR and

UV radiation, but also change the spectral composition of sunlight at ground level [144]. Importantly, changes in air quality resulting from fires and droughts can occur well beyond the location of these events [145–147]. Thus, these conditions arising from fires and droughts can potentially affect photosynthesis and light-driven development in plants [148], as well as litter decomposition and GHG emissions in ecosystems [19] not directly impacted by these extreme events (Sect. 6.1).

### 3 Effects of UV radiation and climate interactions on plants and animals

While moderate UV-B irradiance serves as an informational cue that facilitates the normal regulation of plant growth and metabolism, exposure to excessive UV radiation, and in particular short-wavelength UV-B radiation, can have deleterious effects on terrestrial organisms [e.g. 150]. As sessile, photosynthetic organisms, plants require sunlight for their growth and reproduction, but this also means that they can receive a large cumulative amount of solar UV radiation over their lifetime. This cumulative amount would have been very high in the extreme UV irradiation conditions that would have occurred without the Montreal Protocol [1]; however, because of its implementation appreciable reductions in photosynthesis in terrestrial plants have been avoided. High UV irradiance conditions would also likely impair growth with severe consequences for global carbon storage and climate [3, 10] (Box 1). Under current climate conditions, and in most regions of the world, land plants appear to show adequate protection against UV radiation that limits the deleterious effects of moderate UV-B radiation. There are physiological similarities in responses to adverse conditions (stress), that may determine the extent to which plants can tolerate increased UV radiation in combination with other abiotic factors (e.g. temperature, drought, elevated CO<sub>2</sub>) that occur simultaneously. In this section, we highlight recent progress in identifying the mechanisms by which plants perceive and respond to UV radiation. These findings allow us to better assess the impacts of changes in UV radiation, plant response to rapid increases in UV radiation (as occur following many ECEs), and how UV radiation interacts with environmental stresses (e.g. climate change) to modulate their growth and productivity.

In contrast to the abundant literature on the effects of UV-B radiation on terrestrial plants, far less attention has been paid to the effects of UV-B radiation on terrestrial animals. What research there is on animals, typically addresses vision in the UV-A waveband and its effect on behaviour [150], and the application of these findings for controlling insect pests and pollinators of certain crops [151]. One exception is the increasing research, largely focussed on

agricultural systems, showing that terrestrial invertebrates, including mites [152, 153] and insects such as aphids [154–156], are vulnerable to direct damage from UV-B radiation. There are interesting parallels between invertebrate and plant responses to UV radiation; for example, in the role of DNA-repair [157, 158], antioxidant metabolism [159–162] and pigments [155, 156, 161] in conferring UV-protection. This includes evidence that mites can obtain UV-protective compounds by consuming pollen [160]. It is also clear that avoidance behaviour plays a major part in reducing the exposure of invertebrates to solar UV radiation [152–154, 163].

### 3.1 Perception and response of plants to changing UV radiation

The need to better understand how organisms respond to elevated UV-B radiation, as occurs with stratospheric ozone depletion, stimulated research that eventually led to the discovery of a UV-B photoreceptor in plants (UVR8, which stands for Ultraviolet Resistance Locus 8) [164]. It is now well-documented that UVR8 mediates a number of plant responses to changes in UV-B radiation in the environment. Recently, UVR8 has been found to operate over a spectral region extending through the UV-B and part of the UV-A radiation wavebands [165]. Thus, variation in solar UV radiation attenuated by the stratospheric ozone layer (which screens UV radiation up to *ca.* 335 nm) is well matched to the action spectrum of UVR8 [166, 167]. This might suggest that the evolution of UVR8 allowed plants to perceive and respond to environmental cues related to changes in stratospheric ozone.

The UVR8-signalling pathway likely evolved very early in the transition of plants from aquatic to terrestrial environments [168–170]. Two overlapping signalling pathways for UV responses (UVR8/WRKY36/HY5 and UVR8/COP1/SPA-HY5 pathways) have been conserved during the evolution of green plants [170, 171]. These pathways regulate a series of genetic transcription factors that affect accumulation of flavonoids, functioning of the plant hormone auxin, and growth (i.e. through inhibition of elongation of lateral roots and hypocotyls [172]). Subsequently, diversification of signal transduction to increase crosstalk with other signalling pathways that control the production of additional secondary metabolites, such as brassinosteroids (hormones involved in plant development), enabled fine-tuning of tolerance to UV radiation in photosynthetic organisms. Specific responses in plants that are involved in their acclimation to UV radiation include: the accumulation of flavonoid pigments as UV sunscreens, shorter stature with increased branching, and smaller leaves with thickened cell walls. These changes together with a more conservative strategy (i.e. slower but more efficient growth, photosynthesis, and

water loss [173, 174]) collectively mitigate the potentially deleterious effects of current levels of solar UV radiation on plants.

Among the diverse functions of phenolic compounds in growth, development and reproduction, certain flavonoids and related phenolic acids (e.g. hydroxycinnamic acid derivatives) screen UV radiation in plant tissues and are therefore central to plant UV-acclimation responses. The accumulation of these compounds in leaves, flower petals and pollen is temperature dependent but is also driven by UV-B radiation [135, 175, 176]. Flavonoids fulfil many additional roles in plants, in that they are involved in ameliorating biotic and abiotic environmental-stress, regulating the transport of certain hormones (i.e. auxin) and are required in many species for successful germination and growth of the pollen tube on the stigma of flowers, where they participate in cell signalling and recognition [177–180]. There is also evidence that greater accumulation of flavonoids in pollen grains improves their germination (e.g. in *Clarkia unguiculata*; [181]) and flavonoids function in UV screening in pollen, which is essential to maintain viability [182, 183]. Additionally, flavonoid glycosides (quercetins and kaempferols), hydroxycinnamic acids and anthocyanins in leaves and pollen act as strong antioxidants, and, as such, they scavenge reactive oxygen species (ROS) produced by abiotic stressors such as excessive solar radiation, including UV-B radiation [178, 184].

In assessing plant acclimation to increased UV radiation, it is relevant to consider responses to short-term, rapid fluctuations in UV radiation—as would occur with changing cloud cover or from day-to-day during the break-up of the stratospheric ozone hole—as well as to the longer-term (i.e. decade-scale changes that occur from anthropogenic changes in stratospheric ozone together with climate). The patterns of these responses can be used to evaluate whether plants' epidermal UV screening and photoprotection principally acclimate to immediate changes in UV radiation or if plants mainly rely on other mechanisms that allow trans-generational improvements in protection against UV radiation (i.e. genetic adaptation or epigenetics). There is increasing evidence that the accumulation of photoprotective compounds (including flavonoids, hydroxycinnamic acids and carotenoids) tracks seasonal and even daily variation in UV radiation [185–190]. In general, the magnitude of diurnal changes in UV screening is less than those that occur during the development of leaves. Diurnal changes in UV screening can, however, be of comparable size to the variation in screening that results from day-to-day fluctuations in UV radiation and temperature [176, 186, 191]. Rapid acclimation of UV screening to short-term changes in UV radiation indicates a high level of phenotypic plasticity and suggests that many plants can acclimate to short-term fluctuations in UV irradiance arising from transient reductions



in stratospheric ozone, reduced cloud cover or certain ECEs (Fig. 1; Sect. 2.2). In fact, a comparison of 629 taxa growing together at high-elevation and high-latitude locations subject to strongly contrasting UV irradiances, found phenotypic plasticity in epidermal UV screening according to their immediate growing microenvironment, and this outweighed any differences in adaptation arising from their evolutionary history under disparate climates [192]. Similarly, the importance of the local environment over the place of origin is also highlighted by experiments where species and populations are grown in the same location and habitat (i.e. common-garden experiments [193]).

Although the capacity for rapid acclimation may be advantageous for adjusting to short-term environmental variability, high phenotypic plasticity may interfere with the capacity for genetic adaptation to changing conditions over long time periods [194–196]. Understanding the relative importance of phenotypic plasticity *vs.* genetic adaptation is needed to evaluate the consequences of climate change-induced range shifts that expose plant species to UV irradiances that might be beyond those experienced in their historic ranges (Sect. 4.1).

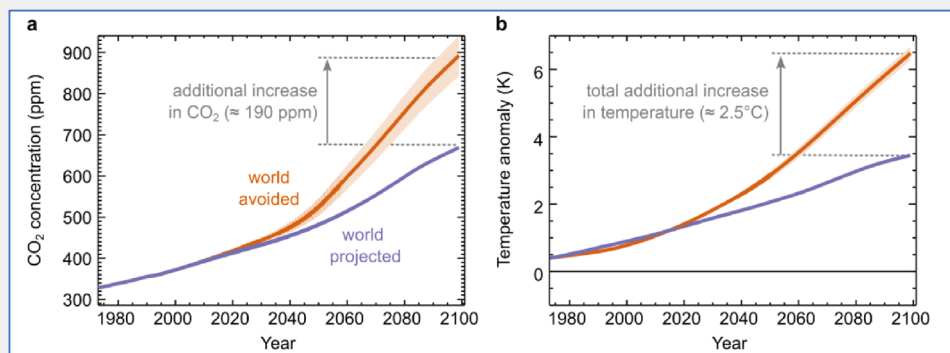
### Box 1 The effect of terrestrial ecosystems on climate change in a world without successful controls of ozone-depleting substances.

A recent modelling exercise [3] estimated that large increases in plant effective UV-B radiation resulting from uncontrolled emissions of ozone-depleting substances (ODS) would have strongly reduced carbon dioxide uptake by terrestrial vegetation in the “world avoided” (i.e., a world without effective control of ODS by the Montreal Protocol) compared with the “world projected” (i.e., the actual world with the Montreal Protocol projected into the future). As a result of the control of ODS emissions, by 2100 it was estimated that:

- Carbon storage by terrestrial vegetation would have decreased by 325–690 billion tons;
- Atmospheric CO<sub>2</sub> concentrations would have increased by an additional 190 ppm, on average (range = 115–235 ppm; panel (a) in the figure below); and
- Global mean surface temperature would have risen an additional 2.5 °C (range = 2.4–2.7 °C; panel (b), which includes the ODS warming effect (1.7 °C) and the UV plant effect (0.85 °C).

Because of the relatively long time period involved, there are significant uncertainties associated with these projections. The size of the effect depends on the action spectrum used to describe the sensitivity of photosynthesis to different wavelengths of UV-B radiation and on extrapolating the dose response relationship between change in plant effective UV-B radiation and primary productivity (see Sect. 8 and [9]).

Nonetheless, this modelling highlights both the potential damage to terrestrial ecosystems in the world avoided scenario and the limits imposed on such models by the scope of known responses of plants and mixed vegetation to changes in UV-B radiation.



Changes in atmospheric CO<sub>2</sub> concentrations (a) and surface temperature (b) resulting from UV-B radiation under scenarios with (violet line) and without (orange line) the Montreal Protocol. Shading around the orange line represents the range of responses from simulations assuming a 50–150% range of plant responses to UV radiation. Figure adapted from [3], reproduced by permission.

Studies examining the mechanisms by which native plant species tolerate naturally high UV-B environments can provide insights into the range of adaptive responses exhibited by plants to UV-B radiation. For example, *Rheum nobile* (Sikkim Rhubarb), an herbaceous plant that grows above 4000 m on the Tibetan Plateau, has large translucent flower bracts containing high concentrations of flavonoids, which form a protective cover over its flowers. This species can, thus, attenuate UV-B radiation within its floral tissues to similar levels across its elevation range [197]. In the same region, other herbaceous plants, such as *Megacarpaea delavayi* (a wild mustard [198]) and five species in the genus *Saussurea* (thistle-like plants in the sunflower family [199]) have a rapid DNA-repair mechanism to mitigate the damaging effects of high UV-B irradiances. Comparative genomic analysis of 377 Tibetan peach populations showed that the expansion of SINE retrotransposons (genetic variations that regulate gene expression), promotes adaptation to UV-B radiation [200]. These, and other evolutionary adaptations of specialist alpine species to extreme UV radiation conditions indicate how plants in general might adapt to high UV-B irradiances. However, the rate of changes in UV irradiance as a result of ozone depletion or climate change is likely to outpace the rates of adaptation in many species, especially long-lived perennials such as trees. Additionally, plants endemic to high elevations often have limited distribution ranges and abundances, and may be among the most vulnerable to habitat loss due to climate change.

### 3.2 Proxies for past solar UV irradiance based on acclimation responses of modern-day plants to UV radiation

Because many of the phenolic UV sunscreens accumulated by plants are resistant to decay, it may be possible to infer historical changes in solar UV radiation from tissue samples of plants that have been preserved in herbaria or in sediment cores. Herbarium specimens offer the potential to retrospectively infer past environmental conditions by assessing how plant traits have changed over the period of their collection (usually decades). However, to be reliable proxies for UV radiation, herbarium specimens must be sampled in a consistent and unbiased manner over time (See [201] for a full discussion of necessary procedures). If these protocols are followed, and if other factors that can modify flavonoid and anthocyanin accumulation in plants (e.g. shading, changes in temperature, availability of moisture or total solar irradiance) are accounted for, one could associate trends in pigmentation of thermostable compounds with historical changes in UV radiation.

Over longer time frames the effects of major global events such as changes in solar activity, volcanic eruptions, or reversals in the Earth's magnetic field (e.g. at the Laschamps

Excursion 42,000 years ago) might be examined through changes in the phenolic composition of pollen or spores. For example, the hydroxycinnamic acid *para*-coumaric acid, which is preserved in fossilised sporopollenin (a compound that forms the outer wall of spores and pollen), continues to be the focus of research seeking such a proxy for UV-B radiation over geological time periods. However, before it can be reliably used it is necessary to ascertain the action spectrum of its response to UV radiation, its rate of degradation, the response time of its synthesis, as well as the consistency of response among species and over time [202]. In the case of fossilised pollen from *Nitraria* (a steppe plant) and conifers, chemical signatures have been shown to differ from those of contemporary (extant) pollen in a predictable and consistent manner such that stable relationships can be modelled [203].

### 3.3 Interactive effects of UV radiation and climate change factors

Ongoing changes in climate, together with associated changes in plant species distribution, are exposing wild plants, forests and crops to new combinations of UV radiation and other climatic conditions [10, 124]. Combinations of particular concern are high UV-B irradiance and drought or temperature, as climate change is increasing the frequency and severity of heat waves and droughts, and these events frequently coincide with high UV radiation, particularly at mid to low latitudes [204].

There are marked similarities in the acclimation responses of plants to increases in UV radiation and drought. A recent meta-analysis found these two sets of responses to be generally consistent irrespective of whether experiments were performed in controlled environments (i.e. growth chambers or greenhouses) or in the field [205]. In general, when plants are exposed to co-occurring drought and increased UV irradiance, the accumulation of defence compounds (e.g. proline and secondary antioxidants, such as flavonoids and anthocyanins) and other stress responses (e.g. decreased leaf area, reduced stomatal opening) is enhanced. Thus, the combined detrimental effects of these stressors on plant function are milder (i.e. reduced production of stress-associated malondialdehyde (MDA) and reactive oxygen species (ROS)), and this reduces the negative effects on photosynthesis and biomass production [205, 206]. The response of plants to increased UV radiation may therefore confer cross-protection against drought [205, 207] and mitigate some of the detrimental effects of drought on plant growth and productivity, unless both stress factors are excessive. Further, it has been postulated that plants may use UV radiation as a signal of impending drought [208]. The functional association between exposure to drought and UV radiation exposure appears to involve common physiological defence and acclimation responses [208, 209]. For example, multiple

studies have shown that overexpression of protective pigments in plants results in enhanced protection against both drought and UV-B radiation [210, 211]. UV-B radiation can even be exploited for seed priming, resulting in enhanced expression of drought tolerance of plants grown from such UV pre-treated seeds [212]. Certain agricultural practices may also negatively impact crop tolerance of both UV radiation and drought. For example, growth allocation to roots relative to shoots often increases in drought-stressed plants, as well as those exposed to high solar UV-B radiation (i.e. increased root:shoot ratios), but high nitrogen availability has the opposite effect on root–shoot allocation [213].

High UV-B irradiance often co-occurs with high temperatures. A recent study of a commercial tomato cultivar (*Solanum lycopersicum* cv. Money Maker) compared plants transferred under near-ambient solar UV radiation to those placed in a UV exclusion treatment in the field. Exposure to UV-B radiation led to partial closure of leaf stomatal pores, reducing transpiration and evaporative cooling, and thus increasing leaf temperature by up to 1.5 °C [214]. These findings are relevant in warmer climates where even small increases in temperature may have substantial consequences for survival of crops [215], as high temperatures are well-known to negatively affect photosynthesis and growth of many plant species. More broadly, a recent meta-analysis across terrestrial, freshwater and marine plants, algae and animals [216] showed that any negative effects of UV-B radiation can be somewhat compensated for by elevated temperatures, although this depends on the habitat and organism involved. This positive effect of warming appears to be restricted to cool climates where organisms often function at temperatures below their physiological optima, and thus is not expected to occur in environments approaching the thermal and physiological limits of organisms [216]. Given the current context of global warming, more detailed temperature and UV-radiation dose–response studies are required to fill this knowledge gap. Furthermore, the scope of such studies needs to go beyond crop yield, as early evidence shows that interactive effects of heat and UV radiation can also affect crop quality [217] (Sect. 5.2).

Apart from high temperatures, the effects of UV radiation on plants can also be modified by low temperatures, and climate change is expected to increase the incidence of extreme cold events in some regions [20] (Sect. 2.2). In studies with the model plant *Arabidopsis thaliana*, the synthesis of flavonoids is strongly enhanced in response to low temperatures (4/2 °C, day/night) compared to moderate temperatures (18/20 °C), just as it is by UV radiation. Where plants are simultaneously exposed to both cold and UV radiation, complex interactive effects are observed, with UV-B decoupling flavonoid accumulation from gene expression, indicating post-translational regulation [218]. Low temperatures and UV-B radiation also produce

a shift in the composition of flavonoid glycosides from kaempferols to quercetins [176]. The shift in composition towards quercetin synthesis at low temperatures suggests an enhancement in antioxidant function [176, 219], which could increase overall plant hardiness.

Temperature is a cue for many organisms, controlling their seasonal development (i.e. phenology). Changes in thermal regime, such as periods of extreme heat or cold or even an absence of cold temperatures, can disrupt the timing of growth, reproduction, and other aspects of phenology [220, 221]. Temporal shifts in phenology can also change the seasonal timing of exposure to UV radiation, as solar UV radiation varies at high-to-mid-latitudes over the course of the year. Shifts in phenology due to changes in climate and UV radiation may result in new combinations of biotic interactions (i.e. competitors and pests; Sect. 4.1) and abiotic stresses that may be outside the tolerances for some species. For plants, these new combinations of abiotic stresses can have detrimental effects on their growth and survival even though each individual stressor may have a negligible effect [222].

Complex effects on plants may also occur when other environmental factors interact with UV radiation. Recent studies have revisited the interactive effects of UV radiation and increased nitrogen deposition [223], ozone pollution [15, 224] and elevated atmospheric CO<sub>2</sub> concentration, where short-term stimulation can be outweighed by long-term downregulation of photosynthesis [225], as noted in our previous assessments [12, 226]. Elucidation of the interactive effects of UV radiation and these other environmental factors is necessary to improve our ability to model and assess the effects of UV radiation on the carbon sequestration of terrestrial vegetation in a changing climate (e.g. Box 1).

## 4 Species distributions and biodiversity

Maintaining the wide variety of plants, animals, and micro-organisms in terrestrial environments (i.e. biodiversity) is essential for ecosystem health, stability, and valuable services provided to humans. The loss of biodiversity can occur directly (e.g. hunting or harvesting) or indirectly (e.g. loss of habitat, climate change, and invasive species). While considerable attention has been given to the effects of climate change on biodiversity [21, 227, 228], far less is known about how solar UV radiation might interact with climate change to influence species distributions and diversity in ecological communities. We examine these effects from available studies and evaluate how the UV radiation exposures of species can potentially change as their distributions shift in response to climate change.

## 4.1 Potential effects of climate change and UV radiation on shifting species distributions

Plant and animal species are migrating or shifting their distribution ranges to higher elevations and latitudes in response to ongoing changes in climate [229–231]. As species occupy higher elevations and latitudes, they may encounter increased or decreased UV radiation, respectively, because of the natural gradients in solar UV radiation that occur with elevation and latitude (Table 1). Some plants and animals are also shifting their ranges in the opposite direction, viz., towards lower elevation (lower UV radiation) and latitude (higher UV radiation), to avoid the increased seasonality of temperature at higher latitudes [232, 233]. How species respond to novel combinations of UV radiation and multiple climatic conditions has direct implications for how they will interact with other species, including their pests and pathogens (Sect. 5.3), with consequences for biodiversity [e.g. 235].

### 4.1.1 Latitudinal change

While the changes in UV radiation received by plants and animals resulting from latitudinal shifts in ranges are generally rather modest, they may affect terrestrial ecosystems and biodiversity. For instance, if one assumes species migrate at their maximal rates to keep pace with climate change (i.e. their average climate velocity for the period 2050–2090; [235]), the UV irradiance under clear sky conditions for herbaceous plants would decline by 4.5%, while that for more mobile plant-eating insects would decline by 16.2% after a century of climate change (Fig. 2A).

Plants encountering reduced UV-B irradiance resulting from range shifts would likely reduce their levels of UV-protective compounds (i.e. epidermal flavonoids and other phenolic compounds) [236, 237]. The multiplicity of roles performed by these plant secondary compounds could, in turn, make some plants more vulnerable to herbivores [238, 239] (Sect. 5.3) as some of these chemicals serve as deterrents for insect herbivores.

Accelerated loss of biodiversity will likely occur as climate change continues to exert its effects on range shifts on plants and animals. Plants use both temperature and day length (photoperiod) as environmental cues for regulation of phenology (flowering, dormancy, budburst, etc.). Trees with long generation times may be especially vulnerable to extinction because they have limited opportunities to genetically adapt to a changing photoperiod and their environmental cues such as UV radiation may be mismatched with their new environment [240–242]. At present, it is unclear how changes in UV radiation in combination with climate change will affect species migrations and adaptation as experimental

and modelling data are not yet available to quantify and fully assess the risk of these interactive effects.

### 4.1.2 Elevational change

For many montane ecosystems, climate change is resulting in the migration of lower elevation species to higher elevations. Climate change is also reducing the envelope of suitable habitats for high elevation alpine species to survive, while increasing competition against emigrating species from lower elevations [21, 243]. However, like latitudinal shifts towards the equator, elevational distribution changes also occur downslope for some species [232, 233]. For species shifting their ranges to higher elevations, their exposure to solar UV radiation would be expected to increase, assuming no change in cloud cover (Fig. 2B). Further, reduced snow cover due to warmer temperatures exposes organisms to fluctuations in temperature and solar radiation, including UV-B radiation [244].

High elevation alpine plants often have heightened accumulation of UV-screening compounds and herbivore defence [140, 245–248]. Across a diversity of plant species from alpine and subalpine zones in Bulgaria, improved photoprotection has been found to effectively prevent greater DNA damage caused by increased UV radiation at higher altitudes. These mechanisms were sufficiently effective since plants growing at the highest elevations had fewer UV-induced DNA dimers than those at lower elevations, with grasses (Family Poaceae) least susceptible to UV-induced DNA damage among a wide diversity of plant families tested [249]. Nevertheless, as species migrate to high elevations, more resources may be allocated towards protection against UV radiation, and this could in turn divert resources away from growth, which could then reduce competitive ability [250, 251]. Depending on the availability of suitable habitats at higher elevations, these changes in species interactions have the potential to negatively affect biodiversity [252] by shifting the balance of competition between species [253, 254].

While climate change is causing many species to migrate to higher elevations, these climate change-induced shifts in distribution ranges are often most pronounced for non-native, invasive species [255–257]. At present, it is unclear if UV radiation affects native and non-native invasive species differently [140, 141, 258–260]. However, invasive species are generally considered to exhibit greater phenotypic plasticity to new environments than native species, although this may depend on availability of resources [261, 262]. In some cases, invasive species have been found to alter their production of UV screening compounds to a greater degree than native species [140]. This flexibility may allow non-native, invasive species to occupy new habitats more rapidly



than native species and, in some cases, outcompete endemic alpine species [234].

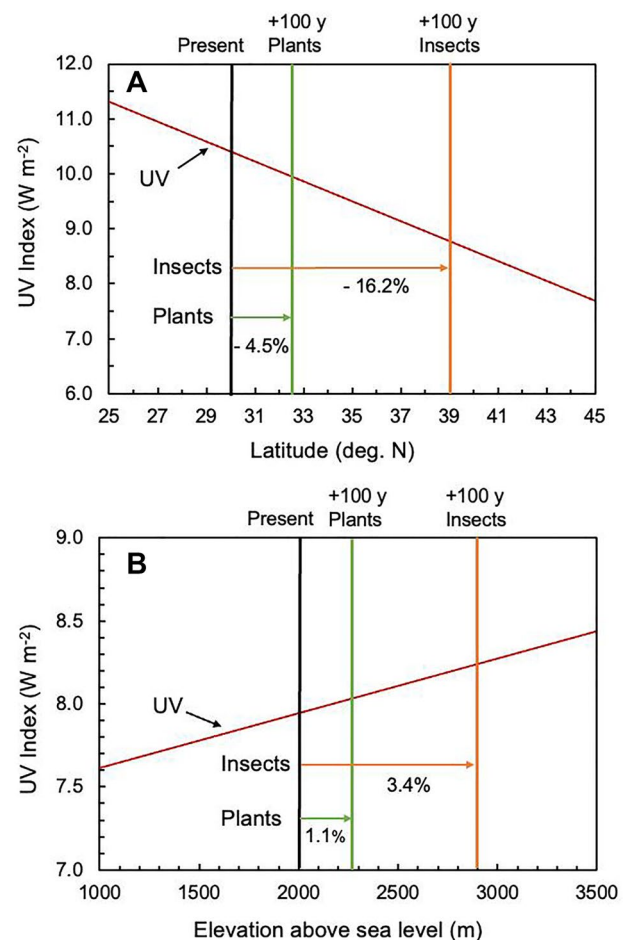
#### 4.2 Assessing the risks to biodiversity from the interactive effects of UV radiation and climate change

Climate change can cause declines in biodiversity by reducing the availability of suitable habitats for species and by differentially shifting their distribution ranges, which then disrupts species interactions and ecosystem function. If species cannot keep pace with climate change, then populations will decline leading to a loss of biodiversity. In this context, species distribution models (SDMs) are used to determine how climate change will affect future habitat suitability of species through changes in key abiotic drivers. These models can be used to inform species conservation as well as management for plant production in agriculture and forestry [265].

Several studies have shown that the inclusion of solar UV-B radiation in models forecasting future distribution ranges of ecologically and agriculturally important crop and tree species improves their statistical predictive power [266–272]. These models are based on different scenarios of climate change (i.e. IPCC scenarios of greenhouse gas emissions) and create projections based on correlative relationships between climate and species occurrence. These projections suggest that the ranges of some species of native plants from open, dry habitats found in arid and semi-arid shrub-steppe biomes will expand to higher elevations [267–272], while the ranges of willows and other related species from wetter habitats will contract [266].

Some studies of plants native to China and central Asia include UV-B radiation among the potential explanatory climatic variables that contribute to species distributions, sourcing data from the global climatology [273]. Future habitat suitability estimated using Maximum Entropy models (MaxEnt; models that apply basic machine learning algorithms to resolve environmental conditions where the species is present across its distribution) reveal incident UV-B radiation together with precipitation and temperature as significant correlates of species occurrence. While these models do not identify the mechanisms underlying these results, the findings suggest that such models could be useful in assessing risks to biodiversity, as well as providing information on potential species distributions and suitable habitats for conservation and planting crops for different scenarios of climate and solar UV-B radiation.

Despite the inclusion of UV-B radiation among significant climatic variables in some studies of species distributions [266–272], most modelling studies to date do not include UV-B radiation and its interaction with other abiotic stressors as potential constraints on species distribution. As



**Fig. 2** Potential changes in exposure to UV radiation as plants and insects migrate to higher latitudes and elevations with climate change. Panel **A** shows the estimated changes in UV radiation as plants and their herbivorous insects migrate poleward after 100 years (y) of climate change. UV radiation data are simulated midday summer (June 21) UV irradiances (here reported as UV Index; red line) based on stratospheric ozone levels in 1980 at sea level (radiative transfer model TUV; [263]). Horizontal arrows show distances migrated for herbaceous plants (green arrow) and plant-eating insects (orange arrow) originating from 30° N after 100 years of climate change assuming maximum rates of migration and average climate velocity for 2050–2090 (from [235]). Panel **B** shows the simulated midday summer (June 21) clear sky UV Index changes with elevation in the European Alps (46° N latitude; red line) and the estimated changes in UV irradiance for plants (green line) and insects (orange line) as they migrate from 2000 m to higher elevations after 100 years of climate change, assuming average current rates of leading edge migration for Western European montane plants (28.2 m/decade) and insects (90.5 m/decade) [264]

more detailed UV-B databases become available, it is likely that UV-B radiation will more routinely be included among climatic variables used to predict species occurrence and changes in biodiversity. As well as species distribution, climatological data that include regional UV-B irradiances can be applied to study whether climatic trends correlate

with patterns in plant functional traits among species. For example, a large-scale study of 1192 grassland species found that UV radiation was negatively correlated with leaf size across the Mongolian and Tibetan Plateau [274], while leaf shape, reflectance, and thickness have also been found to covary with UV-B radiation along environmental gradients [275–277].

## 5 Effects on agriculture and food production

Some of the earliest concerns raised over stratospheric ozone depletion and the accompanying increase in solar UV-B radiation considered the potential for reductions in crop productivity and compromised food security [278, 279]. A prior assessment [38] using results from field studies conducted at high latitude locations indicated that plant productivity declines by about 3% for every 10% increase in plant effective UV-B radiation (i.e. UV-B radiation weighted according to a generalised plant action spectrum [280]). These findings implied that the projected increases in solar UV radiation with changes in stratospheric ozone and climate, assuming full compliance with the Montreal Protocol, would have minimal effects on agricultural productivity. However, few experimental studies to date have been conducted on species growing in those regions with the highest natural levels of UV-B radiation on Earth (i.e. the tropics and high elevations). Previous studies also tended to focus on crop productivity but paid less attention to the effects of UV radiation on food quality. The effects of UV radiation on agroecosystems also extends beyond the direct effects on crop plants, as UV radiation can influence pest–pathogen interactions and the persistence and effectiveness of biocides and agricultural pollutants. The management of solar radiation in greenhouses and advances in artificial UV lighting are exploiting some of the beneficial effects of modest exposures to UV radiation to improve food quality, enhance plant defences against pest and pathogens, and contributing to more sustainable agricultural practices [281].

### 5.1 Agroecosystems vulnerable to changes in UV radiation and climate

As noted in our 2019 Update Assessment [282], and in other reports [283, 284], most field research to date on the effects of UV radiation on crops has been concentrated on regions outside the tropics and at lower elevations. The tropics extend over approximately 33% of the Earth's land surface [285] and harbour a vast reservoir of biodiversity [286] that provides critical resources and essential services for agriculture and food security [287]. Thus, tropical agroecosystems

warrant further attention to safeguard a sustainable future for life on Earth.

Because the projected recovery of stratospheric ozone is highly dependent on changes in GHG concentrations and lifetimes of ODS, there remains some uncertainty about how UV-B radiation might change in the future for tropical regions [10, 288]. Under some scenarios, UV-B radiation could increase by 3% in the tropics due to interactions between stratospheric ozone, climate and aerosols [8, 10]. This increase would further elevate the already high levels of UV-B radiation that occur naturally at low latitudes. At present, the consequences of these relatively modest percentage increases in UV-B radiation on crops or wild plants in this region are uncertain. Available evidence suggests that current levels of UV-B radiation in the tropics can alter the morphology (e.g. smaller leaves, reduced shoot height) and chemistry (e.g. increased flavonoid levels) of native, non-crop tropical plants, but that biomass production is rarely decreased in these species (e.g. [289]). By comparison, several field experiments have shown that certain varieties of temperate-zone crops (e.g. wheat and soybean) [290–292] show decreases in photosynthesis and yield when grown under ambient UV-B radiation in the tropics. These findings suggest that some important crop species grown in the tropics might be vulnerable to relatively small increases in UV-B radiation.

As noted above and in Sect. 4.1, climate change is shifting bioclimatic zones and this is allowing certain crops to be grown at higher elevations than was previously possible [293–298]. For some crop species originating from lower elevations, the more intense UV radiation at high elevations may exceed their tolerances to UV radiation with negative consequences for their physiology and growth [299]. As crop species are grown in these new habitats, they will also encounter new weeds, pests and pathogens, which may disrupt the structure and function of these agroecosystems [300, 301]. Differential effects of climate change on range shifts and phenology can also lead to spatial and/or temporal or seasonal mismatches between pollinators and their plant hosts [302, 303], posing additional risks to food security. Many of these high-elevation agroecosystems support community livelihoods and are important carbon sinks that help mitigate global warming. Thus, their risks from changes in climate and UV radiation are of particular concern.

### 5.2 Effects of UV radiation on food quality

Laboratory and field studies have found significant effects of UV radiation on crop quality with regard to texture, flavour, appearance and nutritional content. It is now well-established that the concentrations of a wide array of natural plant chemicals are modified by UV radiation [304–307] and these changes in chemical composition can have positive and negative effects on food quality. There is abundant

research demonstrating that exposure to modest levels of UV radiation can improve food quality by enhancing crop flavour [308], taste [309], colour [310], nutritional content [311–314], and pharmaceutical content [315–317] in various plants. Given that the intake of fruits and vegetables of many consumers is well below recommended levels [318], the higher nutritional content of crops exposed to UV radiation may generate long-term health benefits. For example, Keffie et al. [319] used solar UV-B radiation to increase vitamin D in oyster mushrooms, which may alleviate vitamin D deficiency in humans. Some have proposed legal regulation for UV treatment of foods, including mushrooms [320].

In some cases, exposure of crops to UV radiation can lead to a decrease in their nutritional value for humans and livestock. For example, some species of tropical grasses show increases in tannins when grown under experimentally elevated UV-B radiation, and this would imply a reduced palatability of forage for cattle [321, 322]. High levels of UV radiation may also increase amounts of other anti-nutritional compounds in plants, such as oxalates, which are generally associated with kidney problems [323]. At present, the full scope of UV-induced anti-nutritional compounds is not fully known nor is the identification of crops most at risk to these changes.

### 5.3 Effects of UV radiation on plant interactions with pests and pathogens

The Food and Agriculture Organization of the United Nations (FAO) estimates that plant pests<sup>2</sup> cause a 20–40% loss in global agricultural production per year, costing ca. \$220 billion USD, with the impacts of invasive insect species adding another \$70 billion USD [324]. It is expected that climate change, including ECEs, will increase the incidence and severity of pests and pathogens in some regions, as these organisms colonise new previously sub-optimal habitats along latitudinal and elevational gradients [325, 326]. The climate-induced parallel range shifts of plants with latitude or elevation into new habitats may constitute additional stress from plant pests (Sect. 4.1) [327]. Rising concentrations of CO<sub>2</sub> and associated global warming together with regional increases in UV radiation may also act together to compromise food security through complex effects on plant pests and disease [328]. While our previous assessments have reported on UV-mediated increases in resistance to specific pests and pathogens [12, 38], we note

that there is a need for more detailed studies on the interactive effects of UV radiation, CO<sub>2</sub> and other climate change factors on plant interactions with pests and pathogens.

Exposure to UV radiation can confer increased resistance of certain crops to pests and diseases through changes in host physiology, morphology, and biochemistry. As noted in Sect. 3.1, UV radiation typically enhances the production of polyphenolic compounds, such as flavonoids. Some of these compounds enhance a plant's defence against herbivores and pathogens (e.g. viral, fungal or bacterial) [239, 282, 329]. Disease and pest attack will also elicit the production of increased amounts of these polyphenolic compounds that can make the host plant unpalatable or toxic (Sect. 5.2) and/or protect the plant through their antioxidant properties (e.g. scavenging of free radicals). These effects on pests or pathogen attack are part of a wider network of interactive effects on plant physiology and morphology potentially altering the susceptibility of crops to these threats [330].

Chemical biocides are widely employed to manage pests and pathogens in crops (Sect. 5.4). However, several biocontrol agents against insect pests have been developed and used as alternatives to chemical pesticides. Of particular interest is a group of fungi that are parasitic on insects (entomopathogenic fungi). Entomopathogenic fungi kill insects by penetrating the outer protective cuticle layer of specific hosts with the help of proteases [331]. These fungi live naturally in soils but can be mass-produced for application to crops where they have been used against pests including spittlebugs and locusts, which affect crops such as maize, sugarcane and beans [331], as well as against various insect pests in rice [332]. However, many of the entomopathogenic fungi are strongly inhibited by UV radiation and temperatures above 30<sup>0</sup> C, which affect their development and pathogenic function against certain insects. Therefore, these abiotic constraints are considered a major barrier to the use of entomopathogenic fungi in controlling insect pests [332–335]. However, the effect of solar UV-B radiation on these fungi remains to be confirmed through experiments where they are grown under realistic solar radiation conditions [336]. Such studies may also allow for selection of fungal biocontrol agents that are more tolerant to UV-B radiation and other climate factors, for use as biocontrol agents to safeguard economically important agricultural systems.

### 5.4 Effects of UV radiation on agricultural biocides

The widespread application of biocides (herbicides and pesticides) for controlling or killing harmful organisms in agricultural field settings results in some accumulation of these chemicals in water, soil, and atmosphere, and may also result in residues in agricultural products. Given that biocides are designed to be bioactive, their adverse effects on non-target organisms and humans are of concern. Direct

<sup>2</sup> A pest in this context is “any species, strain or biotype of plant, animal or pathogenic agent injurious to the plants or plant products”, as per the definition in the International Standard for Phytosanitary Measures No. 5 (ISPM) adopted by the Commission on Phytosanitary Measures of the International Plant Protection Convention.

and indirect photodegradation by solar UV radiation can potentially reduce the environmental residence time of pesticides [19]. However, photochemical degradation can also reduce the functional effectiveness of biocides as crop protectants, which may lead to greater amounts being administered by growers [337, 338]. Direct photodegradation of biocides occurs when a chemical absorbs UV radiation, leading to its breakdown into various degradation products [339]. Indirect photodegradation involves the reaction of the biocide with reactive intermediates formed when natural photosensitisers (e.g. nitrate) absorb solar radiation [340]. Not all biocides are subject to direct photodegradation under solar radiation. For biocides with an action spectrum for direct photodegradation only in the UV-C region (wavelengths 100–280 nm) and not extending into the solar UV-B, only indirect photodegradation occurs under solar radiation.

In the field, the exposure of biocides to solar radiation depends on the manner in which they are applied to crops, as well as the specific characteristics of the crops, including age and canopy structure, which determines their exposure to solar radiation. These factors, together with the chemical composition of the pesticide formulation determine the extent to which they are photodegraded in the field. For example, the additive (co-formulation compound) benoxacor, which is used as a safener (i.e. a compound used in combination with herbicides to reduce negative effects on crops) of the herbicide metolachlor, accelerates the photodegradation of the active ingredient on soil surfaces, lessening its toxicity [341]. The extent to which biocides are photodegraded is also highly dependent on where the biocide residues occur. For example, the photodegradation rate of the herbicide imazethapyr is two orders of magnitude slower when applied to maize and soybean leaves than in aqueous solutions [342]. The leaves of aromatic herbs like thyme emit volatile organic compounds that can further affect the photodegradation of biocides deposited on their leaf surfaces, resulting in the formation of different photoproducts [341, 343–345]. Thus, the importance of direct vs. indirect UV-mediated photodegradation of biocides in the environment appears highly context dependent, and requires further research across a range of crops, environmental conditions and methods of application to clarify modes of action.

Climate change may be an additional factor impacting pesticide photodegradation on leaf surfaces. While photodegradation kinetics typically have a weak temperature dependence, pyrethroid insecticides applied onto spinach plants grown at 16–21°C degraded up to 2 times slower than when plants were grown at lower temperatures (10–15 °C), likely due to differences in the chemical composition of leaf wax [346].

As observed for other contaminants [e.g. 348,112] biocide photodegradation products can be more toxic than their

parent compounds. For example, some breakdown products generated by UV-B radiation of the fungicide chlorothalonil and the insecticide imidacloprid on plant leaves are more toxic to fish than their parent compounds [344].

Functional nanopesticides are being developed using nano-emulsion technologies as an alternative to traditional pesticide applications [348–351]. Encapsulated pesticides in nano-carriers, such as polymers, nanoclays, and metal organic frameworks provide controlled-release kinetics and improved stability against environmental degradation by UV radiation. The use of encapsulated pesticides prevents undesirable pesticide losses and release into the environment that otherwise would cause ecological and health concerns [352]. The development of nano-biocides may contribute to more environmentally friendly and sustainable food production systems (Sect. 7), potentially protecting the integrity of biocides during their application on crops, while still facilitating subsequent degradation of their residues.

## 5.5 Development and application of UV lighting systems in agriculture

Concerns over the effects of elevated UV-B radiation resulting from ozone depletion on food production stimulated considerable research into the effects of UV-B radiation on crops, and much of this early research focused mainly on the leaf-level physiology and shoot growth of traditional crop plants (e.g. soybean, rice, maize; [353]). More recently, studies have examined effects of UV-B radiation on plants of medicinal value, mushrooms and algae [354]. For example, mushrooms [355, 356] and certain microalgae [357] synthesise increased amounts of vitamin D after being exposed to UV-B radiation (Sect. 5.2) [357, 358]. In addition, more attention is being given to studying the effects of UV-B radiation on seeds, fruits, subterranean organs (e.g. roots and tubers), and on derived products, such as wine and olive oil [312, 359–362].

Results from these studies indicate that plants exposed to low or moderate levels of UV-B radiation in controlled environments (e.g. greenhouses, growth chambers) often have improved vigour, enhanced nutraceutical quality and are more resistant to pest and pathogens compared to plants that are grown in the absence of UV-B radiation, as typically occurs in commercial production glasshouses [281].

Other studies have shown how the application of UV-B radiation can modulate different physiological processes important for agriculture. These advances include, (1) accumulation of anthocyanins and other antioxidants in different coloured fruits, such as peach, apple, grapes, and blueberry [313, 362–365] (Sect. 5.2); (2) improving the tolerance of rice and tomato to low temperatures, salinity and drought [366, 367]; (3) the manipulation of flavonoid accumulation in vegetables [368]; (4) the production of smaller cucumber



plants for targeted commercialisation [174]; (5) an increase in anticancer compounds in *Catharanthus roseus* [369] following treatment with a combination of hormones and UV radiation; and (6) extending the shelf-life of fruit by reducing the activity of enzymes involved in fruit rotting [370]. Also, the accumulation of bioactive compounds can be triggered more effectively by applying high UV-B radiation during short periods in specific developmental stages (frequently near harvest) rather than using UV-B radiation over longer periods. This approach has been successfully applied in kale and grapes [361, 371, 372]. These advances have been achieved by translating research that was conducted to better understand the effects of increased UV-B radiation resulting from ozone depletion into commercial practices to improve food quality and production (Sect. 7).

One of the more significant technological advances in plant UV research and horticulture has been the development and use of UV light-emitting diodes (LEDs). Increasingly, LED lighting systems are being used by growers before and after harvest to improve food value. LEDs are more energy-efficient and environmentally friendly than most traditional light sources used in horticulture (e.g. high-pressure sodium vapour or metal halide lamps), and by utilising LEDs that emit both in the UV and PAR regions the control of the spectral composition, intensity and exposure period can be attuned to the light requirements of specific plants and crops [373, 374]. However, at present, only UV-A LEDs have been widely adopted to stimulate the accumulation of desirable plant compounds [375]. There are also some examples of successful application of UV LEDs in reducing certain plant diseases [376] and increasing nutritional quality [375].

## 6 Effects on biogeochemical cycles and climate feedbacks

Terrestrial ecosystems provide many valuable services, including the processing of dead organic material and the storage and recycling of essential nutrients. Both land vegetation and soils are also important carbon sinks that influence the concentrations of atmospheric CO<sub>2</sub> and hence climate. Solar UV radiation affects carbon storage and atmospheric CO<sub>2</sub> by influencing plant productivity [3], and the photodegradation of modern dead plant material (litter) and ancient organic matter preserved in permafrost soils [377, 378], which becomes exposed to solar radiation because of climate change-induced thawing [379–383] (Box 2). Changes in climate and UV radiation can further interact to alter the cycling of other elements (nitrogen being the most important) and the emissions of GHGs other than CO<sub>2</sub>, which can affect stratospheric ozone and climate. Below, we evaluate new findings that address the underlying mechanisms and

climate consequences of the interactive effects of UV radiation and climate change on biogeochemical cycles.

### 6.1 Photodegradation of plant litter

The decomposition of plant litter is a key biogeochemical process determining rates of nutrient cycling and energy flow in terrestrial ecosystems. This process affects vegetation productivity, carbon storage and soil fertility, and releases CO<sub>2</sub> and other GHGs to the atmosphere [384]. Thus, decomposition of litter has important feedback effects to the climate system.

In general, the rate of litter decomposition is regulated by climatic factors (temperature and moisture) and the chemical composition of litter (primarily the amount of lignin and the ratio of carbon to nitrogen (C:N ratio) in the litter), which modifies the activity and composition of the decomposer organisms (fungi, bacteria and invertebrate decomposers). Exposure of litter to solar UV radiation and short-wavelength visible radiation (i.e. blue and green light), can cause the direct breakdown of lignin and other plant cell wall constituents forming non-volatile and volatile compounds (e.g. CO<sub>2</sub> which is released to the atmosphere). This process is referred to as photochemical mineralisation or photomineralisation [384, 385] (Fig. 3A, right panel). Additionally, UV and short-wavelength visible radiation can also accelerate the breakdown of litter by changing its chemistry, making it more palatable to microbes and thereby enhancing microbial decomposition (Fig. 3A, left panel) [386–388]. Promotion of microbial activity can also occur by the photodegradation of waxy surfaces layers (i.e. leaf cuticle) that allows moisture to more readily penetrate litter [389]. These indirect effects of solar radiation on microbial decomposition are collectively referred to as photo-priming or photofacilitation [390, 391]. In some situations, solar UV radiation can negatively affect litter decomposition by altering the composition and activities of the decomposer community (not shown in Fig. 3) [392]. The overall effect of solar radiation on litter decomposition reflects the net effect of these three processes [390].

Among litter components, lignin has been identified as the most photoreactive due to its absorption in the UV and blue–green region of the solar spectrum [384, 391]. However, recent studies have found that cellulose and hemicellulose are even more susceptible to photodegradation than lignin [393, 394]. These discrepancies are an unresolved knowledge gap that could be addressed by identifying differences in the photodegradation action spectra for lignin, cellulose, and hemicellulose. The presence of polyphenolic compounds in plant litter (Sect. 3.1) decreases photodegradation under natural [395] and controlled laboratory conditions [396]. This result suggests that the accumulation of polyphenolic secondary metabolites in green leaves may persist during the early phase of litter decomposition and

attenuate the penetration of UV-B radiation into litter. The surface area of litter exposed to solar radiation is also an important predictor of litter decomposition rate and carbon turnover [397–399].

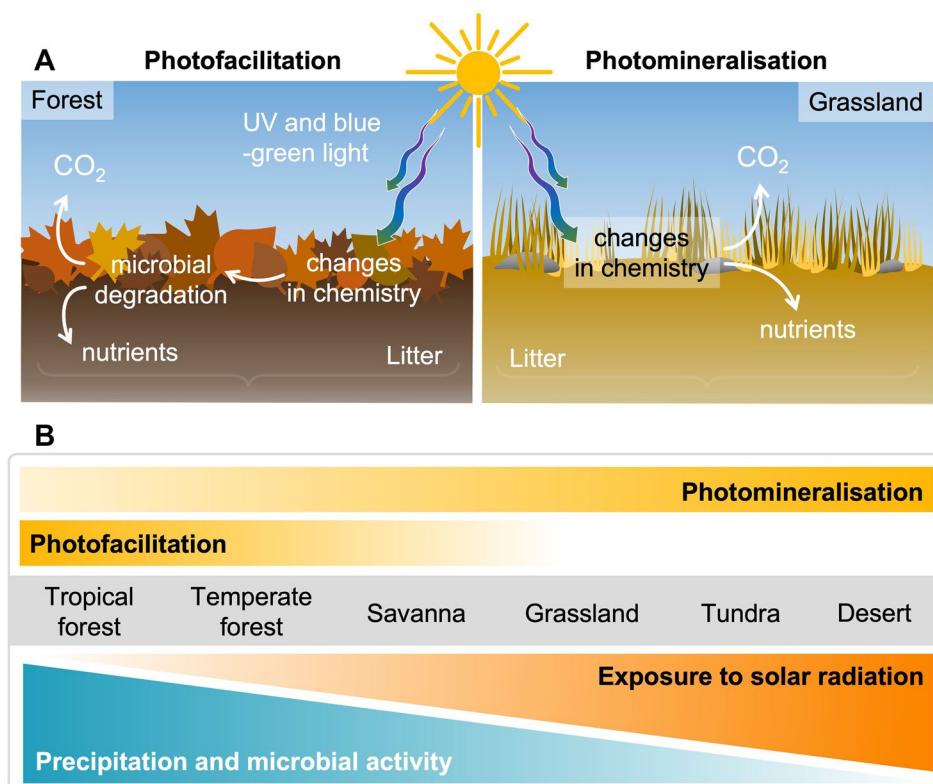
Photodegradation of litter was initially thought to be important only in dryland ecosystems (e.g. deserts and grasslands) where low moisture and high temperatures often constrain the activities of decomposing microbes. Recent studies have established that photodegradation of litter is important not only in semi-arid [400–402] and arid [403–406] ecosystems but also in moist environments that support tropical [407], subtropical [408], temperate and boreal forests [392, 395, 396], alpine steppe [409], and marshes [410].

Calculations of the strength of the terrestrial carbon sink have typically excluded photodegradation of litter in mesic ecosystems (having moderate water supply) due to their high vegetation cover. However, recent field studies found that photodegradation of litter facilitates carbon cycling in canopy openings of temperate and tropical forests, even where understory solar radiation is relatively low [395, 396, 407, 411]. Exposure to the full solar spectrum, resulting from the formation of a forest gap, can increase litter photodegradation rates by up to 120% relative to shaded conditions across a wide diversity of plant species [395]. This number is considerably higher than that for photodegradation in semi-arid regions (60%) [412] or across several habitats or biomes

(23%) [386], underscoring the importance of forest disturbance in mesic ecosystems. Exposure to solar radiation alters lignin structure of litter in the early stages of decomposition, promoting litter degradation via photofacilitation. This fact highlights the role of photofacilitation in mesic ecosystems, where higher water availability favours microbial decomposition compared to drylands [387, 413] (Fig. 3). On the other hand, relatively high UV radiation, which occurs during the time of the year when the forest canopy is leafless, may also have an inhibitory effect on microbial decomposers [396]. The seasonal consequences of these effects of UV radiation on understory microbes and overall ecosystem health and function remain unclear.

Recent studies have clarified the relative importance of the different wavelengths of solar radiation (i.e. UV-B, UV-A and blue–green) in driving photodegradation of litter and these findings have implications for the effects of ozone depletion on this process. A recent meta-analysis found that, globally, solar radiation increases litter mass loss by 15.3 ( $\pm 1.0$ )% relative to litter that has not been exposed to solar radiation [414]. The contribution of UV-B radiation was found to be significant only in specific environments, causing an 18% and 23% loss of litter mass in semi-arid regions and polar regions, respectively. The relatively limited importance of UV-B radiation in promoting loss of litter mass agrees with the results obtained with a new spectral

**Fig. 3** The relative importance of photomineralisation and photofacilitation in litter decomposition across terrestrial biomes and environments. Panel **A** illustrates the processes of photofacilitation and photomineralisation in the photodegradation of surface litter exposed to solar radiation (UV radiation and blue–green light) in representative wet (forest; greater photofacilitation) and dry (grassland; greater photomineralisation) ecosystems. Panel **B** shows relative changes in photofacilitation and photomineralisation across biomes and along gradients of moisture, microbial activity and exposure to solar radiation. Not shown in this figure is the potential leaching of non-volatile breakdown compounds resulting from photodegradation of litter that can occur when it rains, and possible negative direct effects of UV radiation on microbes

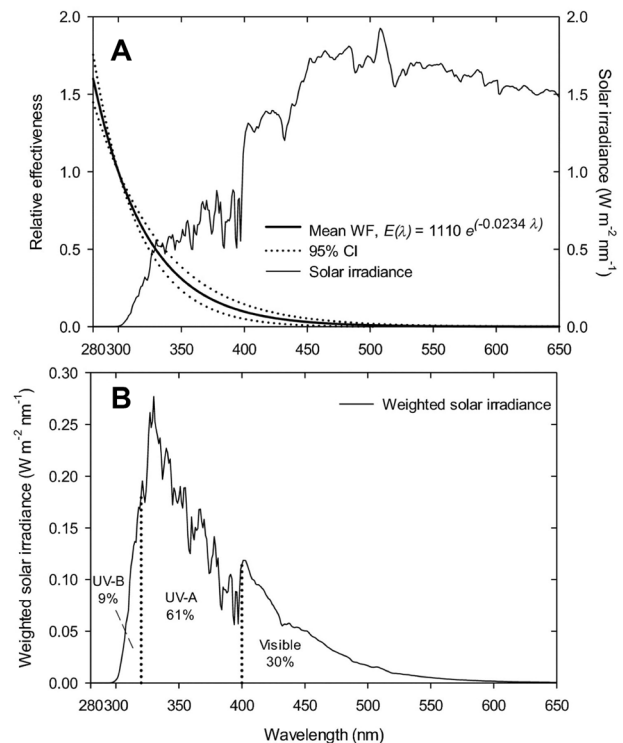


weighting function for litter photomineralisation, which showed that UV-B and UV-A radiation, together with visible blue-green light, are responsible for 9%, 61% and 30%, respectively, of total photochemical (abiotic) carbon loss [404] (Fig. 4). Overall, these rather small effects of UV-B radiation suggests that litter photodegradation would be minimally affected by further changes in stratospheric ozone.

Photodegradation can also be influenced by changes in vegetation cover and soil erosion that result from changes in land use and climate, including ECEs (Sect. 2.2). The loss of forests and other natural or semi-natural vegetation cover due to agricultural practices increases photodegradation of surface litter [19, 397], such that deforestation and land clearing will accelerate release of carbon from the ecosystem and alter patterns of GHG emissions and nutrient cycling [395, 402, 415, 416]. In dryland ecosystems, litter position (e.g. at the soil surface vs. buried or covered in dust) is the predominant factor determining carbon loss from photodegradation [398, 417]. In contrast to forests, these dryland systems are experiencing an increase in woody plant cover as a result of changes in land use and climate and these vegetation shifts result in more shading of ground litter and increased soil erosion and deposition, which decrease litter photodegradation [418, 419]. Additional environmental changes such as increased nitrogen deposition and abandonment or less intensive use of agricultural land may slow litter decomposition through the attenuation of surface UV radiation by increased plant canopy development [409].

Rainfall is another factor affecting litter photodegradation. In an experiment performed in drylands, the addition of supplemental precipitation (simulating a 2.7 times increased rainfall) accelerated loss of litter mass by a factor of 2.6 under near-ambient solar radiation but had no effect if litter was not previously exposed to solar radiation [388]. This result suggests that photodegradation followed by leaching may be another significant mechanism of loss of litter mass in arid ecosystems [388, 420].

Collectively, these findings indicate that the overall effect of photodegradation on the decomposition of plant litter depends on environmental conditions (primarily moisture and temperature), litter quality, the degree of exposure of litter to solar radiation (as influenced by vegetation cover, litter position and degree of soil–litter mixing), and the solar spectral composition of radiation reaching the litter layer [12]. Given the relatively small contribution of UV-B radiation to loss of litter mass and photomineralisation, ongoing and projected changes in stratospheric ozone and their interaction with climate and land-use changes are likely to impact litter photodegradation mainly by modifying its exposure to total solar radiation [12, 19, 145].



**Fig. 4** Action spectrum and weighted solar radiation for the photomineralisation of litter from plants in the North American Sonoran Desert. Panel **A** shows the mean weighting function/action spectrum for the photo-mineralisation of plant litter (heavy solid line; measured as CO<sub>2</sub> loss), with 95% confidence interval (dotted line; CI), along with the average solar noon spectral irradiance over the time period of the study (thin solid line). Panel **B** shows solar radiation at solar noon weighted according to the action spectrum in Panel A, along with the total % effectiveness of the solar UV-B, UV-A and visible wavebands. Adapted from [404]

## 6.2 Photochemical release of nutrients from terrestrial ecosystems

Most studies of photodegradation of organic matter in terrestrial ecosystems have focussed on effects on carbon but, as demonstrated in aquatic ecosystems [111], UV radiation can also affect the storage and cycling of other elements, such as nitrogen and phosphorous. Even in understory environments, where the amount and spectral composition of solar radiation is greatly modified by canopy structure and phenology, UV-B radiation [395], UV-A radiation and blue light can promote the conversion of organic nitrogen into inorganic compounds (nitrogen mineralisation) [411, 415].

A recent meta-analysis of litter degradation studies found that the amount of UV radiation received affected the timing of nitrogen and phosphorous loss compared

to that of carbon [413] due to differences in the relative contribution of microbial vs. photochemical degradation. Under reduced UV radiation, nutrient mineralisation was slow and poorly correlated with overall loss of litter mass, whereas, under increased UV radiation, phosphorous and nitrogen mineralisation was rapid and correlated with carbon mineralisation. These results suggest that microbial processes dominate nutrient cycling under low levels of UV radiation, while abiotic processes, which are characterised by a simultaneous release of nutrients and carbon, are more important at higher UV irradiances. Thus, under

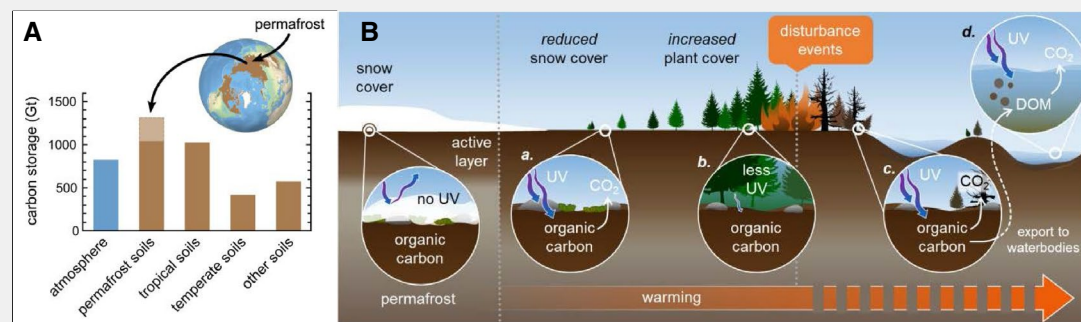
conditions of high UV irradiation the nutrients in litter may be made more rapidly available to plants, potentially reducing competition for nutrients between plants and microbes. These effects could play a significant role in ecosystem functioning but have not yet been thoroughly studied. The release of mineral forms of nitrogen is also likely to produce volatile nitrogen compounds including nitrous oxide ( $\text{N}_2\text{O}$ ), which is both a powerful GHG and ODS [19]. Given the obvious implications for climate and stratospheric ozone, the effect of UV radiation on  $\text{N}_2\text{O}$  emissions by litter remains a critical knowledge gap to be addressed in future studies.

### Box 2 Biogeochemical cycling of warming permafrost under climate change.

The world's soils store large amounts of carbon, approximately two-to-three times more than the atmosphere (Panel A below; [378]). Therefore, even small instability or degradation of soils driven by perturbations in climate can lead to large releases of carbon. A large proportion of soil carbon is stored at high northern latitudes, where it has remained stable in peatlands and permafrost soils over long time periods; often many thousands of years.

Permafrost is defined as any ground that remains completely frozen for at least two consecutive years. There is extensive evidence that climate warming is causing permafrost to thaw [381]. This releases large amounts of organic carbon that was previously locked underground, whereby it becomes available for microbial and photochemical decomposition [384]. These two processes release dissolved organic matter (DOM) and greenhouse gases (GHGs) such as  $\text{CO}_2$  [382, 383]

The effects of climate change on vegetation growing at high latitudes differ according to the region and vegetation type. These effects are illustrated in Panel B below. Warmer winters and springs mean that less precipitation falls as snow, and that the snowpack melts earlier. Reduced snow cover allows more solar radiation to reach the soil, thereby increasing photodegradation (B, circle a). A smaller snowpack also exposes vegetation to the environment above the snowpack for longer, leaving it more susceptible to damage and desiccation (referred to as Arctic browning; [84]). The resultant loss of vegetation cover can lead to soil erosion and loss of ecosystem stability [86, 87]. In this scenario, these ecosystems become sources of carbon and release nutrients. Moribund vegetation is unable to take up these nutrients, more of which are leached into waterways or emitted as gases [384], including the potent greenhouse gas  $\text{N}_2\text{O}$  (Sect. 6.2). Elsewhere, warmer temperatures may extend the growing season and nutrient release from thawing permafrost, which will increase plant growth and vegetation cover (Arctic greening). Additional shade from vegetation intercepts solar radiation before it reaches the soil, thereby limiting photodegradation (B, circle b). However, warming also leads to increased disturbance in these ecosystems, including more frequent wildfires, drought, floods, heatwaves, and herbivore outbreaks. Disturbance events generally destabilise the soil and contribute to the release of organic carbon from permafrost to waterbodies (B, circles c-d) [80], where it is subject to continued photodegradation to  $\text{CO}_2$  (B, circle d.; see also [112]).



**Panel A** Comparison of carbon stocks in various type of soils (brown bars) and the atmosphere (blue bar). The lighter extended bar gives the contribution from permafrost soils greater than 3 m depths that is not relevant to the other soil types. Reproduced and modified from [378]. **Panel B** Schematic showing the effect of warming on permafrost. Prior to climate change, soils and vegetation were covered for much of the year (left). Warming leads to reduced snow cover, permafrost thaw, and subsequent increased frequency of disturbance events like fire, floods, droughts and insect damage causing dieback. Inset circles illustrate how solar UV radiation interacts with these climate driven processes in soil (a-c) and waterbodies (d). Climate change is causing the active top layer of soil to thaw during the summer and freeze again in the autumn – this active layer is becoming deeper and staying unfrozen for longer.



### 6.3 Methane emissions, UV radiation and plants

Methane ( $\text{CH}_4$ ) is a potent greenhouse gas, such that relatively small changes in its emissions can make a significant contribution to climate change [421]. In addition to anthropogenic emissions, methane is released naturally by terrestrial ecosystems, particularly wetlands [422, 423]. At present, solar UV radiation is not considered an important driver of methane emission from terrestrial ecosystems [9, 111, 424]. However, there is concern that climate changes associated with stratospheric ozone depletion at high latitudes (tundra and taiga ecosystems in the Northern Hemisphere and peatlands or wetlands in the Southern Hemisphere) may enhance methane emissions [19].

Plants often serve as conduits of methane produced by bacteria in damp soils [425]. They also contribute to methane emissions through photochemical mineralisation of pectin, waxes, and lignin by UV-B radiation, although this effect is deemed rather small [426]. Methane emission from plants is accelerated by interaction with other stressors such as herbivore damage and high temperatures [427]. Controlled experiments with Scots pine and Norway spruce under ambient conditions in Finland found a positive relationship between methane emissions and solar radiation, which was steeper at warmer temperatures [428]. Even then, in most habitats, direct emission from plants through photodegradation of pectin [429] is considered only a minor contributor to global terrestrial methane emissions [421, 428].

Methane emission from plants also occurs through microbial methane production in the heartwood of trees (reviewed by [421, 424, 430]). From there, methane can be released to the atmosphere by passing through the bark or through the plant's vascular system. This process is currently thought to be the main avenue of plant methane emissions in non-wetland environments, and it is modulated by the moisture and phenolic content of heartwood rather than by UV radiation [431]. Reactive oxygen species (ROS), which are produced in all organisms and can be enhanced by oxidative stress, also take part in reactions that can release methane. Additional research is required to at the global scale to provide for a more complete understanding of the effects of climate and UV radiation on terrestrial methane emissions [421, 424, 427, 432].

### 6.4 Interactions of UV radiation with fire-derived carbon

Forest fires are increasing in severity and frequency and will become even more prevalent as the climate continues to change [20]. Boreal forests are particularly vulnerable to fires as extreme warming is expected in this region [433–439] close to the Arctic circle. Forest fires directly contribute to climate change by releasing GHGs such as

$\text{CO}_2$ , methane, and nitrous oxide [15, 434]. Wildfires also provide an important pathway for opening soil surfaces to UV irradiation, leading to enhanced photodegradation of organic matter with consequent release of  $\text{CO}_2$  (Sect. 6.1; Box 2). Due to the incomplete combustion of wood and other biomass, fires convert a substantial fraction of vegetation into burnt biomass, termed charcoal or pyrogenic carbon (PyC) [436]. Recent estimates indicate that *ca.* 256 Tg carbon ( $\text{Tg}_\text{C}$ )  $\text{yr}^{-1}$  (range = 196–340  $\text{Tg}_\text{C}$   $\text{yr}^{-1}$ ; 1 teragram =  $10^{12}$  g) of biomass were converted into pyrogenic carbon between 1997 and 2016 [440]. During rainfall events following a wildfire, ash and pyrogenic carbon (estimated up to 203  $\text{Tg}_\text{C}$   $\text{yr}^{-1}$  in a modelling study [441]) reach nearby watersheds, resulting in increased loads of organic carbon, nutrients, and metals [442, 443]. The impact of wildfires on surrounding water bodies can last for years, affecting biogeochemical processes and drinking water quality [442, 444]. In addition, fire-derived aerosols can temporarily reduce incident UV radiation reaching the Earth's surface [144] and slow down UV-driven chemical processes in the troposphere [15].

Pyrogenic carbon includes a broad suite of chemicals such as anhydrous sugars, condensed aromatics (often named black carbon), and graphitic carbon [440]. The specific chemical composition of PyC depends on biomass type and charring temperature, and this composition affects its solubility, bioavailability, and photoreactivity [440, 443, 445, 446]. Adding to previous findings [440], recent studies confirmed that black carbon is the most photoreactive fraction of PyC [447] and that microbial mineralisation of PyC can be enhanced by prior exposure to UV radiation (i.e. photofacilitation) [443, 446], similar to plant litter (Sect. 6.1) and dissolved organic matter in water [111].

## 7 Sustainability and the Montreal Protocol

By protecting the stratospheric ozone layer and mitigating some of the effects of climate change, the Montreal Protocol and its Amendments are assisting in the implementation of several of the United Nations Sustainable Development Goals (SDGs). Many findings in our Quadrennial Assessment address SDGs and specific targets that are relevant to agriculture (*SDG 2: Zero hunger*) and terrestrial ecosystems (*SDG 15: Life on land*) (Fig. 5). Other relevant contributions of the Montreal Protocol are related to pollution and contamination (*SDG 3: Good health and well-being*), and climate change (*SDG 13: Climate action*). Specific SDG targets addressed by our findings are described below.

## 7.1 SDG 2: Zero hunger

While small increases in solar UV-B radiation do not appear to pose a threat to crop yield, extreme increases in UV-B radiation, as would have occurred without the Montreal Protocol, would likely have significantly decreased agricultural productivity (Sect. 5) and jeopardised SDG 2 and particularly 2.4 (*Sustainable food production and resilient agricultural practices*). Several studies have shown that moderate UV radiation can alter the chemical composition of food and medicinal plants (Fig. 5a). In most cases, UV radiation increases the nutritional profile of some crop species (e.g. by increasing the concentration of certain antioxidants, e.g. flavonoids), with potential long-term positive outcomes for human health (Sect. 5.2). The latter finding has motivated the development of agricultural practices (e.g. UV-transparent greenhouse coverings and UV-emitting LEDs) that exploit low and medium levels of UV-B radiation to enhance the nutraceutical properties of crops (Sect. 5.5). These practices can be directly deployed in both developed and developing countries to obtain food with an improved nutritional profile for increased food security.

## 7.2 SDG 3: Good health and well-being

Plants exposed to modest levels of UV radiation often display some increase in resistance to pests and pathogens (Sect. 5.3), which could lead to reduced use of pesticides. However, solar UV radiation also degrades certain pesticides (Fig. 5a). This may lead to increased application of pesticides (Sect. 5.4), which could increase the risk of exposure of workers and consumers to these chemicals as well as adding to soil pollution and contamination (SDG 3.9). The net result on pesticide use likely depends on many factors, including changes in UV irradiance, cropping system, and types of pesticides.

Insert Fig,

## 7.3 SDG 13: Climate action

Modelling studies indicate that the Montreal Protocol and its Amendments have played a critical role in protecting global carbon sequestration by terrestrial vegetation, which has, in turn, slowed the build-up of CO<sub>2</sub> in the atmosphere and reduced a certain amount of global warming (Box 1). Also, exposure of plants to modest levels of UV radiation, that would not have continued to occur without the Montreal Protocol, can improve their tolerances to drought (Fig. 5b) and enhance resistance to pests and pathogens, thereby making crops and natural ecosystems more resilient to climate

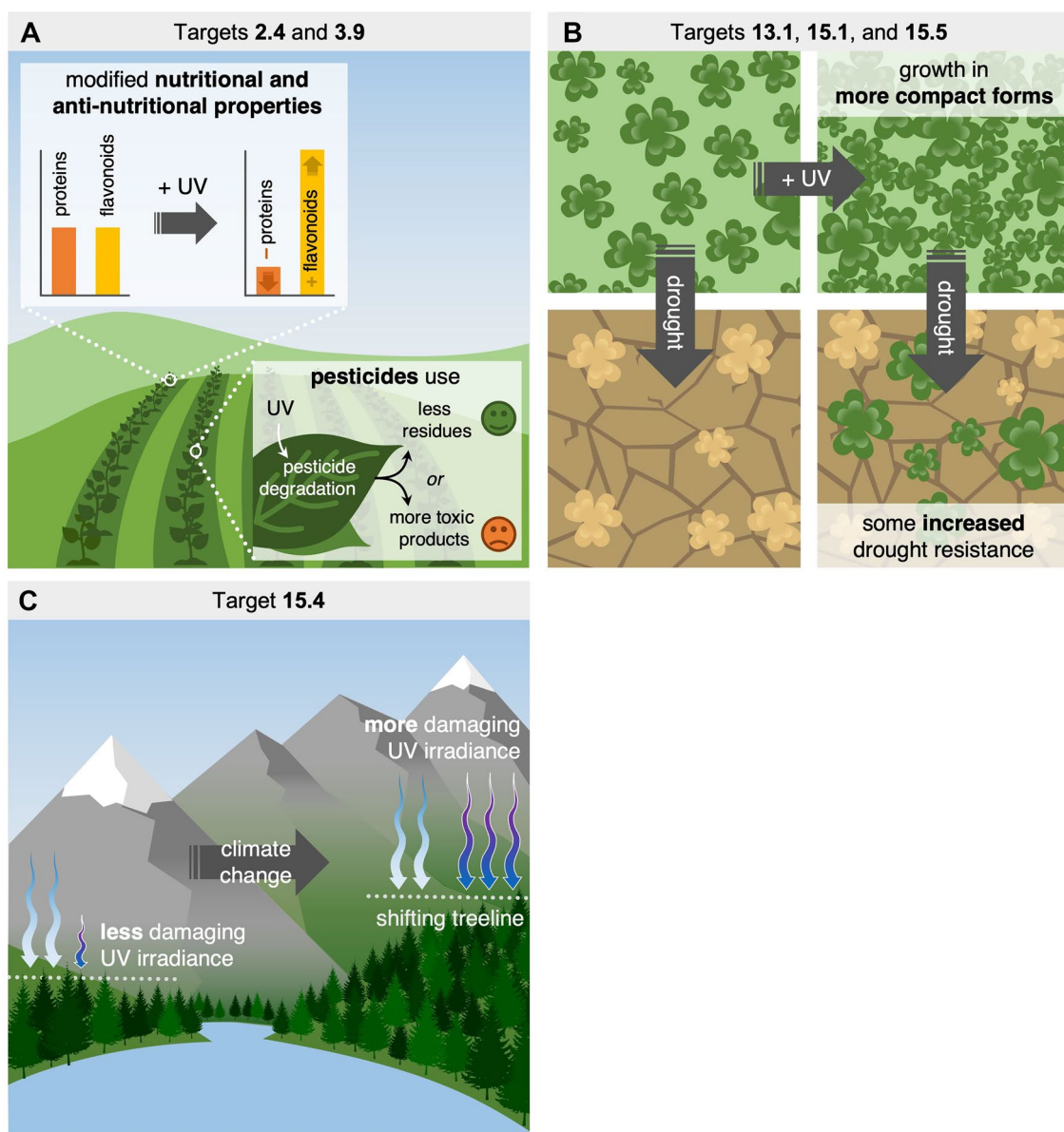
change (Sect. 2.2, 3.3; *SDG Target 13.1: Improve resilience to climate change*). Finally, this Assessment prepared for the Parties to the Montreal Protocol and as a scientific publication contributes to *SDG Target 13.3 (Improve education, awareness-raising and human and institutional capacity on climate change mitigation, adaptation, impact reduction and early warning)*.

## 7.4 SDG 15: Life on land

Increasing temperatures due to climate change are shifting the distribution ranges of plants and animals to higher elevations and latitudes, which changes their exposure to solar UV irradiation (Fig. 5c; Sect. 4.1). Some modelling studies suggest that UV radiation can be important in influencing the distribution shifts in plants (Sect. 4.2), which have the potential to negatively impact biodiversity (*SDG Target 15.1: Conservation of terrestrial ecosystems*). For mountain ecosystems, the shift to higher altitudes is often more pronounced for invasive species, which then occupy ecological niches of endemic alpine species with negative outcomes for biodiversity (*SDG Target 15.4: Conservation of mountain ecosystems*).

## 7.5 SDG 17: Partnership

Monitoring of the stratospheric ozone layer and its interactions with climate change are key to understanding the effects of UV radiation on terrestrial ecosystems, therefore the assessment of how species respond to this climatic pressure represents a challenge imposed on all countries. Partnerships between countries in Northern and Southern Hemispheres have been facilitated by the Montreal Protocol, which has stimulated technology transfer and innovation on the effects of UV radiation on plants and animals among scientific communities worldwide (*SDG targets 17.6, 17.7 and 17.8: North–South cooperation to access science, technology and innovation; Promote development, transfer and dissemination of environmentally sound technologies; and Science, technology and innovation capacity-building mechanisms for least developed countries*). This partnership has facilitated international support for data acquisition and sharing on the stratospheric ozone layer and the effect of UV radiation on terrestrial ecosystems (*SDG target 17.9: Enhance international capacity-building to achieve SDGs*). This has assisted least developed countries to have first-hand information for the implementation of environmental policies towards the achievement of SDGs (*SDG target 17.14: Enhance policy coherence for sustainable development*).



**Fig. 5** Pictorial representation of how the Montreal Protocol and its Amendments align with several Sustainable Development Goals (SDG) and their targets. Panel **A** shows SDGs 2.4 (*Sustainable food production and resilient agricultural practices*) and 3.9 (Deaths and illnesses from hazardous chemicals and soil pollution and contami-

nation). Panels **B** and **C** show SDGs 13.1 (Strengthen resilience and adaptive capacity to climate related disasters; centre panel) and 15.1, 15.4, 15.5 (Protect, restore and promote sustainable use of terrestrial ecosystems, sustainably manage forests, combat desertification, and halt and reverse land degradation and halt biodiversity loss)

## 8 Gaps in knowledge

In this assessment, we have identified several important knowledge gaps. These include:

- **Additional well-designed field studies are needed on all the topics addressed here to increase the confidence in our assessment.** It is well-established that the responses of plants and other organisms to UV radiation

are heavily dependent on other wavelengths of solar radiation as well as environmental factors such as temperature and moisture availability. There is also large inter- and intraspecific variation in sensitivity to UV radiation. Thus, the assessment of the effects of changes in solar UV radiation, stratospheric ozone and climate requires research conducted on a variety of species under natural, field conditions. However, studies conducted under controlled environmental conditions (e.g. growth chambers and glasshouses) can provide important insights into the

mechanisms of effects of UV radiation. In our assessment we have included certain studies carried out under controlled conditions when the results appear plausible and/or are useful for increasing awareness of potential effects and outcomes, but more field studies are clearly needed to reduce many of the uncertainties identified in this assessment.

- **Research into the impacts of Solar Radiation Management (SRM), such as Stratospheric Aerosol Injection (SAI), is needed to keep pace with policy-makers' interest in these technologies.** This is of particular concern, given that the existing evidence might suggest that impacts on terrestrial ecosystems of adopting SAI, and in particular any eventual termination or interruption of SAI following its adoption, are likely to be considerable, persistent and in some cases irreversible [24, 26–28, 448]. Importantly, some models of the effects of SRM on primary productivity by terrestrial ecosystems only draw on estimates derived from relatively simple and short-term calculations of changing canopy-level light-use efficiency under SAI scenarios. Experimental evidence of the relative importance of short-term responses vs the long-term acclimation of photosynthesis to the changes in spectral composition and irradiance brought by SAI have yet to be assessed through controlled experiments. Thus, we are not in a position to confidently assess the effects of SAI on ecosystem-level carbon assimilation at this time.
- **Experimental studies are needed to verify findings from modelling studies aimed at quantifying the environmental consequences of extreme levels of solar UV-B radiation, as would have occurred with uncontrolled emissions of ODS.** While these modelling studies are powerful approaches to understanding the benefits of the Montreal Protocol, and assessing the risks of future changes in the stratospheric ozone layer [e.g. 3], they rely on several assumptions that can lead to large uncertainties in the findings. As experimental studies on organisms exposed to these extreme amounts of UV radiation are lacking, it is often assumed that the effects of UV-B radiation on growth, productivity and reproduction observed under current UV radiation can be linearly extrapolated to higher amounts of UV radiation. This assumption is likely unrealistic, especially for the more extreme ozone depletion scenarios that would have eventually occurred without the Montreal Protocol. In addition, little is known about how the photomorphogenic responses of plants, which are driven by photoreceptors such as UVR8, are affected by extreme levels of UV-B radiation, or about the levels of UV irradiation where damage by the UV-B waveband supersedes the regulatory, photomorphogenic effects.
- **There is a critical need to develop action spectra for plants and other organisms, which more accurately describe biological responses to the different wavelengths of UV radiation under the full solar spectrum.** Action spectra are fundamental to interpreting biological responses to changes in UV radiation that occur with stratospheric ozone depletion and they also serve as spectral weighting functions in both laboratory and field experiments [449]. Large uncertainties in assessing the effects of ozone depletion can occur if inappropriate action spectra are used [450].
- **The establishment of long-term biomonitoring studies would improve our ability to assess how organisms and ecosystems will respond to the ongoing changes in UV radiation and climate.** Changes in UV radiation and climate, especially extreme climate events and combined extreme events (e.g. wildfires), pose significant risks to the health, stability, and biodiversity of terrestrial ecosystems, but little experimental or modelling data exist to quantify these effects. These studies are critically needed for organisms and ecosystems in polar regions, the tropics and high-elevation mountains.
- **The establishment of a global UV radiation biomonitoring network using material from selected organisms (from pollen to plants and animals) could further increase our knowledge and reduce uncertainties on the use of biological proxies for solar UV radiation.** Certain plant material and tissues, such as herbarium specimens and pollen in sediment cores, have the potential to serve as proxies for reconstructing past UV radiation environments on Earth, but presently there are large uncertainties associated with these techniques.
- **Studies are needed to characterise a wider array of interactive effects to adequately assess the consequences and map potential mitigation options of ongoing changes in solar UV radiation together with other contemporary environmental changes.** Advances have been made in understanding how UV radiation interacts with other climate change factors (e.g. UV radiation and drought) to affect the growth and physiology of plants [451] but studies need to be expanded to include multiple interactive factors (e.g. UV radiation, temperature, drought, CO<sub>2</sub> concentrations).
- **There is a need for additional biomedical research examining how UV radiation-induced changes in plant secondary metabolites affects dietary availability of metabolites, and the impacts of these changes on food quality and the epidemiology of human diseases.** Evidence continues to mount showing that exposure of plants to UV radiation alters their secondary chemistry and nutritional quality. But how these changes affect human health is largely unknown. This knowledge gap needs to be addressed to gain a fuller understanding of climate change-associated effects of UV radiation and their consequences for consumers, as well as the develop-



ment of more sustainable agricultural practices (Sect. 5.5 and 7).

- **Research is needed to better understand the effects of UV-B radiation on animals. In comparison to terrestrial plants and ecosystems, there are far fewer studies on the effects of UV-B radiation on animals.** While there are some similarities in experimental approaches used to study plant and animal responses to UV-B radiation (e.g. providing different UV-B radiation treatments using UV-emitting lamps) there are also some important differences that often limit the applicability of UV radiation research on animals. For example, plant research typically uses a filter material (e.g. cellulose diacetate) to remove the short-wavelength UV radiation that is present in lamps but not in solar radiation [336]. Most plant research has also taken into account the effects of different UV wavelengths using action spectra as biological spectral weighting functions in designing and interpreting experiments using UV radiation produced by lamps [336]. Not all studies of the responses of terrestrial animals, including insects and other invertebrates, adopt these approaches. In our assessment, these experimental deficiencies represent a significant limit in placing current understanding of invertebrate responses, mostly obtained using UV-emitting lamps, in the context of variation in solar UV-B radiation in the field.
- **Despite recent advances in understanding the ecological significance of photodegradation in the decomposition of plant litter, further research is needed to refine our mechanistic understanding of this process and assess its importance in the cycling of carbon and other nutrients, and feedbacks to the climate system.** Findings since our last Quadrennial Assessment have revealed that photodegradation of plant litter is not only important in drylands, but across all terrestrial ecosystems. These findings explain, in part, why traditional biogeochemical models of litter decomposition that do not include photodegradation are often inadequate in reproducing measured mass and carbon losses [401, 452, 453]. Despite this general finding, many knowledge gaps remain, notably the quantification of the relative importance of photomineralisation vs photofacilitation in both dry and mesic environments, and whether the spectral weighting function derived from studies in drylands also applies to mesic ecosystems. Nutrient cycling has also been much less studied than carbon cycling, and particularly how changes in nitrogen cycling caused by UV irradiance could feedback on climate change and stratospheric ozone depletion. Open questions also remain concerning the underlying chemistry controlling litter photomineralisation and the role of UV radiation in driving GHG emissions from the thawing of permafrost. Reducing these uncertainties would improve our ability

to assess how changes in UV radiation and climate will impact carbon cycling and feedbacks to the climate system.

## 9 Conclusions

The findings presented in this Quadrennial Assessment indicate that changes in stratospheric ozone, UV radiation and climate can interact in various ways to modify terrestrial ecosystems and biogeochemical cycles. While exposure to solar UV radiation, and in particular the short-wavelength UV-B radiation, has the potential to cause deleterious effects on plants, animals, and microorganisms, most species have evolved mechanisms to tolerate or avoid harmful solar UV radiation at the Earth's surface within the range experienced without significant ozone depletion. The extreme UV irradiances that would have occurred without the Montreal Protocol (i.e. "World-Avoided" scenarios) would likely have exceeded these tolerance limits and greatly reduced the productivity and biodiversity of terrestrial ecosystems. These conditions would also have driven increased photodegradation of organic matter and nutrient cycling, which would have increased emission of GHGs, including nitrous oxide, an ozone-depleting and greenhouse gas. Our findings further indicate that, in some cases, moderate levels of solar UV radiation (i.e. ambient UV irradiances without appreciable ozone depletion) can have some positive effects on organisms and the environment (e.g. improved food quality, enhanced plant defence against pests, improved plant vigour and resistance to other abiotic stresses, and the photodegradation of pesticides). Maintaining these beneficial effects of moderate UV radiation would have been impossible without the Montreal Protocol. Thus, the Montreal Protocol and its Amendments have played, and continue to play, a vital role in maintaining healthy, diverse ecosystems on land that can sustain life on Earth. The Montreal Protocol and its Kigali Amendment are also directly and indirectly protecting the Earth's climate and mitigating some of the negative consequences of climate change by limiting the emissions of GHGs and protecting the carbon sequestration potential of vegetation and the terrestrial carbon pool [3, 5].

Since our last full assessment [12], there have been additional extreme weather events (e.g. heat waves, droughts, and hurricanes) and events resulting from a combination of weather extremes and other drivers (e.g. wildfires) that have all contributed to the disruption and destabilisation of terrestrial ecosystems. These have been particularly pronounced in polar regions where anomalies in stratospheric ozone and ozone-driven climate change have occurred in the last three years [10]. Ozone depletion over Antarctica in certain years has coincided with early summer and has likely resulted in greater exposure to UV radiation of animals, plants and

microbes. These, and other extreme events (as outlined in Sect. 2.2), are expected to increase in frequency and intensity in the future because of climate change [20]. Together with other aspects of climate change, these extreme events will likely alter the UV radiation received by terrestrial organisms to a greater degree than the expected changes in the stratospheric ozone layer—assuming continued and full compliance with the Montreal Protocol. While understanding of the mechanisms of these UV-climate interactions is improving, the scale of their effects in terrestrial ecosystems remain poorly defined at present. Nonetheless, our findings indicate that the Montreal Protocol and its Amendments continue to make valuable contributions towards mitigating some of the negative environmental consequences of climate change as well as addressing several of the SDG targets established in the United Nations 2030 Agenda for Sustainable Development.

**Acknowledgements** Generous contributions by UNEP/Ozone Secretariat for the convened author meeting, and support for W-C.H (also partly supported by the National Science and Technology Council (NSTC) of Taiwan under grant MOST 110 2223-E-006 -004 -MY3). The following authors gratefully acknowledge support: PWB [J.H. Mullahy Endowment for Environmental Biology at Loyola University New Orleans, and the U.S. Global Change Research Program]. TMR [University of Helsinki; University of Cumbria; Norwegian Research Council (QUEST-UV project, and Academy of Finland (decision #324555)]. RGZ [US Environmental Protections Agency; the views expressed in this article are those of the authors and do not necessarily represent the views or policies of the U.S. Environmental Protection Agency]. MAKJ [Science Foundation Ireland (16-IA-4418). RO [Swiss National Science Foundation (SNSF), grant number P500PN\_206690]. Q-WW [CAS Young Talents Program and National Natural Science Foundation of China (41971148)]. SAR [Australian Research Council (DP180100113 & DP200100223)]. BF [RCN (Research Council of Norway), grant # 322954]. JM-A [MCIN/AEI/ and 'ERDF A way of making Europe' (grant PGC2018-093824-B-C42)].

**Author contributions** All authors contributed to the conception and assessment, and carried out extensive revisions of content.

**Funding** Open Access funding provided by University of Helsinki including Helsinki University Central Hospital. Open access funding was provided by the University of Helsinki, Finland.

**Data availability** All data generated or analysed are included in this article.

## Declarations

**Conflict of interest** The authors have no conflict of interest.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not

permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## References

- McKenzie, R., Bernhard, G., Liley, B., Disterhoft, P., Rhodes, S., Bais, A., Morgenstern, O., Newman, P., Oman, L., Brogniez, C., & Simic, S. (2019). Success of Montreal Protocol demonstrated by comparing high-quality UV measurements with “World Avoided” calculations from two chemistry-climate models. *Scientific Reports*, 9(1), 12332. <https://doi.org/10.1038/s41598-019-48625-z>
- Barnes, P. W., Bornman, J. F., Pandey, K. K., Bernhard, G. H., Bais, A. F., Neale, R. E., Robson, T. M., Neale, P. J., Williamson, C. E., Zepp, R. G., Madronich, S., Wilson, S. R., Andrady, A. L., Heikkilä, A. M., & Robinson, S. A. (2021). The success of the Montreal Protocol in mitigating interactive effects of stratospheric ozone depletion and climate change on the environment. *Global Change Biology*, 27(22), 5681–5683. <https://doi.org/10.1111/gcb.15841>
- Young, P. J., Harper, A. B., Huntingford, C., Paul, N. D., Morgenstern, O., Newman, P. A., Oman, L. D., Madronich, S., & Garcia, R. R. (2021). The Montreal Protocol protects the terrestrial carbon sink. *Nature*, 596(7872), 384–388. <https://doi.org/10.1038/s41586-021-03737-3>
- Smith, S. L., O'Neill, H. B., Isaksen, K., Noetzli, J., & Romanovsky, V. E. (2022). The changing thermal state of permafrost. *Nature Reviews Earth & Environment*, 3(1), 10–23. <https://doi.org/10.1038/s43017-021-00240-1>
- Goyal, R., England, M. H., Sen Gupta, A., & Jucker, M. (2019). Reduction in surface climate change achieved by the 1987 Montreal Protocol. *Environmental Research Letters*, 14(12), 124041. <https://doi.org/10.1088/1748-9326/ab4874>
- Purohit, P., Borgford-Parnell, N., Klimont, Z., & Höglund-Isaksson, L. (2022). Achieving Paris climate goals calls for increasing ambition of the Kigali Amendment. *Nature Climate Change*, 12(4), 339–342. <https://doi.org/10.1038/s41558-022-01310-y>
- WMO (World Meteorological Organization), Scientific Assessment of Ozone Depletion: 2018, Global Ozone Research and Monitoring Project–Report No. 58, 588 pp., Geneva, Switzerland, 2018. <http://ozone.unep.org/science/assessment/sap>
- World Meteorological Organization (WMO). Scientific Assessment of Ozone Depletion: 2022, GAW Report No. 278, 509 pp.; WMO: Geneva, 2022. <https://ozone.unep.org/science/assessment/sap>
- Bais, A. F., Bernhard, G., McKenzie, R. L., Aucamp, P. J., Young, P. J., Ilyas, M., Jöckel, M., & Deushi, M. (2019). Ozone-climate interactions and effects on solar ultraviolet radiation. *Photochemical & Photobiological Sciences*, 18, 602–640. <https://doi.org/10.1039/C8PP90059K>
- Bernhard, G. H., Bais, A. F., Aucamp, P. J., Klekociuk, A. R., Liley, J. B., & McKenzie, R. L. (2023) Stratospheric ozone, UV radiation, and climate interactions. *Photochemical & Photobiological Sciences*. <https://doi.org/10.1007/s43630-023-00373-w>
- Wang, J. A., & Friedl, M. A. (2019). The role of land cover change in Arctic-Boreal greening and browning trends. *Environmental Research Letters*, 14(12), 125007. <https://doi.org/10.1088/1748-9326/ab5429>
- Bornman, J. F., Barnes, P. W., Robson, T. M., Robinson, S. A., Jansen, M. A. K., Ballaré, C. L., & Flint, S. D. (2019). Linkages between stratospheric ozone, UV radiation and climate change

- and their implications for terrestrial ecosystems. *Photochemical & Photobiological Sciences*, 18, 681–716. <https://doi.org/10.1039/C8PP90061B>
13. Bais, A. F., McKenzie, R. L., Bernhard, G., Aucamp, P. J., Ilyas, M., Madronich, S., & Tourpali, K. (2015). Ozone depletion and climate change: Impacts on UV radiation. *Photochemical & Photobiological Sciences*, 14(1), 19–52. <https://doi.org/10.1039/c4pp90032d>
  14. Lamy, K., Portafaix, T., Josse, B., Brogniez, C., Godin-Beekmann, S., Bencherif, H., Revell, L., Akiyoshi, H., Bekki, S., Heggin, M. I., Jöckel, P., Kirner, O., Liley, B., Marecal, V., Morgenstern, O., Stenke, A., Zeng, G., Abraham, N. L., Archibald, A. T., ... Yoshida, K. (2019). Clear-sky ultraviolet radiation modelling using output from the Chemistry Climate Model Initiative. *Atmospheric Chemistry and Physics*, 19(15), 10087–10110. <https://doi.org/10.5194/acp-19-10087-2019>
  15. Madronich, S., Sulzberger, B., Longstreth, J. D., Schikowski, T., Andersen, M. P. S., Solomon, K. R., & Wilson, S. R. (2023). Changes in tropospheric air quality related to the protection of stratospheric ozone in a changing climate. *Photochemical & Photobiological Sciences*, this issue. <https://doi.org/10.1007/s43630-023-00369-6>
  16. Pu, W., Cui, J., Wu, D., Shi, T., Chen, Y., Xing, Y., Zhou, Y., & Wang, X. (2021). Unprecedented snow darkening and melting in New Zealand due to 2019–2020 Australian wildfires. *Fundamental Research*, 1(3), 224–231. <https://doi.org/10.1016/j.fmre.2021.04.001>
  17. Damany-Pearce, L., Johnson, B., Wells, A., Osborne, M., Allan, J., Belcher, C., Jones, A., & Haywood, J. (2022). Australian wildfires cause the largest stratospheric warming since Pinatubo and extends the lifetime of the Antarctic ozone hole. *Scientific Reports*, 12(1), 12665. <https://doi.org/10.1038/s41598-022-15794-3>
  18. Bernath, P., Boone, C., & Crouse, J. (2022). Wildfire smoke destroys stratospheric ozone. *Science*, 375(6586), 1292–1295. <https://doi.org/10.1126/science.abm5611>
  19. Sulzberger, B., Austin, A. T., Cory, R. M., Zepp, R. G., & Paul, N. D. (2019). Solar UV radiation in a changing world: Roles of cryosphere-land-water-atmosphere interfaces in global biogeochemical cycles. *Photochemical & Photobiological Sciences*, 18, 747–774. <https://doi.org/10.1039/C8PP90063A>
  20. IPCC (2021). Climate Change 2021: The Physical Science Basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change. [V. Masson-Demotte, P. Zhai, A. Pirani, S.L. Connors, C.Péan, S. Berger, N. Caud, Y. Chen, L. Goldfarb, M.I. Gomis, M. Huang, K. Leitzell, E. Lonnoy, J.B.R. Matthews, T.K. Maycock, T. Waterfield, O. Yelekçi, R. Yu and B. Zhou (eds.)], (pp. In Press). <https://doi.org/10.1017/9781009157896>
  21. IPCC (2022). Climate change 2022: Impacts, adaptation, and vulnerability. Contribution of Working Group II to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change. [D. C. R. H.-O. Pörtner, M. Tignor, E.S. Poloczanska, K. Mintenbeck, A. Alegría, M. Craig, S. Langsdorf, S. Löschke, V. Möller, A. Okem, R. Rama (eds.)], (pp. In Press). Cambridge University Press, Cambridge, UK and New York, NY, USA, 3056 pp., <https://doi.org/10.1017/9781009325844>
  22. Crutzen, P. J. (2006). Albedo enhancement by stratospheric sulfur injections: A contribution to resolve a policy dilemma? *Climatic Change*, 77(3–4), 211–220. <https://doi.org/10.1007/s10584-006-9101-y>
  23. Kravitz, B., & MacMartin, D. G. (2020). Uncertainty and the basis for confidence in solar geoengineering research. *Nature Reviews Earth & Environment*, 1(1), 64–75. <https://doi.org/10.1038/s43017-019-0004-7>
  24. Madronich, S., Tilmes, S., Kravitz, B., MacMartin, D., & Richter, J. (2018). Response of surface ultraviolet and visible radiation to stratospheric SO<sub>2</sub> injections. *Atmosphere*. <https://doi.org/10.3390/atmos9110432>
  25. Tilmes, S., Müller, R., & Salawitch, R. (2008). The sensitivity of Polar ozone depletion to proposed geoengineering schemes. *Science*, 320(5880), 1201. <https://doi.org/10.1126/science.1153966>
  26. Trisos, C. H., Amatulli, G., Gurevitch, J., Robock, A., Xia, L., & Zambri, B. (2018). Potentially dangerous consequences for biodiversity of solar geoengineering implementation and termination. *Nature Ecology & Evolution*, 2(3), 475–482. <https://doi.org/10.1038/s41559-017-0431-0>
  27. Lockley, A., Xu, Y., Tilmes, S., Sugiyama, M., Rothman, D., & Hinde, A. (2022). 18 Politically relevant solar geoengineering scenarios. *Socio-Environmental Systems Modelling*. <https://doi.org/10.18174/sesmo.18127>
  28. Zarnetske, P. L., Gurevitch, J., Franklin, J., Groffman, P. M., Harrison, C. S., Hellmann, J. J., Hoffman, F. M., Kothari, S., Robock, A., Tilmes, S., Visioni, D., Wu, J., Xia, L., & Yang, C. E. (2021). Potential ecological impacts of climate intervention by reflecting sunlight to cool Earth. *Proceedings of the National Academy of Sciences USA*. <https://doi.org/10.1073/pnas.1921854118>
  29. Barnes, P. W., Robson, T. M., Neale, P. J., Williamson, C. E., Zepp, R. G., Madronich, S., Wilson, S. R., Andrad, A. L., Heikkilä, A. M., Bernhard, G. H., Bais, A. F., Neale, R. E., Bornman, J. F., Jansen, M. A. K., Klekociuk, A. R., Martinez-Abaigar, J., Robinson, S. A., Wang, Q. W., Banaszak, A. T., ... Young, A. R. (2022). Environmental effects of stratospheric ozone depletion, UV radiation, and interactions with climate change: UNEP Environmental Effects Assessment Panel, Update 2021. *Photochemical & Photobiological Sciences*. <https://doi.org/10.1007/s43630-022-00176-5>
  30. Friedel, M., Chiodo, G., Stenke, A., Domeisen, D., Fueglistaler, S., Anet, J., & Peter, T. (2022). Springtime arctic ozone depletion forces northern hemisphere climate anomalies. *Nature Geosciences*, 15, 541–547. <https://doi.org/10.1038/s41561-022-00974-7>
  31. Kwon, H., Choi, H., Kim, B.-M., Kim, S.-W., & Kim, S.-J. (2020). Recent weakening of the southern stratospheric polar vortex and its impact on the surface climate over Antarctica. *Environmental Research Letters*. <https://doi.org/10.1088/1748-9326/ab9d3d>
  32. Jucker, M., & Goyal, R. (2021). Ozone-forced southern annular mode during Antarctic stratospheric warming events. *Geophysical Research Letters*. <https://doi.org/10.1002/essoar.10507626.1>
  33. Xia, Y., Hu, Y., Huang, Y., Zhao, C., Xie, F., & Yang, Y. (2021). Significant contribution of severe ozone loss to the Siberian-Arctic surface warming in spring 2020. *Geophysical Research Letters*. <https://doi.org/10.1029/2021GL092509>
  34. Robinson, S. A., & Erickson, D. J., III. (2015). Not just about sunburn—the ozone hole’s profound effect on climate has significant implications for Southern Hemisphere ecosystems. *Global Change Biology*, 21(2), 515–527. <https://doi.org/10.1111/gcb.12739>
  35. Bernhard, G. H., McKenzie, R. L., Lantz, K., & Stierle, S. (2022). Updated analysis of data from Palmer Station, Antarctica (64° S), and San Diego, California (32° N), confirms large effect of the Antarctic ozone hole on UV radiation. *Photochemical & Photobiological Sciences*, 21(3), 373–384. <https://doi.org/10.1007/s43630-022-00178-3>
  36. Robinson, S. A., Wasley, J., & Tobin, A. K. (2003). Living on the edge - plants and global change in continental and maritime



- Antarctica. *Global Change Biology*, 9(12), 1681–1717. <https://doi.org/10.1046/j.1365-2486.2003.00693.x>
37. Newsham, K. K., & Robinson, S. A. (2009). Responses of plants in polar regions to UVB exposure: A meta-analysis. *Global Change Biology*, 15(11), 2574–2589. <https://doi.org/10.1111/j.1365-2486.2009.01944.x>
  38. Ballaré, C. L., Caldwell, M. M., Flint, S. D., Robinson, S. A., & Bornman, J. F. (2011). Effects of solar ultraviolet radiation on terrestrial ecosystems Patterns., mechanisms, and interactions with climate change. *Photochemical & Photobiological Sciences*. <https://doi.org/10.1039/c0pp90035d>
  39. Banerjee, A., Fyfe, J. C., Polvani, L. M., Waugh, D., & Chang, K.-L. (2020). A pause in Southern Hemisphere circulation trends due to the Montreal Protocol. *Nature*, 579(7800), 544–548. <https://doi.org/10.1038/s41586-020-2120-4>
  40. Fogt, R. L., & Marshall, G. J. (2020). The Southern Annular Mode: Variability, trends, and climate impacts across the Southern Hemisphere. *WIREs Climate Change*, 11(4), e652. <https://doi.org/10.1002/wcc.652>
  41. Gillett, Z. E., Arblaster, J. M., Dittus, A. J., Deushi, M., Jöckel, P., Kinnison, D. E., Morgenstern, O., Plummer, D. A., Revell, L. E., Rozanov, E., Schofield, R., Stenke, A., Stone, K. A., & Tilmes, S. (2019). Evaluating the relationship between interannual variations in the Antarctic ozone hole and Southern Hemisphere surface climate in chemistry–climate models. *Journal of Climate*, 32(11), 3131–3151. <https://doi.org/10.1175/jcli-d-18-0273.1>
  42. Morales, M. S., Cook, E. R., Barichivich, J., Christie, D. A., Villalba, R., LeQuesne, C., Srur, A. M., Ferrero, M. E., Gonzalez-Reyes, A., Couvreur, F., Matuskovsky, V., Aravena, J. C., Lara, A., Mundo, I. A., Rojas, F., Prieto, M. R., Smerdon, J. E., Bianchi, L. O., Masiokas, M. H., ... Boninsegna, J. A. (2020). Six hundred years of South American tree rings reveal an increase in severe hydroclimatic events since mid-20th century. *Proceedings of the National Academy of Sciences USA*, 117(29), 16816–16823. <https://doi.org/10.1073/pnas.2002411117>
  43. Damiani, A., Cordero, R. R., Llanillo, P. J., Feron, S., Boisier, J. P., Garreaud, R., Rondanelli, R., Irie, H., & Watanabe, S. (2020). Connection between Antarctic Ozone and climate: Interannual precipitation changes in the Southern Hemisphere. *Atmosphere*, 11(6), 579. <https://doi.org/10.3390/atmos11060579>
  44. Doddridge, E. W., Marshall, J., Song, H., Campin, J.-M., & Kelley, M. (2021). Southern Ocean heat storage, reemergence, and winter sea ice decline induced by summertime winds. *Journal of Climate*, 34(4), 1403–1415. <https://doi.org/10.1175/jcli-d-20-0322.1>
  45. England, M., Polvani, L., & Sun, L. (2018). Contrasting the Antarctic and Arctic atmospheric responses to projected sea ice loss in the late twenty-first century. *Journal of Climate*, 31(16), 6353–6370. <https://doi.org/10.1175/jcli-d-17-0666.1>
  46. Li, S., Liu, W., Lyu, K., & Zhang, X. (2021). The effects of historical ozone changes on Southern Ocean heat uptake and storage. *Climate Dynamics*, 57(7–8), 2269–2285. <https://doi.org/10.1007/s00382-021-05803-y>
  47. Marshall, J., Ferreira, D., Bitz, C. M., Solomon, S., & Plumb, A. (2015). Antarctic Ocean and sea ice response to ozone depletion: A two-time-scale problem. *Journal of Climate*, 28(3), 1206–1226. <https://doi.org/10.1175/jcli-d-14-00313.1>
  48. Son, S.-W., Han, B.-R., Garfinkel, C. I., Kim, S.-Y., Park, R., Abraham, N. L., Akiyoshi, H., Archibald, A. T., Butchart, N., Chipperfield, M. P., Dameris, M., Deushi, M., Dhomse, S. S., Hardiman, S. C., Jöckel, P., Kinnison, D., Michou, M., Morgenstern, O., O'Connor, F. M., ... Zeng, G. (2018). Tropospheric jet response to Antarctic ozone depletion: An update with Chemistry–Climate Model Initiative (CCMI) models. *Environmental Research Letters*, 13(5), 054024. <https://doi.org/10.1088/1748-9326/aabf21>
  49. Waugh, D. W., Thomas, J. L., Polvani, L. M., Marshall, J., Kostov, Y., Kelley, M., Gnanadesikan, A., Ferreira, D., Doddridge, E. W., Codron, F., & Seviour, W. J. M. (2019). The Southern Ocean sea surface temperature response to ozone depletion: A multimodel comparison. *Journal of Climate*, 32(16), 5107–5121. <https://doi.org/10.1175/jcli-d-19-0109.1>
  50. Polvani, L. M., Banerjee, A., Chemke, R., Doddridge, E. W., Ferreira, D., Gnanadesikan, A., Holland, M. A., Kostov, Y., Marshall, J., Seviour, W. J. M., Solomon, S., & Waugh, D. W. (2021). Interannual SAM modulation of Antarctic sea ice extent does not account for its long-term trends, pointing to a limited role for ozone depletion. *Geophysical Research Letters*. <https://doi.org/10.1029/2021gl094871>
  51. Stuecker, M. F., Bitz, C. M., Armour, K. C., Proistosescu, C., Kang, S. M., Xie, S.-P., Kim, D., McGregor, S., Zhang, W., Zhao, S., Cai, W., Dong, Y., & Jin, F.-F. (2018). Polar amplification dominated by local forcing and feedbacks. *Nature Climate Change*, 8(12), 1076–1081. <https://doi.org/10.1038/s41558-018-0339-y>
  52. Bergstrom, D. M., Dickson, C. R., Baker, D. J., Winham, J., Selkirk, P. M., & McGeoch, M. A. (2021). Ecosystem collapse on a Sub-Antarctic island. In J. G. Canadell & R. B. Jackson (Eds.), *Ecosystem Collapse and Climate Change* (pp. 13–25). Springer International Publishing.
  53. Robinson, S. A., King, D. H., Bramley-Alves, J., Waterman, M. J., Ashcroft, M. B., Wasley, J., Turnbull, J. D., Miller, R. E., Ryan-Colton, E., Benny, T., Mullany, K., Clarke, L. J., Barry, L. A., & Hua, Q. (2018). Rapid change in East Antarctic terrestrial vegetation in response to regional drying. *Nature Climate Change*, 8(10), 879–884. <https://doi.org/10.1038/s41558-018-0280-0>
  54. Kramarova, N., Newman, P. A., Nash, E. R., Strahan, S. E., Long, C. S., Johnson, B., Pitts, M., Santee, M. L., Petropavlovskikh, I., Coy, L., & de Laat, J. (2020). 2019 Antarctic ozone hole In J. Blunden, & D. S. Arndt (Eds.), “State of the Climate in 2019”, *Bulletin of the American Meteorological Society* (Vol. 101, pp. S310–S312)
  55. Milinevsky, G., Evtushevsky, O., Klekociuk, A., Wang, Y., Grytsai, A., Shulga, V., & Ivaniha, O. (2019). Early indications of anomalous behaviour in the 2019 spring ozone hole over Antarctica. *International Journal of Remote Sensing*, 41(19), 7530–7540. <https://doi.org/10.1080/2150704X.2020.1763497>
  56. Shen, X., Wang, L., & Osprey, S. (2020). Tropospheric forcing of the 2019 Antarctic sudden stratospheric warming. *Geophysical Research Letters*. <https://doi.org/10.1029/2020gl089343>
  57. Hendon, H. H., Thompson, D. J. W., Lim, E.-P., Butler, A. H., Newman, P. A., Coy, L., Scaife, A., Polichtchouk, I., Garreaud, R. S., & T.G., S., & Nakamura, H. (2019). Rare forecasted climate event under way in the Southern Hemisphere. *Nature*, 573(7775), 495. <https://doi.org/10.1038/d41586-019-02858-0>
  58. Jucker, M., Reichler, T., & Waugh, D. W. (2021). How frequent are Antarctic sudden stratospheric warmings in present and future climate? *Geophysical Research Letters*. <https://doi.org/10.1029/2021gl093215>
  59. Lim, E.-P., Hendon, H. H., Boschat, G., Hudson, D., Thompson, D. W. J., Dowdy, A. J., & Arblaster, J. M. (2019). Australian hot and dry extremes induced by weakenings of the stratospheric polar vortex. *Nature Geoscience*. <https://doi.org/10.1038/s41561-019-0456-x>
  60. Lim, E.-P., Hendon, H. H., Butler, A. H., Garreaud, R. D., Polichtchouk, I., Shepherd, T. G., Scaife, A., Comer, R., Coy, L., Newman, P. A., Thompson, D. J. W., & Nakamura, H. (2020). The 2019 Antarctic sudden stratospheric warming. *SPARC Newsletter*, 54, 10–13.



61. Newman, P., Nash, E. R., Kramarova, N., & Butler, A. (2020). The 2019 southern stratospheric warming. In: T. Scambos, & S. Stammerjohn (Eds.) "State of the Climate in 2019", *Bulletin of the American Meteorological Society* (Vol. 101, pp. S297–S298, Vol. 8).
62. Noguchi, S., Kuroda, Y., Kodera, K., & Watanabe, S. (2020). Robust enhancement of tropical convective activity by the 2019 Antarctic sudden stratospheric warming. *Geophysical Research Letters*. <https://doi.org/10.1029/2020GL088743>
63. Robinson, S. A., Klekociuk, A. R., King, D. H., Pizarro Rojas, M., Zúñiga, G. E., & Bergstrom, D. M. (2020). The 2019/2020 summer of Antarctic heatwaves. *Global Change Biology*, 26(6), 3178–3180. <https://doi.org/10.1111/gcb.15083>
64. Yamazaki, Y., Matthias, V., Miyoshi, Y., Stolle, C., Siddiqui, T., Kervlishvili, G., Laštovička, J., Kozubek, M., Ward, W., Themens, D. R., Kristoffersen, S., & Alken, P. (2020). September 2019 Antarctic sudden stratospheric warming: quasi-6-Day wave burst and ionospheric effects. *Geophysical Research Letters*. <https://doi.org/10.1029/2019gl086577>
65. Allen, D. R., Fromm, M. D., Kablick, G. P., III., & Nedoluha, G. E. (2020). Smoke with induced rotation and lofting (SWIRL) in the stratosphere. *Journal of the Atmospheric Sciences*, 77(12), 4297–4316. <https://doi.org/10.1175/JAS-D-20-0131.1>
66. Boone, C. D., Bernath, P. F., & Fromm, M. D. (2020). Pyrocumulonimbus stratospheric plume injections measured by the ACE-FTS. *Geophysical Research Letters*. <https://doi.org/10.1029/2020GL088442>
67. Hirsch, E., & Koren, I. (2021). Record-breaking aerosol levels explained by smoke injection into the stratosphere. *Science*, 371(6535), 1269–1274. <https://doi.org/10.1126/science.abe1415>
68. Kablick, G. P., III., Allen, D. R., Fromm, M. D., & Nedoluha, G. E. (2020). Australian PyroCb smoke generates synoptic-scale stratospheric anticyclones. *Geophysical Research Letters*. <https://doi.org/10.1029/2020GL088101>
69. Khaykin, S., Legras, B., Bucci, S., Sellitto, P., Isaksen, I., Tencé, F., Bekki, S., Bourassa, A., Rieger, L., Zawada, D., Jumelet, J., & Godin-Beekmann, S. (2020). The 2019/20 Australian wildfires generated a persistent smoke-charged vortex rising up to 35 km altitude. *Communications Earth & Environment*, 1(1), 1–12. <https://doi.org/10.1038/s43247-020-00022-5>
70. Ohnaiser, K., Ansmann, A., Baars, H., Seifert, P., Barja, B., Jimenez, C., Radenz, M., Teisseire, A., Floutsis, A., Haarig, M., Foth, A., Chudnovsky, A., Engelmann, R., Zamorano, F., Bühl, J., & Wandinger, U. (2020). Smoke of extreme Australian bushfires observed in the stratosphere over Punta Arenas, Chile, in January 2020: Optical thickness, lidar ratios, and depolarization ratios at 355 and 532 nm. *Atmospheric Chemistry and Physics*, 20(13), 8003–8015. <https://doi.org/10.5194/acp-20-8003-2020>
71. Schwartz, M. J., Santee, M. L., Pumphrey, H. C., Manney, G. L., Lambert, A., Livesey, N. J., Millán, L., Neu, J. L., Read, W. G., & Werner, F. (2020). Australian new year's pyroCb impact on stratospheric composition. *Geophysical Research Letters*. <https://doi.org/10.1029/2020GL090831>
72. Solomon, S., Dube, K., Stone, K., Yu, P., Kinnison, D., Toon, O. B., Strahan, S. E., Rosenlof, K. H., Portmann, R., Davis, S., Randel, W., Bernath, P., Boone, C., Bardeen, C. G., Bourassa, A., Zawada, D., & Degenstein, D. (2022). On the stratospheric chemistry of midlatitude wildfire smoke. *Proceedings of the National Academy of Sciences USA*. <https://doi.org/10.1073/pnas.2117325119>
73. Yu, P., Davis, S. M., Toon, O. B., Portmann, R. W., Bardeen, C. G., Barnes, J. E., Telg, H., Maloney, C., & Rosenlof, K. H. (2021). Persistent stratospheric warming due to 2019–2020 Australian wildfire smoke. *Geophysical Research Letters*. <https://doi.org/10.1029/2021GL092609>
74. Klekociuk, A. R., Tully, M. B., Krummel, P. B., Henderson, S. I., Smale, D., Querel, R., Nichol, S., Alexander, S. P., Fraser, P. J., & Nedoluha, G. (2022). The Antarctic ozone hole during 2020. *Journal of Southern Hemisphere Earth Systems Science*. <https://doi.org/10.1071/es21015>
75. Stone, K. A., Solomon, S., Kinnison, D. E., & Mills, M. J. (2021). On recent large Antarctic ozone holes and ozone recovery metrics. *Geophysical Research Letters*. <https://doi.org/10.1029/2021gl095232>
76. Yook, S., Thompson, D. W. J., & Solomon, S. (2022). Climate impacts and potential drivers of the unprecedented Antarctic ozone holes of 2020 and 2021. *Geophysical Research Letters*. <https://doi.org/10.1029/2022GL098064>
77. Lecouffe, A., Godin-Beekmann, S., Pazmiño, A., & Hauchecorne, A. (2022). Evolution of the intensity and duration of the Southern Hemisphere stratospheric polar vortex edge for the period 1979–2020. *Atmospheric Chemistry and Physics*, 22(6), 4187–4200. <https://doi.org/10.5194/acp-22-4187-2022>
78. Ceppi, P., & Shepherd, T. G. (2019). The role of the stratospheric polar vortex for the austral jet response to greenhouse gas forcing. *Geophysical Research Letters*, 46(12), 6972–6979. <https://doi.org/10.1029/2019gl082883>
79. Lee, J. R., Waterman, M. J., Shaw, J. D., Bergstrom, D. M., Lynch, H. J., Wall, D. H., & Robinson, S. A. (2022). Islands in the ice: Potential impacts of habitat transformation on Antarctic biodiversity. *Global Change Biology*. <https://doi.org/10.1111/gcb.16331>
80. Lembrechts, J. J., van den Hoogen, J., Aalto, J., Ashcroft, M. B., De Frenne, P., Kemppinen, J., Kopecky, M., Luoto, M., Maclean, I. M. D., Crowther, T. W., Bailey, J. J., Haesen, S., Klimes, D. H., Niittynen, P., Scheffers, B. R., Van Meerbeek, K., Aartsma, P., Abdalaze, O., Abedi, M., ... Lenoir, J. (2022) Global maps of soil temperature. *Global Change Biology*. Doi: <https://doi.org/10.1111/gcb.16060>.
81. Cannone, N., Malfasi, F., Favero-Longo, S. E., Convey, P., & Guglielmin, M. (2022). Acceleration of climate warming and plant dynamics in Antarctica. *Current Biology*. <https://doi.org/10.1016/j.cub.2022.01.074>
82. Chown, S. L., Leihy, R. I., Naish, T. R., Brooks, C. M., Convey, P., Henley, B. J., Mackintosh, A. N., Phillips, L. M., Kennicutt, M. C., II., & Grant, S. M. (Eds.). (2022). *Antarctic climate change and the environment: A decadal synopsis and recommendations for action*. Scientific Committee on Antarctic Research.
83. Myers-Smith, I. H., Kerby, J. T., Phoenix, G. K., Bjerke, J. W., Epstein, H. E., Assmann, J. J., John, C., Andreu-Hayles, L., Angers-Blondin, S., Beck, P. S. A., Berner, L. T., Bhatt, U. S., Bjorkman, A. D., Blok, D., Bryn, A., Christiansen, C. T., Cornelissen, J. H. C., Cunliffe, A. M., Elmendorf, S. C., ... Wipf, S. (2020). Complexity revealed in the greening of the Arctic. *Nature Climate Change*, 10(2), 106–117. <https://doi.org/10.1038/s41558-019-0688-1>
84. Callaghan, T. V., Cazzolla Gatti, R., & Phoenix, G. (2022). The need to understand the stability of arctic vegetation during rapid climate change: An assessment of imbalance in the literature. *Ambio*, 51(4), 1034–1044. <https://doi.org/10.1007/s13280-021-01607-w>
85. Phoenix, G. K., & Treharne, R. (2022). Arctic greening and browning: Challenges and a cascade of complexities. *Global Change Biology*, 28(11), 3481–3483. <https://doi.org/10.1111/gcb.16118>
86. Treharne, R., Bjerke, J. W., Tommervik, H., Stenard, L., & Phoenix, G. K. (2019). Arctic browning: Impacts of extreme climatic events on heathland ecosystem CO<sub>2</sub> fluxes. *Global Change Biology*, 25(2), 489–503. <https://doi.org/10.1111/gcb.14500>

87. Robinson, S. A. (2022). Climate change and extreme events are changing the biology of Polar Regions. *Global Change Biology*. <https://doi.org/10.1111/gcb.16309>
88. Bokhorst, S., Cornelissen, J. H. C., & Veraverbeke, S. (2022). Long-term legacies of seasonal extremes in Arctic ecosystem functioning. *Global Change Biology*, 28(10), 3161–3162. <https://doi.org/10.1111/gcb.16078>
89. Perera-Castro, A. V., Waterman, M. J., Turnbull, J. D., Ashcroft, M. B., McKinley, E., Watling, J. R., Bramley-Alves, J., Casanova-Katny, A., Zuniga, G., Flexas, J., & Robinson, S. A. (2020). It is hot in the sun: Antarctic mosses have high temperature optima for photosynthesis despite cold climate. *Frontiers in Plant Science*, 11, 1178. <https://doi.org/10.3389/fpls.2020.01178>
90. Lawrence, Z. D., Perlwitz, J., Butler, A. H., Manney, G. L., Newman, P. A., Lee, S. H., & Nash, E. R. (2020). The remarkably strong Arctic stratospheric polar vortex of winter Links to record-breaking Arctic oscillation and ozone loss. *Journal of Geophysical Research Atmospheres*. <https://doi.org/10.1029/2020JD033271>
91. Manney, G. L., Livesey, N. J., Santee, M. L., Froidevaux, L., Lambert, A., Lawrence, Z. D., Millán, L. F., Neu, J. L., Read, W. G., Schwartz, M. J., & Fuller, R. A. (2020). Record-low Arctic stratospheric ozone in 2020: MLS observations of chemical processes and comparisons with previous extreme winters. *Geophysical Research Letters*. <https://doi.org/10.1029/2020gl089063>
92. Bernhard, G. H., Fioletov, V. E., Grooß, J.-U., Ialongo, I., Johnsen, B., Lakkala, K., Manney, G. L., Müller, R., & Svendby, T. (2021). Ozone and ultraviolet radiation. In: J. Blunden, & T. Boyer (Eds.). Vol. 102 “State of the Climate in 2020”, *Bull. Amer. Meteor. Soc.* (, pp. S299–S303).
93. Bernhard, G. H., Fioletov, V. E., Grooß, J. U., Ialongo, I., Johnsen, B., Lakkala, K., Manney, G. L., Müller, R., & Svendby, T. (2020). Record-breaking increases in Arctic solar ultraviolet radiation caused by exceptionally large ozone depletion in 2020. *Geophysical Research Letters*. <https://doi.org/10.1029/2020gl090844>
94. Domeisen, D. I. V., & Butler, A. H. (2020). Stratospheric drivers of extreme events at the Earth’s surface. *Communications Earth & Environment*. <https://doi.org/10.1038/s43247-020-00060-z>
95. Overland, J. E., & Wang, M. (2021). The 2020 Siberian heat wave. *International Journal of Climatology*, 41, E2341–E2346. <https://doi.org/10.1002/joc.6850>
96. Zhang, J., Tian, W., Pyle, J. A., Keeble, J., Abraham, N. L., Chipperfield, M. P., Xie, F., Yang, Q., Mu, L., Ren, H.-L., Wang, L., & Xu, M. (2022). Responses of Arctic sea ice to stratospheric ozone depletion. *Science Bulletin*, 67(11), 1182–1190. <https://doi.org/10.1016/j.scib.2022.03.015>
97. Abram, N. J., Henley, B. J., Sen Gupta, A., Lippmann, T. J. R., Clarke, H., Dowdy, A. J., Sharples, J. J., Nolan, R. H., Zhang, T., Wooster, M. J., Wurtzel, J. B., Meissner, K. J., Pitman, A. J., Ukkola, A. M., Murphy, B. P., Tapper, N. J., & Boer, M. M. (2021). Connections of climate change and variability to large and extreme forest fires in southeast Australia. *Communications Earth & Environment*. <https://doi.org/10.1038/s43247-020-00065-8>
98. Zhang, Y., Beggs, P. J., McGushin, A., Bambrick, H., Trueck, S., Hanigan, I. C., Morgan, G. G., Berry, H. L., Linnenluecke, M. K., Johnston, F. H., Capon, A. G., & Watts, N. (2020). The 2020 special report of the MJA–Lancet Countdown on health and climate change: Lessons learnt from Australia’s “Black Summer.” *Medical Journal of Australia*, 213(11), 490–492.e410. <https://doi.org/10.5694/mja2.50869>
99. Graham, A. M., Pringle, K. J., Pope, R. J., Arnold, S. R., Conibear, L. A., Burns, H., Rigby, R., Borchers-Arriagada, N., Butt, E. W., Kiely, L., Reddington, C., Spracklen, D. V., Woodhouse, M. T., Knote, C., & McQuaid, J. B. (2021). Impact of the Australian an megafires on air quality and health. *Geohealth*. <https://doi.org/10.1029/2021GH000454>
100. Cowled, B. D., Hillman, A., Ward, M. P., Clutterbuck, H., Doyle, M., Webb Ware, J., Thomas, M., Plain, K., Barwell, R., Laurence, M., & Pfeiffer, C. (2022). The black summer bushfires: Impacts and risk factors for livestock bushfire injury in south-eastern Australia. *Australian Veterinary Journal*, 100(7), 306–317. <https://doi.org/10.1111/avj.13165>
101. Dickman, C. R. (2021). Ecological consequences of Australia’s “Black Summer” bushfires: Managing for recovery. *Integrated Environmental Assessment and Management*, 17(6), 1162–1167. <https://doi.org/10.1002/ieam.4496>
102. González-Herrero, S., Barriopedro, D., Trigo, R. M., López-Bustins, J. A., & Oliva, M. (2022). Climate warming amplified the 2020 record-breaking heatwave in the Antarctic Peninsula. *Communications Earth & Environment*. <https://doi.org/10.1038/s43247-022-00450-5>
103. Bergstrom, D. M., Wienecke, B. C., van den Hoff, J., Hughes, L., Lindenmayer, D. B., Ainsworth, T. D., Baker, C. M., Bland, L., Bowman, D. M. J. S., Brooks, S. T., Canadell, J. G., Constable, A. J., Dafforn, K. A., Depledge, M. H., Dickson, C. R., Duke, N. C., Helmstedt, K. J., Holz, A., Johnson, C. R., ... Shaw, J. D. (2021). Combating ecosystem collapse from the tropics to the Antarctic. *Global Change Biology*, 27(9), 1692–1703. <https://doi.org/10.1111/gcb.15539>
104. Cortes-Antiquera, R., Pizarro, M., Contreras, R. A., Kohler, H., & Zuniga, G. E. (2021). Heat shock tolerance in *Deschampsia antarctica* desv. cultivated in vitro is mediated by enzymatic and non-enzymatic antioxidants. *Frontiers in Plant Science*, 12, 635491. <https://doi.org/10.3389/fpls.2021.635491>
105. Montie, S., Thomsen, M. S., Rack, W., & Broady, P. A. (2020). Extreme summer marine heatwaves increase chlorophyll a in the Southern Ocean. *Antarctic Science*, 32(6), 508–509. <https://doi.org/10.1017/S0954102020000401>
106. Samuels, T., Rynearson, T. A., & Collins, S. (2021). Surviving heatwaves: Thermal experience predicts life and death in a southern ocean diatom. *Frontiers in Marine Science*. <https://doi.org/10.3389/fmars.2021.600343>
107. De Leij, R., Grange, L. J., & Peck, L. S. (2022). Functional thermal limits are determined by rate of warming during simulated marine heatwaves. *Marine Ecology Progress Series*, 685, 183–196. <https://doi.org/10.3354/meps13980>
108. Bernhard, P., Zwieback, S., & Hajnsek, I. (2022). Accelerated mobilization of organic carbon from retrogressive thaw slumps on the northern Taymyr Peninsula. *The Cryosphere*, 16(7), 2819–2835. <https://doi.org/10.5194/tc-16-2819-2022>
109. Boonstra, R., Bodner, K., Bosson, C., Delehanty, B., Richardson, E. S., Lunn, N. J., Derocher, A. E., & Molnár, P. K. (2020). The stress of Arctic warming on polar bears. *Global Change Biology*, 26(8), 4197–4214. <https://doi.org/10.1111/gcb.15142>
110. Stirling, I., & Derocher, A. E. (2012). Effects of climate warming on polar bears: A review of the evidence. *Global Change Biology*, 18(9), 2694–2706. <https://doi.org/10.1111/j.1365-2486.2012.02753.x>
111. Neale, P. J., Williamson, C. E., Banaszak, A. T., Häder, D.-P., Hylander, S., Ossola, R., Rose, K. A., Wängberg, S. Å., & Zepp, R. G. (2023). The response of aquatic ecosystems to the interactive effects of stratospheric ozone depletion, UV radiation, and climate change. *Photochemical & Photobiological Sciences*. <https://doi.org/10.1007/s43630-023-00370-z>
112. Smith, M. D. (2011). The ecological role of climate extremes: Current understanding and future prospects. *Journal of Ecology*, 99(3), 651–655. <https://doi.org/10.1111/j.1365-2745.2011.01833.x>
113. Chen, Y., Liao, Z., Shi, Y., Tian, Y., & Zhai, P. (2021). Detectable increases in sequential flood-heatwave events across China

- during 1961–2018. *Geophysical Research Letters*. <https://doi.org/10.1029/2021GL092549>
114. Fischer, E. M., Sippel, S., & Knutti, R. (2021). Increasing probability of record-shattering climate extremes. *Nature Climate Change*, 11(8), 689–695. <https://doi.org/10.1038/s41558-021-01092-9>
  115. Goss, M., Swain, D. L., Abatzoglou, J. T., Sarhadi, A., Kolden, C. A., Williams, A. P., & Diffenbaugh, N. S. (2020). Climate change is increasing the likelihood of extreme autumn wildfire conditions across California. *Environmental Research Letters*, 15(9), 094016. <https://doi.org/10.1088/1748-9326/ab83a7>
  116. Kennard, D. K., Matlaga, D., Sharpe, J., King, C., Alonso-Rodríguez, A. M., Reed, S. C., Cavaleri, M. A., & Wood, T. E. (2020). Tropical understory herbaceous community responds more strongly to hurricane disturbance than to experimental warming. *Ecology and Evolution*, 10(16), 8906–8915. <https://doi.org/10.1002/ece3.6589>
  117. Li, F., Wan, X., Wang, H., Orsolini, Y. J., Cong, Z., Gao, Y., & Kang, S. (2020). Arctic sea-ice loss intensifies aerosol transport to the Tibetan Plateau. *Nature Climate Change*, 10(11), 1037–1044. <https://doi.org/10.1038/s41558-020-0881-2>
  118. Musselman, K. N., Addor, N., Vano, J. A., & Molotch, N. P. (2021). Winter melt trends portend widespread declines in snow water resources. *Nature Climate Change*. <https://doi.org/10.1038/s41558-021-01014-9>
  119. Dodd, R. J., Chadwick, D. R., Harris, I. M., Hines, A., Hollis, D., Economou, T., Gwynn-Jones, D., Scullion, J., Robinson, D. A., & Jones, D. L. (2021). Spatial co-localisation of extreme weather events: A clear and present danger. *Ecology Letters*, 24(1), 60–72. <https://doi.org/10.1111/ele.13620>
  120. Filazzola, A., Matter, S. F., & MacIvor, J. S. (2021). The direct and indirect effects of extreme climate events on insects. *Science of the Total Environment*, 769, 145161. <https://doi.org/10.1016/j.scitotenv.2021.145161>
  121. IPCC. (2019). Summary for Policymakers. In J. S. P. R. Shukla, E. CalvoBuendia, V. MassonDelmotte, H. O. Pörtner, D. C. Roberts, P. Zhai, R. Slade, S. Connors, R. van Diemen, M. Ferrat, E. Haughey, S. Luz, S. Neogi, M. Pathak, J. Petzold, J. Portugal Pereira, P. Vyas, E. Huntley, K. Kissick, ... J. Malley (Eds.), *Climate Change and Land an IPCC special report on climate change desertification land degradation sustainable land management food security and greenhouse gas fluxes in terrestrial ecosystems*. IPCC.
  122. Ruthrof, K. X., Fontaine, J. B., Breshears, D. D., Field, J. P., & Allen, C. D. (2021) Extreme events trigger terrestrial and marine ecosystem collapses in the Southwestern USA and Southwestern Australia. In *Ecosystem Collapse and Climate Change* (pp. 187–217, Ecological Studies). [https://doi.org/10.1007/978-3-030-71330-0\\_8](https://doi.org/10.1007/978-3-030-71330-0_8)
  123. Walsh, J. E., Ballinger, T. J., Euskirchen, E. S., Hanna, E., Mård, J., Overland, J. E., Tangen, H., & Vihma, T. (2020). Extreme weather and climate events in northern areas: A review. *Earth-Science Reviews*. <https://doi.org/10.1016/j.earscirev.2020.103324>
  124. Neale, R. E., Barnes, P. W., Robson, T. M., Neale, P. J., Williamson, C. E., Zepp, R. G., Wilson, S. R., Madronich, S., Andrad, A. L., Heikkilä, A. M., Bernhard, G. H., Bais, A. F., Aucamp, P. J., Banaszak, A. T., Bornman, J. F., Bruckman, L. S., Byrne, S. N., Foereid, B., Häder, D. P., ... Zhu, M. (2021). Environmental effects of stratospheric ozone depletion, UV radiation, and interactions with climate change: UNEP Environmental Effects Assessment Panel, Update 2020. *Photochemical & Photobiological Sciences*, 20, 1–67. <https://doi.org/10.1007/s43630-020-00001-x>
  125. Silva, C. A., Santilli, G., Sano, E. E., & Laneve, G. (2021). Fire occurrences and greenhouse gas emissions from deforestation in the Brazilian Amazon. *Remote Sensing*. <https://doi.org/10.3390/rs13030376>
  126. Shiraishi, T., & Hirata, R. (2021). Estimation of carbon dioxide emissions from the megafires of Australia in 2019–2020. *Scientific Reports*, 11(1), 8267. <https://doi.org/10.1038/s41598-021-87721-x>
  127. Leitold, V., Morton, D. C., Martinuzzi, S., Paynter, I., Uriarte, M., Keller, M., Ferraz, A., Cook, B. D., Corp, L. A., & González, G. (2022). Tracking the rates and mechanisms of canopy damage and recovery following hurricane maria using multitemporal lidar data. *Ecosystems*, 25(4), 892–910. <https://doi.org/10.1007/s10021-021-00688-8>
  128. Wang, J. A., Randerson, J. T., Goulden, M. L., Knight, C. A., & Battles, J. J. (2022). Losses of tree cover in California driven by increasing fire disturbance and climate stress. *AGU Advances*. <https://doi.org/10.1029/2021av000654>
  129. Messier, C., Parent, S., & Bergeron, Y. (1998). Effects of overstory and understory vegetation on the understory light environment in mixed boreal forests. *Journal of Vegetation Science*, 9(4), 511–520. <https://doi.org/10.2307/3237266>
  130. Tanner, E. V., Kapos, V., & Healey, J. (1991). Hurricane effects on forest ecosystems in the Caribbean. *Biotropica*, 513–521. <https://www.jstor.org/stable/2388274>
  131. Wang, Q. W., Robson, T. M., Pieristè, M., Oguro, M., Oguchi, R., Murai, Y., & Kurokawa, H. (2020). Testing trait plasticity over the range of spectral composition of sunlight in forb species differing in shade tolerance. *Journal of Ecology*. <https://doi.org/10.1111/1365-2745.13384>
  132. Lovelock, C. E., Osmond, C. B., & Jebb, M. (1994). Photoinhibition and recovery in tropical plant species: Response to disturbance. *Oecologia*, 97(3), 297–307. <https://doi.org/10.1007/BF00317318>
  133. Nichol, C. J., Pieruschka, R., Takayama, K., & B. F. R., Kolber, Z., Rascher, U., Grace, J., Robinson, S. A., Pogson, B., & Osmond, B. (2012). Canopy conundrums: Building on the Biosphere 2 experience to scale measurements of inner and outer canopy photoprotection from the leaf to the landscape. *Functional Plant Biology*, 39(1), 1–24. <https://doi.org/10.1071/fp11255>
  134. Barnes, P. W., Kersting, A. R., Flint, S. D., Beyschlag, W., & Ryel, R. J. (2013). Adjustments in epidermal UV-transmittance of leaves in sun-shade transitions. *Physiologia Plantarum*, 149, 200–213. <https://doi.org/10.1111/pp1.12025>
  135. Barnes, P. W., Robson, T. M., Tobler, M. A., Bottger, I. N., & Flint, S. D. (2017). Plant responses to fluctuating UV environments. In B. Jordan (Ed.), *UV-B Radiation and Plant Life: Molecular Biology to Ecology* (pp. 72–89). CABI. <https://doi.org/10.1079/9781780648590.0072>
  136. Dahal, D., Liu, S., & Oeding, J. (2014). The carbon cycle and hurricanes in the United States between 1900 and 2011. *Scientific Reports*. <https://doi.org/10.1038/srep05197>
  137. Veraverbeke, S., Delcourt, C. J. F., Kukavskaya, E., Mack, M., Walker, X., Hessilt, T., Rogers, B., & Scholten, R. C. (2021). Direct and longer-term carbon emissions from arctic-boreal fires: A short review of recent advances. *Current Opinion in Environmental Science & Health*. <https://doi.org/10.1016/j.coesh.2021.100277>
  138. Prestes, N. C. C., & d. S., Massi, K. G., Silva, E. A., Nogueira, D. S., de Oliveira, E. A., Freitag, R., Marimon, B. S., Marimon-Junior, B. H., Keller, M., & Feldpausch, T. R. (2020). Fire effects on understory forest regeneration in Southern Amazonia. *Frontiers in Forests and Global Change*. <https://doi.org/10.3389/ffgc.2020.00010>
  139. Knuesting, J., Brinkmann, M. C., Silva, B., Schorsch, M., Bendix, J., Beck, E., & Scheibe, R. (2018). Who will win where and why? An ecophysiological dissection of the competition



- between a tropical pasture grass and the invasive weed Bracken over an elevation range of 1000 m in the tropical Andes. *PLoS ONE*, 13(8), e0202255. <https://doi.org/10.1371/journal.pone.0202255>
140. Barnes, P. W., Ryel, R. J., & Flint, S. D. (2017). UV screening in native and non-native plant species in the tropical alpine: Implications for climate change-driven migration of species to higher elevations. *Frontiers in Plant Science*, 8(1451), 12–21. <https://doi.org/10.3389/fpls.2017.01451>
  141. Watermann, L. Y., Hock, M., Blake, C., & Erfmeier, A. (2019). Plant invasion into high elevations implies adaptation to high UV-B environments: A multi-species experiment. *Biological Invasions*. <https://doi.org/10.1007/s10530-019-02173-9>
  142. Rammer, W., Braziliunas, K. H., Hansen, W. D., Ratajczak, Z., Westerling, A. L., Turner, M. G., & Seidl, R. (2021). Widespread regeneration failure in forests of Greater Yellowstone under scenarios of future climate and fire. *Global Change Biology*, 27(18), 4339–4351. <https://doi.org/10.1111/gcb.15726>
  143. Werner, C., Meredith, L. K., Ladd, S. N., Ingrisch, J., Kübert, A., van Haren, J., Bahn, M., Bailey, K., Bamberger, I., Beyer, M., Blomdahl, D., Byron, J., Daber, E., Deleeuw, J., Dippold, M. A., Fudyma, J., Gil-Loaiza, J., Honeker, L. K., Hu, J., ... Williams, J. (2021). Ecosystem fluxes during drought and recovery in an experimental forest. *Science*, 374(6574), 1514–1518. <https://doi.org/10.1126/science.abj6789>
  144. Scordo, F., Chandra, S., Suenaga, E., Kelson, S. J., Culpepper, J., Scaff, L., Tromboni, F., Caldwell, T. J., Seitz, C., Fiorenza, J. E., Williamson, C. E., Sadro, S., Rose, K. C., & Poulson, S. R. (2021). Smoke from regional wildfires alters lake ecology. *Scientific Reports*, 11(1), 10922. <https://doi.org/10.1038/s41598-021-89926-6>
  145. Wilson, S. R., Madronich, S., Longstreth, J. D., & Solomon, K. R. (2019). Interactive effects of changing stratospheric ozone and climate on composition of the troposphere, air quality, and consequences for human and ecosystem health. *Photochemical & Photobiological Sciences*, 18, 775–803. <https://doi.org/10.1039/C8PP90064G>
  146. Francis, D., Fonseca, R., Nelli, N., Cuesta, J., Weston, M., Evan, A., & Temimi, M. (2020). The atmospheric drivers of the major Saharan dust storm in June 2020. *Geophysical Research Letters*. <https://doi.org/10.1029/2020gl090102>
  147. Zheng, G., Sedlacek, A. J., Aiken, A. C., Feng, Y., Watson, T. B., Raveh-Rubin, S., Uin, J., Lewis, E. R., & Wang, J. (2020). Long-range transported North American wildfire aerosols observed in marine boundary layer of eastern North Atlantic. *Environment International*, 139, 105680. <https://doi.org/10.1016/j.envint.2020.105680>
  148. Durand, M., Murchie, E. H., Lindfors, A. V., Urban, O., Aphalo, P. J., & Robson, T. M. (2021). Diffuse solar radiation and canopy photosynthesis in a changing environment. *Agricultural and Forest Meteorology*, 311, 108684. <https://doi.org/10.1016/j.agrfor.2021.108684>
  149. Chen, Z., Gao, W., Reddy, K. R., Chen, M., Taduri, S., Meyers, S. L., & Shankle, M. W. (2020). Ultraviolet (UV) B effects on growth and yield of three contrasting sweet potato cultivars. *Photosynthetica*. <https://doi.org/10.32615/ps.2019.137>
  150. Cronin, T. W., & Bok, M. J. (2016). Photoreception and vision in the ultraviolet. *Journal of Experimental Biology*, 219(18), 2790–2801. <https://doi.org/10.1242/jeb.128769>
  151. Fennell, J. T., Fountain, M. T., & Paul, N. D. (2019). Direct effects of protective cladding material on insect pests in crops. *Crop Protection*, 121, 147–156. <https://doi.org/10.1016/j.cropro.2019.04.003>
  152. Onzo, A., Sabelis, M. W., & Hanna, R. (2010). Effects of ultraviolet radiation on predatory mites and the role of refuges in plant structures. *Environmental Entomology*, 39(2), 695–701. <https://doi.org/10.1603/en09206>
  153. Ohtsuka, K., & Osakabe, M. (2009). Deleterious effects of UV-B radiation on herbivorous spider mites: They can avoid it by remaining on lower leaf surfaces. *Environmental Entomology*, 38(3), 920–929. <https://doi.org/10.1603/022.038.0346>
  154. Yin, W. D., Hoffmann, A. A., Gu, X. B., & Ma, C. S. (2018). Behavioral thermoregulation in a small herbivore avoids direct UVB damage. *Journal of Insect Physiology*, 107, 276–283. <https://doi.org/10.1016/j.jinsphys.2017.12.002>
  155. Hu, Z. Q., Zhao, H. Y., & Thieme, T. (2013). The effects of enhanced ultraviolet-B radiation on the biology of green and brown morphs of *Sitobion avenae* (Hemiptera: Aphididae). *Environmental Entomology*, 42(3), 578–585. <https://doi.org/10.1603/en12136>
  156. Burdick, S. C., Prischmann-Voldseth, D. A., & Harmon, J. P. (2015). Density and distribution of soybean aphid, *Aphis glycines* Matsumura (Hemiptera: Aphididae) in response to UV radiation. *Population Ecology*. <https://doi.org/10.1007/s10144-015-0501-6>
  157. Yoshioka, Y., Gotoh, T., & Suzuki, T. (2018). UV-B susceptibility and photoreactivation in embryonic development of the two-spotted spider mite. *Tetranychus urticae*. *Experimental and Applied Acarology*, 75(2), 155–166. <https://doi.org/10.1007/s10493-018-0263-x>
  158. Murata, Y., & Osakabe, M. (2017). Photo-enzymatic repair of UVB-induced DNA damage in the two-spotted spider mite *Tetranychus urticae*. *Experimental and Applied Acarology*, 71(1), 15–34. <https://doi.org/10.1007/s10493-016-0100-z>
  159. Tian, C. B., Li, Y. Y., Wang, X., Fan, W. H., Wang, G., Liang, J. Y., Wang, Z. Y., & Liu, H. (2019). Effects of UV-B radiation on the survival, egg hatchability and transcript expression of antioxidant enzymes in a high-temperature adapted strain of *Neoseiulus barkeri*. *Experimental and Applied Acarology*, 77(4), 527–543. <https://doi.org/10.1007/s10493-019-00361-9>
  160. Sugioka, N., Kawakami, M., Hirai, N., & Osakabe, M. (2018). A pollen diet confers ultraviolet-B resistance in phytoseiid mites by providing antioxidants. *Frontiers in Ecology and Evolution*, 6, 133. <https://doi.org/10.3389/fevo.2018.00133>
  161. Koveos, D. S., Suzuki, T., Terzidou, A., Kokkari, A., Floros, G., Damos, P., & Kouloussis, N. A. (2017). Egg hatching response to a range of ultraviolet-B (UV-B) radiation doses for four predatory mites and the herbivorous spider mite *Tetranychus urticae*. *Experimental and Applied Acarology*, 71(1), 35–46. <https://doi.org/10.1007/s10493-016-0102-x>
  162. Cui, H. Y., Zeng, Y. Y., Reddy, G. V. P., Gao, F., Li, Z. H., & Zhao, Z. H. (2021). UV radiation increases mortality and decreases the antioxidant activity in a tephritid fly. *Food and Energy Security*. <https://doi.org/10.1002/fes3.297>
  163. Fennell, J. T., Wilby, A., Sobeih, W., & Paul, N. D. (2020). New understanding of the direct effects of spectral balance on behaviour in *Myzus persicae*. *Journal of Insect Physiology*, 126, 104096. <https://doi.org/10.1016/j.jinsphys.2020.104096>
  164. Rizzini, L., Favory, J.-J., Cloix, C., Faggionato, D., O'Hara, A., Kaiserli, E., Baumeister, R., Schaefer, E., Nagy, F., Jenkins, G. I., & Ulm, R. (2011). Perception of UV-B by the *Arabidopsis* UVR8 protein. *Science*, 332(6025), 103–106. <https://doi.org/10.1126/science.1200660>
  165. Rai, N., O'Hara, A., Farkas, D., Safronov, O., Ratanasopa, K., Wang, F., Lindfors, A. V., Jenkins, G. I., Lehto, T., Salojärvi, J., Brosche, M., Strid, A., Aphalo, P. J., & Morales, L. O. (2020). The photoreceptor UVR8 mediates the perception of both UV-B and UV-A wavelengths up to 350 nm of sunlight with responsiveness moderated by cryptochromes. *Plant, Cell & Environment*. <https://doi.org/10.1111/pce.13752>
  166. Rai, N., Morales, L. O., & Aphalo, P. J. (2021). Perception of solar UV radiation by plants: photoreceptors and mechanisms.



- Plant Physiology*, 186(3), 1382–1396. <https://doi.org/10.1093/plphys/kiab162>
167. Madronich, S., McKenzie, R. L., Caldwell, M., & Björn, L. O. (1995). Changes in ultraviolet radiation reaching the earth's surface. *Ambio A Journal of Human Environment*, 24(3), 143–152.
  168. Fernández, M. B., Tossi, V., Lamattina, L., & Cassia, R. (2016). A comprehensive phylogeny reveals functional conservation of the UV-B photoreceptor UVR8 from green algae to higher plants. *Frontiers in Plant Science*, 7, 1698. <https://doi.org/10.3389/fpls.2016.01698>
  169. Balcerowicz, M. (2022). Ancient sun protection: The evolutionary origin of plant UV-B signaling. *Plant Physiology*, 188(1), 29–31. <https://doi.org/10.1093/plphys/kiab517>
  170. Zhang, Z., Xu, C., Zhang, S., Shi, C., Cheng, H., Liu, H., & Zhong, B. (2022). Origin and adaptive evolution of UV RESISTANCE LOCUS 8-mediated signaling during plant terrestrialization. *Plant Physiology*, 188(1), 332–346. <https://doi.org/10.1093/plphys/kiab486>
  171. Podolec, R., Demarsy, E., & Ulm, R. (2021). Perception and signaling of ultraviolet-B radiation in plants. *Annual Review of Plant Biology*, 72, 793–822. <https://doi.org/10.1146/annurev-arplant-050718-095946>
  172. Podolec, R., Lau, K., Wagnon, T. B., Hothorn, M., & Ulm, R. (2021). A constitutively monomeric UVR8 photoreceptor confers enhanced UV-B photomorphogenesis. *Proceedings of the National Academy of Sciences USA*. <https://doi.org/10.1073/pnas.2017284118>
  173. Wang, D., Sun, Y., Tu, M., Zhang, P., Wang, X., Wang, T., & Li, J. (2021). Response of *Zebrina pendula* leaves to enhanced UV-B radiation. *Functional Plant Biology*, 48(9), 851–859. <https://doi.org/10.1071/FP20274>
  174. Qian, M., Rosenqvist, E., Prinsen, E., Pescheck, F., Flygare, A.-M., Kalbina, I., Jansen, M. A. K., & Strid, Å. (2021). Downsizing in plants - UV induces pronounced morphological changes in cucumber in the absence of stress. *Biorxiv*. <https://doi.org/10.1101/2021.02.27.432481>
  175. Bidel, L. P. R., Meyer, S., Talhouet, A. C., Baudin, X., Daniel, C., Cazals, G., & Streb, P. (2020). Epidermal UVA screening capacity measured in situ as an indicator of light acclimation state of leaves of a very plastic alpine plant *Soldanella alpina* L. *Plant Physiology and Biochemistry*, 151, 10–20. <https://doi.org/10.1016/j.plaphy.2020.02.045>
  176. Nichelmann, L., & Pescheck, F. (2021). Solar UV-B effects on composition and UV screening efficiency of foliar phenolics in *Arabidopsis thaliana* are augmented by temperature. *Physiologia Plantarum*. <https://doi.org/10.1111/ppl.13554>
  177. Mo, Y., Nagel, C., & Taylor, L. P. (1992). Biochemical complementation of chalcone synthase mutants defines a role for flavonols in functional pollen. *Proceedings of the National Academy of Sciences USA*, 89(15), 7213–7217. <https://doi.org/10.1073/pnas.89.15.7213>
  178. Muhlemann, J. K., Younts, T. L. B., & Muday, G. K. (2018). Flavonols control pollen tube growth and integrity by regulating ROS homeostasis during high-temperature stress. *Proceedings of the National Academy of Sciences of the United States of America*, 115(47), E11188–E11197. <https://doi.org/10.1073/pnas.1811492115>
  179. Wang, L., Lam, P. Y., Lui, A. C. W., Zhu, F. Y., Chen, M. X., Liu, H., Zhang, J., & Lo, C. (2020). Flavonoids are indispensable for complete male fertility in rice. *Journal of Experimental Botany*, 71(16), 4715–4728. <https://doi.org/10.1093/jxb/eraa204>
  180. Brunetti, C., Di Ferdinando, M., Fini, A., Pollastri, S., & Tattini, M. (2013). Flavonoids as antioxidants and developmental regulators: Relative significance in plants and humans. *International Journal of Molecular Sciences*, 14(2), 3540–3555. <https://doi.org/10.3390/ijms14023540>
  181. Peach, K., Liu, J. W., & Mazer, S. J. (2020). Climate predicts uv floral pattern size, anthocyanin concentration, and pollen performance in *Clarkia unguiculata*. *Frontiers in Plant Science*, 11, 847. <https://doi.org/10.3389/fpls.2020.00847>
  182. Koski, M. H., Berardi, A. E., & Galloway, L. F. (2020). Pollen colour morphs take different paths to fitness. *Journal of Evolutionary Biology*, 33, 388–400. <https://doi.org/10.1111/jeb.13599>
  183. Xue, J. S., Zhang, B., Zhan, H., Lv, Y. L., Jia, X. L., Wang, T., Yang, N. Y., Lou, Y. X., Zhang, Z. B., Hu, W. J., Gui, J., Cao, J., Xu, P., Zhou, Y., Hu, J. F., Li, L., & Yang, Z. N. (2020). Phenylpropanoid derivatives are essential components of sporopollenin in vascular plants. *Molecular Plant*, 13(11), 1644–1653. <https://doi.org/10.1016/j.molp.2020.08.005>
  184. Agati, G., Brunetti, C., Fini, A., Gori, A., Guidi, L., Landi, M., Sebastiani, F., & Tattini, M. (2020). Are flavonoids effective antioxidants in plants? *Twenty years of our investigation. Antioxidants (Basel)*. <https://doi.org/10.3390/antiox9111098>
  185. Barnes, P. W., Tobler, M. A., Keefover-Ring, K., Flint, S. D., Barkley, A. E., Ryel, R. J., & Lindroth, R. L. (2016). Rapid modulation of ultraviolet shielding in plants is influenced by solar ultraviolet radiation and linked to alterations in flavonoids. *Plant Cell and Environment*, 39(1), 222–230. <https://doi.org/10.1111/pce.12609>
  186. Neugart, S., Tobler, M. A., & Barnes, P. W. (2021). Rapid adjustment in epidermal UV sunscreen: Comparison of optical measurement techniques and response to changing solar UV radiation conditions. *Physiologia Plantarum*, 173(3), 725–735. <https://doi.org/10.1111/ppl.13517>
  187. Gori, A., Nascimento, L. B., Ferrini, F., Centritto, M., & Brunetti, C. (2020). Seasonal and diurnal variation in leaf phenolics of three medicinal Mediterranean wild species: What is the best harvesting moment to obtain the richest and the most antioxidant extracts? *Molecules*, 25(4), 956. <https://doi.org/10.3390/molecules25040956>
  188. Badmus, U. O., Ač, A., Klem, K., Urban, O., & Jansen, M. A. K. (2022). A meta-analysis of the effects of UV radiation on the plant carotenoid pool. *Plant Physiology and Biochemistry*, 183, 36–45. <https://doi.org/10.1016/j.plaphy.2022.05.001>
  189. Piccolella, S., Crescente, G., Pacifico, F., & Pacifico, S. (2018). Wild aromatic plants bioactivity: A function of their (poly) phenol seasonality? A case study from Mediterranean area. *Phytochemistry Reviews*, 17(4), 785–799. <https://doi.org/10.1007/s11101-018-9558-0>
  190. Gori, A., Brunetti, C., Dos Santos Nascimento, L. B., Marino, G., Guidi, L., Ferrini, F., Centritto, M., Fini, A., & Tattini, M. (2021). Photoprotective role of photosynthetic and non-photosynthetic pigments in *Phillyrea latifolia*: Is their “antioxidant” function prominent in leaves exposed to severe summer drought? *International Journal of Molecular Sciences*. <https://doi.org/10.3390/ijms22158303>
  191. Neugart, S., Tobler, M. A., & Barnes, P. W. (2021). The function of flavonoids in the diurnal rhythm under rapidly changing UV conditions-A model study on okra. *Plants (Basel)*. <https://doi.org/10.3390/plants10112268>
  192. Hartikainen, S. M., & Robson, T. M. (2022). The roles of species' relatedness and climate of origin in determining optical leaf traits over a large set of taxa growing at high elevation and high latitude. *Frontiers in Plant Science*, 13, 1058162. <https://doi.org/10.3389/fpls.2022.1058162>
  193. Zhou, S., Yan, X., Yang, J., Qian, C., Yin, X., Fan, X., Fang, T., Gao, Y., Chang, Y., & Liu, W. (2021). Variations in flavonoid metabolites along altitudinal gradient in a desert medicinal plant *Agriophyllum squarrosum*. *Frontiers in Plant Science*, 12, 1306. <https://doi.org/10.3389/fpls.2021.683265>
  194. Eriksson, M., & Rafajlovic, M. (2022). The role of phenotypic plasticity in the establishment of range margins. *Philosophical*

- Transactions of the Royal Society of London B Biological Sciences*, 377(1846), 20210012. <https://doi.org/10.1098/rstb.2021.0012>
195. Schmid, M., & Guillaume, F. (2017). The role of phenotypic plasticity on population differentiation. *Heredity*, 119(4), 214–225. <https://doi.org/10.1038/hdy.2017.36>
  196. Crispo, E. (2008). Modifying effects of phenotypic plasticity on interactions among natural selection, adaptation and gene flow. *Journal of Evolutionary Biology*, 21(6), 1460–1469. <https://doi.org/10.1111/j.1420-9101.2008.01592.x>
  197. Song, B., Gao, Y., Stöcklin, J., Song, M., Sun, L., & Sun, H. (2020). Ultraviolet screening increases with elevation in translucent bracts of *Rheum nobile* (Polygonaceae), an alpine ‘glass-house’ plant from the high Himalayas. *Botanical Journal of the Linnean Society*. <https://doi.org/10.1093/botlinnean/boaa005>
  198. Yang, Q., Bi, H., Yang, W., Li, T., Jiang, J., Zhang, L., Liu, J., & Hu, Q. (2020). The genome sequence of alpine *Megacarpaea delavayi* identifies species-specific whole-genome duplication. *Frontiers in Genetics*. <https://doi.org/10.3389/fgene.2020.00812>
  199. Zhang, X., Sun, Y., Landis, J. B., Shen, J., Zhang, H., Kuang, T., Sun, W., Sun, J., Tamiyu, B. B., Deng, T., Sun, H., & Wang, H. (2021). Transcriptomes of *Saussurea* (Asteraceae) provide insights into high-altitude adaptation. *Plants-Basel*. <https://doi.org/10.3390/plants10081715>
  200. Wang, X., Liu, S., Zuo, H., Zheng, W., Zhang, S., Huang, Y., Pingcuo, G., Ying, H., Zhao, F., Li, Y., Liu, J., Yi, T.-S., Zan, Y., Larkin, R. M., Deng, X., Zeng, X., & Xu, Q. (2021). Genomic basis of high-altitude adaptation in Tibetan *Prunus* fruit trees. *Current Biology*, 31(17), 3848. <https://doi.org/10.1016/j.cub.2021.06.062>
  201. Panchen, Z. A., Doubt, J., Kharouba, H. M., & Johnston, M. O. (2019). Patterns and biases in an Arctic herbarium specimen collection: Implications for phenological research. *Applications in Plant Sciences*, 7(3), e1229. <https://doi.org/10.1002/aps3.1229>
  202. Seddon, A. W. R., Festi, D., Robson, T. M., & Zimmermann, B. (2019). Fossil pollen and spores as a tool for reconstructing ancient solar-ultraviolet irradiance received by plants: An assessment of prospects and challenges using proxy-system modelling. *Photochemical & Photobiological Sciences*, 18(2), 275–294. <https://doi.org/10.1039/C8PP00490K>
  203. Jardine, P. E., Hoorn, C., Beer, M. A. M., Barbolini, N., Woutersen, A., Bogota-Angel, G., Gosling, W. D., Fraser, W. T., Lomax, B. H., Huang, H., Sciumbata, M., He, H., Dupont-Nivet, G., & Kustatscher, E. (2021). Sporopollenin chemistry and its durability in the geological record: An integration of extant and fossil chemical data across the seed plants. *Palaeontology*, 64(2), 285–305. <https://doi.org/10.1111/pala.12523>
  204. IPCC. (2022). Summary for Policymakers. In D.C.R.H.-O. Pörtner, E. S. Poloczanska, K. Mintenbeck, M. Tignor, A. Alegría, M. Craig, S. Langsdorf, S. Löschke, V. Möller, & A. Okem (Eds.), *Climate Change 2022: Impacts, Adaptation, and Vulnerability Contribution of Working Group II to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press. <https://doi.org/10.1017/9781009325844.001>
  205. Jansen, M. A. K., Ač, A., Klem, K., & Urban, O. (2022). A meta-analysis of the interactive effects of UV and drought on plants. *Plant, Cell & Environment*, 45(1), 41–54. <https://doi.org/10.1111/pce.14221>
  206. Díaz-Guerra, L., Llorens, L., Julkunen-Tiitto, R., Nogués, I., Font, J., González, J. A., & Verdaguer, D. (2019). Leaf biochemical adjustments in two Mediterranean resprouter species facing enhanced UV levels and reduced water availability before and after aerial biomass removal. *Plant Physiology and Biochemistry*, 137, 130–143. <https://doi.org/10.1016/j.plaphy.2019.01.031>
  207. Verdaguer, D., Díaz-Guerra, L., Font, J., González, J. A., & Llorens, L. (2018). Contrasting seasonal morphological and physio-biochemical responses to UV radiation and reduced rainfall of two mature naturally growing Mediterranean shrubs in the context of climate change. *Environmental and Experimental Botany*, 147, 189–201. <https://doi.org/10.1016/j.envexpbot.2017.12.007>
  208. Aphalo, P. J., & Sadras, V. O. (2021). Explaining preemptive acclimation by linking information to plant phenotype. *Journal of Experimental Botany*. <https://doi.org/10.1093/jxb/erab537>
  209. la Sàenz-de, O., & D., Morales, L. O., Strid, A., Torres-Pacheco, I., & Guevara-Gonzalez, R. G. (2021). Ultraviolet-B exposure and exogenous hydrogen peroxide application lead to cross-tolerance toward drought in *Nicotiana tabacum* L. *Physiologia Plantarum*. <https://doi.org/10.1111/ppl.13448>
  210. Gourlay, G., Hawkins, B. J., Albert, A., Schnitzler, J. P., & Peter Constabel, C. (2022). Condensed tannins as antioxidants that protect poplar against oxidative stress from drought and UV-B. *Plant, Cell & Environment*, 45(2), 362–377. <https://doi.org/10.1111/pce.14242>
  211. Wang, H., Liu, S., Wang, T., Liu, H., Xu, X., Chen, K., & Zhang, P. (2020). The moss flavone synthase I positively regulates the tolerance of plants to drought stress and UV-B radiation. *Plant Science*, 298, 110591. <https://doi.org/10.1016/j.plantsci.2020.110591>
  212. Thomas, T. T., & D., & Puthur, J. T. (2017). UV radiation priming: A means of amplifying the inherent potential for abiotic stress tolerance in crop plants. *Environmental and Experimental Botany*, 138, 57–66. <https://doi.org/10.1016/j.envexpbot.2017.03.003>
  213. Klem, K., Oravec, M., Holub, P., Šimor, J., Findurová, H., Surá, K., Veselá, B., Hodaňová, P., Jansen, M. A. K., & Urban, O. (2022). Interactive effects of nitrogen, UV and PAR on barley morphology and biochemistry are associated with the leaf C: N balance. *Plant Physiology and Biochemistry*, 172, 111–124. <https://doi.org/10.1016/j.plaphy.2022.01.006>
  214. Williams, T. B., Dodd, I. C., Sobeih, W. Y., & Paul, N. D. (2022). Ultraviolet radiation causes leaf warming due to partial stomatal closure. *Horticulture Research*. <https://doi.org/10.1093/hr/uhab066>
  215. Ribeiro, A. F., Russo, A., Gouveia, C. M., & Pires, C. A. (2020). Drought-related hot summers: A joint probability analysis in the Iberian Peninsula. *Weather and Climate Extremes*, 30, 100279. <https://doi.org/10.1016/j.wace.2020.100279>
  216. Jin, P., Overmans, S., Duarte, C. M., Agustí, S., & Bates, A. (2019). Increasing temperature within thermal limits compensates negative ultraviolet-B radiation effects in terrestrial and aquatic organisms. *Global Ecology and Biogeography*. <https://doi.org/10.1111/geb.12973>
  217. Fang, H. C., Dong, Y. H., Yue, X. X., Chen, X. L., He, N. B., Hu, J. F., Jiang, S. H., Xu, H. F., Wang, Y. C., Su, M. Y., Zhang, J., Zhang, Z. Y., Wang, N., & Chen, X. S. (2019). MdCOL4 interaction mediates crosstalk between UV-B and high temperature to control fruit coloration in apple. *Plant and Cell Physiology*, 60, 1055–1066. <https://doi.org/10.1093/pcp/pcz023>
  218. Schulz, E., Tohge, T., Winkler, J. B., Albert, A., Schäffner, A. R., Fernie, A. R., Zuther, E., & Hinch, D. K. (2021). Natural variation among *Arabidopsis* accessions in the regulation of flavonoid metabolism and stress gene expression by combined UV radiation and cold. *Plant and Cell Physiology*, 62(3), 502–514. <https://doi.org/10.1093/pcp/pcab013>
  219. Coffey, A., & Jansen, M. A. K. (2019). Effects of natural solar UV-B radiation on three *Arabidopsis* accessions are strongly affected by seasonal weather conditions. *Plant Physiology and Biochemistry*, 134, 64–72. <https://doi.org/10.1016/j.plaphy.2018.06.016>
  220. Sangüesa-Barreda, G., Di Filippo, A., Piovesan, G., Rozas, V., Di Fiore, L., García-Hidalgo, M., García-Cervigón, A. I.,

- Muñoz-Garachana, D., Baliva, M., & Olano, J. M. (2021). Warmer springs have increased the frequency and extension of late-frost defoliations in southern European beech forests. *Science of the Total Environment*, 775, 145860. <https://doi.org/10.1016/j.scitotenv.2021.145860>
221. Slatyer, R. A., Umbers, K. D. L., & Arnold, P. A. (2021). Ecological responses to variation in seasonal snow cover. *Conservation Biology*. <https://doi.org/10.1111/cobi.13727>
  222. Zandalinas, S. I., Sengupta, S., Fritsch, F. B., Azad, R. K., Nechushtai, R., & Mittler, R. (2021). The impact of multifactorial stress combination on plant growth and survival. *New Phytologist*, 230(3), 1034–1048. <https://doi.org/10.1111/nph.17232>
  223. Deng, B., Liu, X., Zheng, L., Liu, Q., Guo, X., & Zhang, L. (2019). Effects of nitrogen deposition and UV-B radiation on seedling performance of Chinese tallow tree (*Triadica sebifera*): A photosynthesis perspective. *Forest Ecology and Management*, 433, 453–458. <https://doi.org/10.1016/j.foreco.2018.11.038>
  224. Tripathi, R., Rai, K., Singh, S., Agrawal, M., & Agrawal, S. (2019). Role of supplemental UV-B in changing the level of ozone toxicity in two cultivars of sunflower: Growth, seed yield and oil quality. *Ecotoxicology*, 28(3), 277–293. <https://doi.org/10.1007/s10646-019-02020-6>
  225. Urban, O., Hrstka, M., Holub, P., Veselá, B., Večeřová, K., Novotná, K., Grace, J., & Klem, K. (2019). Interactive effects of ultraviolet radiation and elevated CO<sub>2</sub> concentration on photosynthetic characteristics of European beech saplings during the vegetation season. *Plant Physiology and Biochemistry*, 134, 20–30. <https://doi.org/10.1016/j.plaphy.2018.08.026>
  226. Bornman, J. F., Barnes, P. W., Robinson, S. A., Ballaré, C. L., Flint, S. D., & Caldwell, M. M. (2015). Solar ultraviolet radiation and ozone depletion-driven climate change: effects on terrestrial ecosystems [Review]. *Photochemical & Photobiological Sciences*, 14(1), 88–107. <https://doi.org/10.1039/c4pp90034k>
  227. Nunez, S., Arets, E., Alkemade, R., Verwer, C., & Leemans, R. (2019). Assessing the impacts of climate change on biodiversity: Is below 2 °C enough? *Climatic Change*, 154(3–4), 351–365. <https://doi.org/10.1007/s10584-019-02420-x>
  228. Weiskopf, S. R., Rubenstein, M. A., Crozier, L. G., Gaichas, S., Griffiths, R., Halofsky, J. E., Hyde, K. J. W., Morelli, T. L., Morissette, J. T., Muñoz, R. C., Pershing, A. J., Peterson, D. L., Poudel, R., Staudinger, M. D., Sutton-Grier, A. E., Thompson, L., Vose, J., Weltzin, J. F., & Whyte, K. P. (2020). Climate change effects on biodiversity, ecosystems, ecosystem services, and natural resource management in the United States. *Science of the Total Environment*, 733, 137782. <https://doi.org/10.1016/j.scitotenv.2020.137782>
  229. Pecl, G. T., Araújo, M. B., Bell, J. D., Blanchard, J., Bonebrake, T. C., Chen, I.-C., Clark, T. D., Colwell, R. K., Danielsen, F., Evengård, B., Falconi, L., Ferrier, S., Frusher, S., Garcia, R. A., Griffiths, R. B., Hobday, A. J., Janion-Scheepers, C., Jarzyna, M. A., Jennings, S., ... Williams, S. E. (2017). Biodiversity redistribution under climate change: Impacts on ecosystems and human well-being. *Science*. <https://doi.org/10.1126/science.aai9214>
  230. Vitasse, Y., Ursenbacher, S., Klein, G., Bohnenstengel, T., Chittaro, Y., Delestrade, A., Monnerat, C., Rebetez, M., Rixen, C., Strebel, N., Schmidt, B. R., Wipf, S., Wohlgemuth, T., Yoccoz, N. G., & Lenoir, J. (2021). Phenological and elevational shifts of plants, animals and fungi under climate change in the European Alps. *Biological Reviews*, 96(5), 1816–1835. <https://doi.org/10.1111/brv.12727>
  231. Chowdhury, S., Fuller, R. A., Dingle, H., Chapman, J. W., & Zalucki, M. P. (2021). Migration in butterflies: A global overview. *Biological Reviews*, 96(4), 1462–1483. <https://doi.org/10.1111/brv.12714>
  232. Lenoir, J., Gégout, J.-C., Guisan, A., Vittoz, P., Wohlgemuth, T., Zimmermann, N. E., Dullinger, S., Pauli, H., Willner, W., & Svenning, J.-C. (2010). Going against the flow: Potential mechanisms for unexpected downslope range shifts in a warming climate. *Ecography*, 33(2), 295–303. <https://doi.org/10.1111/j.1600-0587.2010.06279.x>
  233. Tagliari, M. M., Danthu, P., Tsy, L. P., & J.-M., Cornu, C., Lenoir, J., Carvalho-Rocha, V., & Vieilledent, G. (2021). Not all species will migrate poleward as the climate warms: The case of the seven baobab species in Madagascar. *Global Change Biology*, 27(23), 6071–6085. <https://doi.org/10.1111/gcb.15859>
  234. Tomiolo, S., & Ward, D. (2018). Species migrations and range shifts: A synthesis of causes and consequences. *Perspectives in Plant Ecology, Evolution and Systematics*, 33, 62–77. <https://doi.org/10.1016/j.ppees.2018.06.001>
  235. IPCC, 2014: Climate change 2014: Impacts, adaptation, and vulnerability Part A: Global and sectoral aspects. Contribution of working group II to the fifth assessment report of the intergovernmental panel on climate change [Field, C.B., V.R. Barros, D.J. Dokken, K.J. Mach, M.D. Mastrandrea, T.E. Bilir, M. Chatterjee, K.L. Ebi, Y.O. Estrada, R.C. Genova, R. B. Girma, E.S. Kissel, A.N. Levy, S. MacCracken, P.R. Mastrandrea, and L.L. White (eds.)]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 1132 pp
  236. Tripp, E. A., Zhuang, Y., Schreiber, M., Stone, H., & Berardi, A. E. (2018). Evolutionary and ecological drivers of plant flavonoids across a large latitudinal gradient. *Molecular Phylogenetics and Evolution*, 128, 147–161. <https://doi.org/10.1016/j.ympev.2018.07.004>
  237. Castagna, A., Csepregi, K., Neugart, S., Zipoli, G., Večeřová, K., Jakab, G., Jug, T., Llorens, L., Martínez-Abaigar, J., Martínez-Lüscher, J., Núñez-Olivera, E., Ranieri, A., Schoedl-Hummel, K., Schreiner, M., Teszlák, P., Tittmann, S., Urban, O., Verdaguier, D., Jansen, M. A. K., & Hideg, É. (2017). Environmental plasticity of Pinot noir grapevine leaves; a trans-European study of morphological and biochemical changes along a 1500 km latitudinal climatic gradient. *Plant, Cell & Environment*, 40(11), 2790–2805. <https://doi.org/10.1111/pce.13054>
  238. Escobar-Bravo, R., Klinkhamer, P. G. L., & Leiss, K. A. (2017). Interactive effects of UV-B light with abiotic factors on plant growth and chemistry, and their consequences for defense against arthropod herbivores. *Frontiers in Plant Science*, 8, 278. <https://doi.org/10.3389/fpls.2017.00278>
  239. Escobar-Bravo, R., Nederpel, C., Naranjo, S., Kim, H. K., Rodríguez-López, M. J., Chen, G., Glauser, G., Leiss, K. A., & Klinkhamer, P. G. (2021). Ultraviolet radiation modulates both constitutive and inducible plant defenses against thrips but is dose and plant genotype dependent. *Journal of Pest Science*, 94(1), 69–81. <https://doi.org/10.1007/s10340-019-01166-w>
  240. Tedla, B., Dang, Q.-L., & Inoue, S. (2019). White birch has limited phenotypic plasticity to take advantage of increased photoperiods at higher latitudes north of the seed origin. *Forest Ecology and Management*, 451, 117565. <https://doi.org/10.1016/j.foreco.2019.117565>
  241. Way, D. A., & Montgomery, R. A. (2014). Photoperiod constraints on tree phenology, performance and migration in a warming world. *Plant, Cell & Environment*, 38(9), 1725–1736. <https://doi.org/10.1111/pce.12431>
  242. Tedla, B., Dang, Q. L., & Inoue, S. (2020). CO<sub>2</sub> elevation and photoperiods north of seed origin change autumn and spring phenology as well as cold hardiness in boreal white birch. *Frontiers in Plant Science*, 11, 506. <https://doi.org/10.3389/fpls.2020.00506>
  243. Mamantov, M. A., Gibson-Reinemer, D. K., Linck, E. B., & Sheldon, K. S. (2021). Climate-driven range shifts of montane



- species vary with elevation. *Global Ecology and Biogeography*, 30(4), 784–794. <https://doi.org/10.1111/geb.13246>
244. Wu, Y., Yang, Y., Liu, C., Hou, Y., Yang, S., Wang, L., & Zhang, X. (2021). Potential suitable habitat of two economically important forest trees (*Acer truncatum* and *Xanthoceras sorbifolium*) in east Asia under current and future climate scenarios. *Forests*. <https://doi.org/10.3390/f12091263>
  245. Kergunteuil, A., Descombes, P., Glauser, G., Pellissier, L., & Rasmann, S. (2018). Plant physical and chemical defence variation along elevation gradients: A functional trait-based approach. *Oecologia*, 187(2), 561–571. <https://doi.org/10.1007/s00442-018-4162-y>
  246. Pellissier, L., Moreira, X., Danner, H., Serrano, M., Salamin, N., van Dam, N. M., & Rasmann, S. (2016). The simultaneous inducibility of phytochemicals related to plant direct and indirect defences against herbivores is stronger at low elevation. *Journal of Ecology*, 104(4), 1116–1125. <https://doi.org/10.1111/1365-2745.12580>
  247. Moreira, X., Petry, W. K., Mooney, K. A., Rasmann, S., & Abdala-Roberts, L. (2018). Elevational gradients in plant defences and insect herbivory: Recent advances in the field and prospects for future research. *Ecography*, 41(9), 1485–1496. <https://doi.org/10.1111/ecog.03184>
  248. Fernández-Marín, B., Sáenz-Cenicerós, A., Solanki, T., Robson, T. M., & García-Plazaola, J. I. (2021). Alpine forbs rely on different photoprotective strategies during spring snowmelt. *Physiologia Plantarum*, 172(3), 1506–1517. <https://doi.org/10.1111/ppl.13342>
  249. Gateva, S. P., Jovtchev, G. V., Angelova, T. P., Nonova, T., Tyutyundzhiev, N. G., Geleva, E., Katrandzhiev, K. A., Nikolova, N., Dimitrov, D., & Angelov, V. (2022). Effect of UV radiation and other abiotic stress factors on DNA of different wild plant species grown in three successive seasons in alpine and subalpine regions. *Phyton*, 91(2), 293–313. <https://doi.org/10.32604/phyton.2022.016397>
  250. Fraser, D. P., Sharma, A., Fletcher, T., Budge, S., Moncrieff, C., Dodd, A. N., & Franklin, K. A. (2017). UV-B antagonises shade avoidance and increases levels of the flavonoid quercetin in coriander (*Coriandrum sativum*). *Scientific Reports*, 7(1), 17758. <https://doi.org/10.1038/s41598-017-18073-8>
  251. Ballaré, C. L., & Austin, A. T. (2019). Recalculating growth and defense strategies under competition: Key roles of photoreceptors and jasmonates. *Journal of Experimental Botany*, 70, 3425–3434. <https://doi.org/10.1093/jxb/erz237>
  252. Seastedt, T., & Oldfather, M. (2021). Climate change, ecosystem processes and biological diversity responses in high elevation communities. *Climate*. <https://doi.org/10.3390/cli9050087>
  253. Robson, T. M., Klem, K., Urban, O., & Jansen, M. A. K. (2015). Re-interpreting plant morphological responses to UV-B radiation. *Plant, Cell & Environment*, 38(5), 856–866. <https://doi.org/10.1111/pce.12374>
  254. Barnes, P. W., Ballaré, C. L., & Caldwell, M. M. (1996). Photomorphogenic effects of UV-B radiation on plants: Consequences for light competition. *Journal of Plant Physiology*, 148(1–2), 15–20. [https://doi.org/10.1016/S0176-1617\(96\)80288-4](https://doi.org/10.1016/S0176-1617(96)80288-4)
  255. Wolf, A., Zimmerman, N. B., Anderegg, W. R. L., Busby, P. E., & Christensen, J. (2016). Altitudinal shifts of the native and introduced flora of California in the context of 20th-century warming. *Global Ecology and Biogeography*, 25(4), 418–429. <https://doi.org/10.1111/geb.12423>
  256. Dainese, M., Aikio, S., Hulme, P. E., Bertolli, A., Prosser, F., & Marini, L. (2017). Human disturbance and upward expansion of plants in a warming climate. *Nature Climate Change*, 7, 577–580. <https://doi.org/10.1038/nclimate3337>
  257. Mosena, A., Steinlein, T., & Beyschlag, W. (2018). Reconstructing the historical spread of non-native plants in the North American West from herbarium specimens. *Flora*, 242, 45–52. <https://doi.org/10.1016/j.flora.2018.03.002>
  258. Wang, H., Ma, X. C., Zhang, L., Siemann, E., & Zou, J. W. (2016). UV-B has larger negative impacts on invasive populations of *Triadica sebifera* but ozone impacts do not vary. *Journal of Plant Ecology*, 9(1), 61–68. <https://doi.org/10.1093/jpe/rtv045>
  259. Václavík, T., Beckmann, M., Cord, A. F., & Bindewald, A. M. (2017). Effects of UV-B radiation on leaf hair traits of invasive plants—Combining historical herbarium records with novel remote sensing data. *PLoS ONE*, 12(4), e0175671. <https://doi.org/10.1371/journal.pone.0175671>
  260. Hock, M., Hofmann, R. W., Müller, C., & Erfmeier, A. (2019). Exotic plant species are locally adapted but not to high ultraviolet-B radiation: a reciprocal multispecies experiment. *Ecology*, 100(5), e02665. <https://doi.org/10.1002/ecy.2665>
  261. Jovtchev, G., Stankov, A., Ravnachka, I., Gateva, S., Dimitrov, D., Tyutyundzhiev, N., Nikolova, N., & Angelov, C. (2019). How can the natural radiation background affect DNA integrity in angiosperm plant species at different altitudes in Rila Mountain (Southwest Bulgaria)? *Environmental Science and Pollution Research*, 26(13), 13592–13601. <https://doi.org/10.1007/s11356-019-04872-1>
  262. Davidson, A. M., Jennions, M., & Nicotra, A. B. (2011). Do invasive species show higher phenotypic plasticity than native species and if so, is it adaptive? A meta-analysis. *Ecology Letters*, 14(4), 419–431. <https://doi.org/10.1111/j.1461-0248.2011.01596.x>
  263. Madronich, S. (1987). Photodissociation in the atmosphere: 1. Actinic flux and the effects of ground reflections and clouds. *Journal of Geophysical Research: Atmospheres*, 92(D8), 9740–9752. <https://doi.org/10.1029/JD092iD08p09740>
  264. Zu, K., Wang, Z., Zhu, X., Lenoir, J., Shrestha, N., Lyu, T., Luo, A., Li, Y., Ji, C., Peng, S., Meng, J., & Zhou, J. (2021). Upward shift and elevational range contractions of subtropical mountain plants in response to climate change. *Science of the Total Environment*, 783, 146896. <https://doi.org/10.1016/j.scitotenv.2021.146896>
  265. Benito Garzon, M., Robson, T. M., & Hampe, A. (2019). DeltaTraitSDMs: Species distribution models that account for local adaptation and phenotypic plasticity. *New Phytologist*, 222(4), 1757–1765. <https://doi.org/10.1111/nph.15716>
  266. Li, W., Shi, M., Huang, Y., Chen, K., Sun, H., & Chen, J. (2019). Climatic change can influence species diversity patterns and potential habitats of Salicaceae plants in China. *Forests*, 10(3), 220. <https://doi.org/10.3390/f10030220>
  267. Wang, D., Cui, B., Duan, S., Chen, J., Fan, H., Lu, B., & Zheng, J. (2019). Moving north in China: The habitat of *Pedicularis kansuensis* in the context of climate change. *Science of the Total Environment*, 697, 133979. <https://doi.org/10.1016/j.scitotenv.2019.133979>
  268. Li, M., He, J., Zhao, Z., Lyu, R., Yao, M., Cheng, J., & Xie, L. (2020). Predictive modelling of the distribution of *Clematis* sect *Fruticella* s. str under climate change reveals a range expansion during the Last Glacial Maximum. *PeerJ*. <https://doi.org/10.7717/peerj.8729>
  269. Zhang, K., Yao, L., Meng, J., & Tao, J. (2018). Maxent modeling for predicting the potential geographical distribution of two peony species under climate change. *Science of the Total Environment*, 634, 1326–1334. <https://doi.org/10.1016/j.scitotenv.2018.04.112>
  270. Peng, L. P., Cheng, F. Y., Hu, X. G., Mao, J. F., Xu, X. X., Zhong, Y., Li, S. Y., & Xian, H. L. (2019). Modelling environmentally suitable areas for the potential introduction and cultivation of the



- emerging oil crop *Paeonia ostii* in China. *Scientific Reports*, 9(1), 3213. <https://doi.org/10.1038/s41598-019-39449-y>
271. Zhang, K., Zhang, Y., & Tao, J. (2019). Predicting the potential distribution of *Paeonia veitchii* (Paeoniaceae) in China by incorporating climate change into a Maxent model. *Forests*. <https://doi.org/10.3390/f10020190>
  272. Hu, X. G., Jin, Y. Q., Wang, X. R., Mao, J. F., & Li, Y. (2015). Predicting impacts of future climate change on the distribution of the widespread conifer *Platycladus orientalis*. *PLoS ONE*, 10(7), e0132326. <https://doi.org/10.1371/journal.pone.0132326>
  273. Beckmann, M., Vaclavik, T., Manceur, A. M., Sprtova, L., von Wehrden, H., Welk, E., & Cord, A. F. (2014). gIUV: A global UV-B radiation data set for macroecological studies. *Methods in Ecology and Evolution*, 5(4), 372–383. <https://doi.org/10.1111/2041-210x.12168>
  274. Ren, T., He, N., Liu, Z., Li, M., Zhang, J., Li, A., Wei, C., Lü, X., & Han, X. (2021). Environmental filtering rather than phylogeny determines plant leaf size in three floristically distinctive plateaus. *Ecological Indicators*. <https://doi.org/10.1016/j.ecoli.2021.108049>
  275. Ye, M., Zhu, X., Gao, P., Jiang, L., & Wu, R. (2020). Identification of quantitative trait loci for altitude adaptation of tree leaf shape with *Populus szechuanica* in the Qinghai-Tibetan Plateau. *Frontiers in Plant Science*, 11, 632. <https://doi.org/10.3389/fpls.2020.00632>
  276. Li, X., Ke, X., Zhou, H., & Tang, Y. (2019). Contrasting altitudinal patterns of leaf UV reflectance and absorbance in four herbaceous species on the Qinghai-Tibetan Plateau. *Journal of Plant Ecology*, 12(2), 245–254. <https://doi.org/10.1093/jpe/rty016>
  277. Wang, Q.-W., Liu, C., Robson, T. M., Hikosaka, K., & Kurokawa, H. (2021). Leaf density and chemical composition explain variation in leaf mass area with spectral composition among 11 widespread forbs in a common garden. *Physiologia Plantarum*, 173(3), 698–708. <https://doi.org/10.1111/ppl.13512>
  278. Teramura, A. H. (1980). Effects of ultraviolet-B irradiances on soybean. I. Importance of photosynthetically active radiation in evaluating ultraviolet-B irradiance effects on soybean and wheat growth. *Physiologia Plantarum*, 48, 333–339. <https://doi.org/10.1111/j.1399-3054.1980.tb03264.x>
  279. Sisson, W. B., & Caldwell, M. M. (1977). Atmospheric ozone depletion: Reduction of photosynthesis and growth of a sensitive higher plant exposed to enhanced UV-B radiation. *Journal of Experimental Botany*, 28(104), 691–705. <https://doi.org/10.1093/jxb/28.3.691>
  280. Caldwell, M. M. (1971). Solar UV irradiation and the growth and development of higher plants. *Photophysiology*, 6, 131–177.
  281. Robson, T. M., Pieristè, M., Durand, M., Kotilainen, T. K., & Aphalo, P. J. (2022). The benefits of informed management of sunlight in production greenhouses and polytunnels. *Plants People Planet*. <https://doi.org/10.1002/ppp3.10258>
  282. Bernhard, G. H., Neale, R. E., Barnes, P. W., Neale, P. J., Zepp, R. G., Wilson, S. R., Andrady, A. L., Bais, A. F., McKenzie, R. L., Aucamp, P. J., Young, P. J., Liley, J. B., Lucas, R. M., Yazar, S., Rhodes, L. E., Byrne, S. N., Hollestein, L. M., Olsen, C. M., Young, A. R., ... White, C. C. (2020). Environmental effects of stratospheric ozone depletion, UV radiation and interactions with climate change: UNEP Environmental Effects Assessment Panel, update 2019. *Photochemical & Photobiological Sciences*, 19, 542–584. <https://doi.org/10.1039/d0pp90011g>
  283. Feeley, K. J., Stroud, J. T., & Perez, T. M. (2017). Most 'global' reviews of species' responses to climate change are not truly global. *Diversity and Distributions*, 23(3), 231–234. <https://doi.org/10.1111/ddi.12517>
  284. Sheldon, K. S. (2019). Climate change in the tropics: Ecological and evolutionary responses at low latitudes. *Annual Review of Ecology, Evolution, and Systematics*, 50(1), 303–333. <https://doi.org/10.1146/annurev-ecolsys-110218-025005>
  285. Feeley, K. J., & Stroud, J. T. (2018). Where on Earth are the "tropics"? *Frontiers of Biogeography*. <https://doi.org/10.21425/f5fbg38649>
  286. Raven, P. H., Gereau, R. E., Phillipson, P. B., Chatelain, C., Jenkins, C. N., & Ulloa Ulloa, C. (2010). The distribution of biodiversity richness in the tropics. *Science Advances*. <https://doi.org/10.1126/sciadv.abc6228>
  287. Secretariat of the Convention on Biological Diversity. (2008). Biodiversity and Agriculture: Safeguarding Biodiversity and Securing Food for the World. Montreal, 56 pp
  288. Chipperfield, M. P., Bekki, S., Dhomse, S., Harris, N. R. P., Hassler, B., Hossaini, R., Steinbrecht, W., Thieblemont, R., & Weber, M. (2017). Detecting recovery of the stratospheric ozone layer. *Nature*, 549(7671), 211–218. <https://doi.org/10.1038/nature23681>
  289. Krause, G. H., Jahns, P., Virgo, A., Garcia, M., Aranda, J., Wellmann, E., & Winter, K. (2007). Photoprotection, photosynthesis and growth of tropical tree seedlings under near-ambient and strongly reduced solar ultraviolet-B radiation. *Journal of Plant Physiology*, 164, 1311–1322. <https://doi.org/10.1016/j.jplph.2006.09.004>
  290. Kataria, S., & Guruprasad, K. N. (2015). Exclusion of solar UV radiation improves photosynthetic performance and yield of wheat varieties. *Plant Physiology and Biochemistry*, 97, 400–411. <https://doi.org/10.1016/j.plaphy.2015.10.001>
  291. Kataria, S., & Baghel, L. (2016). Influence of UV exclusion and selenium on carbon fixation, nitrogen fixation and yield of soybean variety JS-335. *South African Journal of Botany*, 103, 126–134. <https://doi.org/10.1016/j.sajb.2015.09.003>
  292. Mazza, C. A., Gimenez, P. I., Kantolic, A. G., & Ballaré, C. L. (2013). Beneficial effects of solar UV-B radiation on soybean yield mediated by reduced insect herbivory under field conditions. *Physiologia Plantarum*, 147(3), 307–315. <https://doi.org/10.1111/j.1399-3054.2012.01661.x>
  293. Mansour, G., Ghanem, C., Mercenaro, L., Nassif, N., Hassoun, G., & Del Caro, A. (2022). Effects of altitude on the chemical composition of grapes and wine: a review. *OENO One*, 56(1), 227–239. <https://doi.org/10.20870/oeno-one.2022.56.1.4895>
  294. Hinojos Mendoza, G., Gutierrez Ramos, C. A., Heredia Corral, D. M., Soto Cruz, R., & Garbolino, E. (2020). Assessing suitable areas of common grapevine (*Vitis vinifera* L.) for current and future climate situations: The CDS Toolbox SDM. *Atmosphere*, 11(11), 1201. <https://doi.org/10.3390/atmos11111201>
  295. Moriondo, M., Jones, G. V., Bois, B., Dibari, C., Ferrise, R., Trombi, G., & Bindi, M. (2013). Projected shifts of wine regions in response to climate change. *Climatic Change*, 119(3), 825–839. <https://doi.org/10.1007/s10584-013-0739-y>
  296. Hannah, L., Roehrdanz, P. R., Ikegami, M., Shepard, A. V., Shaw, M. R., Tabor, G. M., Zhi, L., Marquet, P. A., & Hijmans, R. J. (2013). Climate change, wine, and conservation. *Proceedings of the National Academy of Sciences*, 110(17), 6907–6912. <https://doi.org/10.1073/pnas.1210127110>
  297. Malhotra, S. (2017). Horticultural crops and climate change: A review. *Indian Journal of Agricultural Sciences*, 87(1), 12–22.
  298. Ginbo, T. (2022). Heterogeneous impacts of climate change on crop yields across altitudes in Ethiopia. *Climatic Change*. <https://doi.org/10.1007/s10584-022-03306-1>
  299. Derebe, A. D., Gobena Roro, A., Tessfaye Asfaw, B., Worku Ayele, W., & Hvorslef-Eide, A. K. (2019). Effects of solar UV-B radiation exclusion on physiology, growth and yields of taro (*Colocasia esculenta* (L.)) at different altitudes in tropical environments of Southern Ethiopia. *Scientia Horticulturae*, 256, 108563. <https://doi.org/10.1016/j.scienta.2019.108563>

300. Tonnang, H. E., Sokame, B. M., Abdel-Rahman, E. M., & Dubois, T. (2022). Measuring and modelling crop yield losses due to invasive insect pests under climate change. *Current Opinion in Insect Science*. <https://doi.org/10.1016/j.cois.2022.100873>
301. Wallingford, P. D., Morelli, T. L., Allen, J. M., Beaury, E. M., Blumenthal, D. M., Bradley, B. A., Dukes, J. S., Early, R., Fusco, E. J., Goldberg, D. E., Ibáñez, I., Laginhas, B. B., Vilà, M., & Sorte, C. J. B. (2020). Adjusting the lens of invasion biology to focus on the impacts of climate-driven range shifts. *Nature Climate Change*, 10(5), 398–405. <https://doi.org/10.1038/s41558-020-0768-2>
302. Schweiger, O., Biesmeijer, J. C., Bommarco, R., Hickler, T., Hulme, P. E., Klotz, S., Kühn, I., Moora, M., Nielsen, A., Ohlemüller, R., Petanidou, T., Potts, S. G., Pyšek, P., Stout, J. C., Sykes, M. T., Tscheulin, T., Vilà, M., Walther, G.-R., Westphal, C., ... Settele, J. (2010). Multiple stressors on biotic interactions: How climate change and alien species interact to affect pollination. *Biological Reviews*, 85(4), 777–795. <https://doi.org/10.1111/j.1469-185X.2010.00125.x>
303. Weaver, S. A., & Mallinger, R. E. (2022). A specialist bee and its host plants experience phenological shifts at different rates in response to climate change. *Ecology*. <https://doi.org/10.1002/ecy.3658>
304. Jaiswal, D., Pandey, A., Mukherjee, A., Agrawal, M., & Agrawal, S. B. (2020). Alterations in growth, antioxidative defense and medicinally important compounds of *Curcuma caesia* Roxb. Under elevated ultraviolet-B radiation. *Environmental and Experimental Botany*, 177, 104152. <https://doi.org/10.1016/j.envexpbot.2020.104152>
305. Neugart, S., & Schreiner, M. (2018). UVB and UVA as eustressors in horticultural and agricultural crops. *Scientia Horticulturae*, 234, 370–381. <https://doi.org/10.1016/j.scienta.2018.02.021>
306. Robson, T. M., Aphalo, P. J., Banas, A. K., Barnes, P. W., Brelsford, C. C., Jenkins, G. I., Kotilainen, T., Labuz, J., Martínez-Abaigar, J., Morales, L. O., Neugart, S., Pieristè, M., Rai, N., Vandenbussche, F., & Jansen, M. (2019). A perspective on ecologically relevant plant-UV research and its practical application. *Photochemical & Photobiological Sciences*, 18, 970–988. <https://doi.org/10.1039/c8pp00526e>
307. Takshak, S., & Agrawal, S. B. (2019). Defense potential of secondary metabolites in medicinal plants under UV-B stress. *Journal of Photochemistry and Photobiology B: Biology*, 193, 51–88. <https://doi.org/10.1016/j.jphotobiol.2019.02.002>
308. Lin, N., Liu, X. Y., Zhu, W. F., Cheng, X., Wang, X. H., Wan, X. C., & Liu, L. L. (2021). Ambient Ultraviolet B signal modulates tea flavor characteristics via shifting a metabolic flux in flavonoid biosynthesis. *Journal of Agricultural and Food Chemistry*, 69, 3401–3414. <https://doi.org/10.1021/acs.jafc.0c07009>
309. Palma, F. F., & C., Castro-Alves, V., Rosenqvist, E., Ottosen, C. O., Strid, Å., & Morales, L. O. (2021). Effects of UV radiation on transcript and metabolite accumulation are dependent on monochromatic light background in cucumber. *Physiologia Plantarum*, 173, 750–761. <https://doi.org/10.1111/ppl.13551>
310. Quintero-Arias, D. G., Acuña-Caita, J. F., Asensio, C., & Valenzuela, J. L. (2021). Ultraviolet transparency of plastic films determines the quality of lettuce (*Lactuca sativa*, L.) grown in a greenhouse. *Agronomy*, 11(2), 358. <https://doi.org/10.3390/agronomy11020358>
311. Ma, M., Wang, P., Yang, R., Zhou, T., & Gu, Z. (2019). UV-B mediates isoflavone accumulation and oxidative-antioxidant system responses in germinating soybean. *Food Chemistry*, 275, 628–636. <https://doi.org/10.1016/j.foodchem.2018.09.158>
312. Qi, W., Ma, J., Zhang, J., Gui, M., Li, J., & Zhang, L. (2020). Effects of low doses of UV-B radiation supplementation on tuber quality in purple potato (*Solanum tuberosum* L.). *Plant Signaling & Behavior*, 15(9), 1783490. <https://doi.org/10.1080/15592324.2020.1783490>
313. Santin, M., Castagna, A., Miras-Moreno, B., Rocchetti, G., Lucini, L., Hauser, M. T., & Ranieri, A. (2020). Beyond the visible and below the peel: How UV-B radiation influences the phenolic profile in the pulp of peach fruit A biochemical and molecular study. *Frontiers in Plant Science*, 11, 579063. <https://doi.org/10.3389/fpls.2020.579063>
314. Ferreyra, M. L. F., Serra, P., & Casati, P. (2021). Recent advances on the roles of flavonoids as plant protective molecules after UV and high light exposure. *Physiologia Plantarum*, 173(3), 736–749. <https://doi.org/10.1111/ppl.13543>
315. Brousseau, V. D., Wu, B.-S., MacPherson, S., Morello, V., & Lefsrud, M. (2021). Cannabinoids and terpenes: How production of photo-protectants can be manipulated to enhance *Cannabis sativa*. *Phytochemistry. Frontiers in Plant Science*, 12, 620021. <https://doi.org/10.3389/fpls.2021.620021>
316. Tripathi, D., Meena, R. P., & Pandey-Rai, S. (2021). Short term UV-B radiation mediated modulation of physiological traits and withanolides production in *Withania coagulans* (L.) Dunal under in-vitro condition. *Physiology and Molecular Biology of Plants*, 27(8), 1823–1835. <https://doi.org/10.1007/s12298-021-01046-7>
317. Zhong, Z., Liu, S., Han, S., Li, Y., Tao, M., Liu, A., He, Q., Chen, S., Dufresne, C., Zhu, W., & Tian, J. (2021). Integrative omic analysis reveals the improvement of alkaloid accumulation by ultraviolet-B radiation and its upstream regulation in *Catharanthus roseus*. *Industrial Crops and Products*, 166, 113448. <https://doi.org/10.1016/j.indcrop.2021.113448>
318. FAO (2020). Fruit and vegetables—your dietary essentials. The international year of fruits and vegetables, 2021, background paper. (pp. 82). Rome, Italy.
319. Keffie, T. S., Samuel, A., Woldegiorgis, A. Z., Lambert, C., Nohr, D., & Biesalski, H. K. (2021). Consumption of sun-exposed oyster mushrooms help patients fight tuberculosis. *Asian Journal of Complementary and Alternative Medicine. Volume*, 9(2).
320. Turck, D., Bohn, T., Castenmiller, J., De Henauw, S., Hirsch-Ernst, K. I., Maciuk, A., Mangelsdorf, I., McArdle, H. J., Naska, A., Pelaez, C., Pentieva, K., Siani, A., Thies, F., Tsaouri, S., Vinceti, M., Cubadda, F., Frenzel, T., Heinonen, M., Marchelli, R., ... Knutsen, H. K. (2022). Safety of vitamin D<sub>2</sub> mushroom powder as a Novel food pursuant to Regulation (EU) 2015/2283 (NF 2019/1471). *European Food Safety Authority Journal*, 20(6), e07326. <https://doi.org/10.2903/j.efsa.2022.7326>
321. Hamid, A., Singh, S., Agrawal, M., & Agrawal, S. B. (2019). *Heteropogon contortus* bl-1 (pilli grass) and elevated UV-B radiation: The role of growth, physiological, and biochemical traits in determining forage productivity and quality. *Photochemistry and Photobiology*, 95(2), 572–580. <https://doi.org/10.1111/php.12990>
322. Hamid, A., Singh, S., Agrawal, M., & Agrawal, S. B. (2020). Effects of plant age on performance of the tropical perennial fodder grass, *Cenchrus ciliaris* L. subjected to elevated ultraviolet-B radiation. *Plant Biology*, 22(5), 805–812. <https://doi.org/10.1111/plb.13116>
323. Golob, A., Stibilj, V., Kreft, I., Vogel-Mikus, K., Gaberscik, A., & Germ, M. (2018). Selenium treatment alters the effects of UV radiation on chemical and production parameters in hybrid buckwheat. *Acta Agriculturae Scandinavica Section B-Soil and Plant Science*, 68, 5–15.
324. FAO (2019). New standards to curb the global spread of plant pests and diseases. <http://www.fao.org/news/story/en/item/1187738/icode/>
325. Jones, R. A. C., & Naidu, R. A. (2019). Global dimensions of plant virus diseases: Current status and future perspectives. *Annual Review of Virology*, 6(1), 387–409. <https://doi.org/10.1146/annurev-virology-092818-015606>

326. Bebbler, D. P., Ramotowski, M. A., & Gurr, S. J. (2013). Crop pests and pathogens move polewards in a warming world. *Nature Climate Change*, 3(11), 985–988. <https://doi.org/10.1038/nclim ate1990>
327. Chaloner, T. M., Gurr, S. J., & Bebbler, D. P. (2021). Plant pathogen infection risk tracks global crop yields under climate change. *Nature Climate Change*, 11(8), 710–715. <https://doi.org/10.1038/s41558-021-01104-8>
328. Juroszek, P., Racca, P., Link, S., Farhumand, J., & Kleinhenz, B. (2020). Overview on the review articles published during the past 30 years relating to the potential climate change effects on plant pathogens and crop disease risks. *Plant Pathology*, 69(2), 179–193. <https://doi.org/10.1111/ppa.13119>
329. Pierik, R., & Ballaré, C. L. (2021). Control of plant growth and defense by photoreceptors: From mechanisms to opportunities in agriculture. *Molecular Plant*, 14(1), 61–76. <https://doi.org/10.1016/j.molp.2020.11.021>
330. Pérez-Hedo, M., Alonso-Valiente, M., Vacas, S., Gallego, C., Pons, C., Arbona, V., Rambla, J. L., Navarro-Llopis, V., Granell, A., & Urbaneja, A. (2021). Plant exposure to herbivore-induced plant volatiles: A sustainable approach through eliciting plant defenses. *Journal of Pest Science*, 94(4), 1221–1235. <https://doi.org/10.1007/s10340-021-01334-x>
331. Brunner-Mendoza, C., Reyes-Montes, M., & d. R., Moonjely, S., Bidochka, M. J., & Toriello, C. (2019). A review on the genus *Metarhizium* as an entomopathogenic microbial biocontrol agent with emphasis on its use and utility in Mexico. *Biocontrol Science and Technology*, 29(1), 83–102. <https://doi.org/10.1080/09583157.2018.1531111>
332. Tong, S. M., & Feng, M. G. (2022). Molecular basis and regulatory mechanisms underlying fungal insecticides' resistance to solar ultraviolet irradiation. *Pest Management Science*, 78(1), 30–42. <https://doi.org/10.1002/ps.6600>
333. Mann, A. J., & Davis, T. S. (2021). Entomopathogenic fungi to control bark beetles: A review of ecological recommendations. *Pest Management Science*, 77(9), 3841–3846. <https://doi.org/10.1002/ps.6364>
334. Sutanto, K. D., Husain, M., Rasool, K. G., Malik, A. F., Al-Qahtani, W. H., & Aldawood, A. S. (2022). Persistency of indigenous and exotic entomopathogenic fungi isolates under Ultraviolet B (UV-B) irradiation to enhance field application efficacy and obtain sustainable control of the red palm weevil. *Insects*, 13(1), 103. <https://doi.org/10.3390/insects13010103>
335. Bernardo, C., & d. C., Pereira-Junior, R. A., Luz, C., Mascarin, G. M., & Kamp Fernandes, E. K. (2020). Differential susceptibility of blastospores and aerial conidia of entomopathogenic fungi to heat and UV-B stresses. *Fungal Biology*, 124(8), 714–722. <https://doi.org/10.1016/j.funbio.2020.04.003>
336. Aphalo, P. J., Albert, A., Björn, L. O., McLeod, A., Robson, T. M., & Rosenqvist, E. (Eds.). (2012). *Beyond the visible: A handbook of best practice in plant UV photobiology* (COST Action FA0906 UV4growth). Helsinki, Finland <http://hdl.handle.net/10138/37558>: University of Helsinki, Department of Biosciences, Division of Plant Biology
337. Puglis, H. J., & Boone, M. D. (2011). Effects of technical-grade active ingredient vs commercial formulation of seven pesticides in the presence or absence of UV radiation on survival of green frog tadpoles. *Archives of Environmental Contamination and Toxicology*, 60(1), 145–155. <https://doi.org/10.1007/s00244-010-9528-z>
338. Kah, M., Walch, H., & Hofmann, T. (2018). Environmental fate of nanopesticides: Durability, sorption and photodegradation of nanoformulated clothianidin. *Environmental Science. Nano*, 5(4), 882–889. <https://doi.org/10.1039/c8en00038g>
339. Schwarzenbach, R. P., Gschwend, P. M., & Imboden, D. M. (2002). Direct Photolysis. In R. P. Schwarzenbach, P. M. Gschwend, & D. M. Imboden (Eds.), *Environmental Organic Chemistry* (pp. 611–654). John Wiley.
340. Schwarzenbach, R. P., Gschwend, P. M., & Imboden, D. M. (2002). Indirect Photolysis: Reactions with Photooxidants in Natural Waters and in the Atmosphere. In R. P. Schwarzenbach, P. M. Gschwend, & D. M. Imboden (Eds.), *Environmental Organic Chemistry* (pp. 655–686). John Wiley.
341. Su, L., Caywood, L. M., Sivey, J. D., & Dai, N. (2019). Sunlight photolysis of safener benoxacor and herbicide metolachlor as mixtures on simulated soil surfaces. *Environmental Science & Technology*, 53(12), 6784–6793. <https://doi.org/10.1021/acs.est.9b01243>
342. Anderson, S. C., Chu, L., Bouma, C., Beukelman, L., McLouth, R., Larson, E., & Nienow, A. M. (2019). Comparison of the photodegradation of imazethapyr in aqueous solution, on epicuticular waxes, and on intact corn (*Zea mays*) and soybean (*Glycine max*) leaves. *Journal of Environmental Science and Health, Part B*, 54(2), 129–137. <https://doi.org/10.1080/03601234.2018.1511400>
343. Buyl Lee, S., & Chung Suh, M. (2021). Regulatory mechanisms underlying cuticular wax biosynthesis. *Journal of Experimental Botany*, 73(9), 2799–2816. <https://doi.org/10.1093/jxb/erab509>
344. Arbid, Y., Sleiman, M., & Richard, C. (2022). Photochemical interactions between pesticides and plant volatiles. *Science of the Total Environment*, 807(1), 150716. <https://doi.org/10.1016/j.scitotenv.2021.150716>
345. Palma, D., Arbid, Y., Sleiman, M., de Sainte-Claire, P., & Richard, C. (2020). New route to toxic nitro and nitroso products upon irradiation of micropollutant mixtures containing imidacloprid: Role of NOx and effect of natural organic matter. *Environmental Science & Technology*, 54(6), 3325–3333. <https://doi.org/10.1021/acs.est.9b07304>
346. Xi, N., Li, Y., Chen, J., Yang, Y., Duan, J., & Xia, X. (2021). Elevated temperatures decrease the photodegradation rate of pyrethroid insecticides on spinach leaves: Implications for the effect of climate warming. *Environmental Science & Technology*, 55(2), 1167–1177. <https://doi.org/10.1021/acs.est.0c06959>
347. Qu, S., Kolodziej, E. P., Long, S. A., Gloer, J. B., Patterson, E. V., Baltrusaitis, J., Jones, G. D., Benchetler, P. V., Cole, E. A., Kimbrough, K. C., Tarnoff, M. D., & Cwierny, D. M. (2013). Product-to-parent reversion of Trenbolone: Unrecognized risks for endocrine disruption. *Science*, 342(6156), 347–351. <https://doi.org/10.1126/science.1243192>
348. Mishra, P., Tyagi, B. K., Mukherjee, A., & Chandrasekaran, N. (2019). Potential application of nanopesticides to pest control in the food and agriculture sector. In *Food Applications of Nanotechnology* (pp. 493–509): CRC Press.
349. Côa, F., Bortolozzo, L. S., Petry, R., Da Silva, G. H., Martins, C. H. Z., de Medeiros, A. M. Z., Sabino, C. M. S., Costa, R. S., Khan, L. U., Delite, F. S., & Martinez, D. S. T. (2020). Environmental toxicity of nanopesticides against non-target organisms: The state of the art. In: L. F. Fraceto, V. L. S.S. de Castro, R. Grillo, D. Ávila, H. Caixeta Oliveira, & R. Lima (Eds.), *Nano-pesticides: From research and development to mechanisms of action and sustainable use in agriculture* (pp. 227–279). Cham: Springer International Publishing. [https://doi.org/10.1007/978-3-030-44873-8\\_8](https://doi.org/10.1007/978-3-030-44873-8_8)
350. Mishra, P., Dutta, S., Haldar, M., Dey, P., Kumar, D., Mukherjee, A., & Chandrasekaran, N. (2019). Enhanced mosquitocidal efficacy of colloidal dispersion of pyrethroid nanometric emulsion with benignity towards non-target species. *Ecotoxicology and Environmental Safety*, 176, 258–269. <https://doi.org/10.1016/j.ecoenv.2019.03.096>
351. Wang, D., Saleh, N. B., Byro, A., Zepp, R., Sahle-Demessie, E., Luxton, T. P., Ho, K. T., Burgess, R. M., Flury, M., White, J. C., & Su, C. (2022). Nano-enabled pesticides for sustainable



- agriculture and global food security. *Nature Nanotechnology*, 17(4), 347–360. <https://doi.org/10.1038/s41565-022-01082-8>
352. Kumar, S., Nehra, M., Dilbaghi, N., Marrazza, G., Hassan, A. A., & Kim, K.-H. (2019). Nano-based smart pesticide formulations: Emerging opportunities for agriculture. *Journal of Controlled Release*, 294, 131–153. <https://doi.org/10.1016/j.jconrel.2018.12.012>
  353. Teramura, A. H. (1983). Effects of ultraviolet-B radiation on the growth and yield of crop plants. *Physiologia Plantarum*, 58, 415–427. <https://doi.org/10.1111/j.1399-3054.1983.tb04203.x>
  354. Takshak, S., & Agrawal, S. B. (2019). Defense potential of secondary metabolites in medicinal plants under UV-B stress. *Journal of Photochemistry & Photobiology, B: Biology*, 193, 51–88. <https://doi.org/10.1016/j.jphotobiol.2019.02.002>
  355. Jiang, Q. Y., Zhang, M., & Mujumdar, A. S. (2020). UV induced conversion during drying of ergosterol to vitamin D in various mushrooms: Effect of different drying conditions. *Trends in Food Science & Technology*, 105, 200–210. <https://doi.org/10.1016/j.tifs.2020.09.011>
  356. Leung, M. F., & Cheung, P. C. K. (2021). Vitamins D and D<sub>2</sub> in cultivated mushrooms under ultraviolet irradiation and their bioavailability in humans: A mini-review. *International Journal of Medicinal Mushrooms*, 23, 1–15. <https://doi.org/10.1615/IntJMedMushrooms.2021040390>
  357. Ljubic, A., Thulesen, E. T., Jacobsen, C., & Jakobsen, J. (2021). UVB exposure stimulates production of vitamin D<sub>3</sub> in selected microalgae. *Algal Research-Biomass Biofuels and Bioproducts*, 59, 102472. <https://doi.org/10.1016/j.algal.2021.102472>
  358. Cardwell, G., Bornman, J. F., James, A. P., & Black, L. J. (2018). A review of mushrooms as a potential source of dietary vitamin D. *Nutrients*, 10(10), 1498. <https://doi.org/10.3390/nu10101498>
  359. Mariz-Ponte, N., Martins, S., Goncalves, A., Correia, C. M., Ribeiro, C., Dias, M. C., & Santos, C. (2019). The potential use of the UV-A and UV-B to improve tomato quality and preference for consumers. *Scientia Horticulturae*, 246, 777–784. <https://doi.org/10.1016/j.scienta.2018.11.058>
  360. Muller, R., Acosta-Motos, J. R., Grosskinsky, D. K., Hernandez, J. A., Lutken, H., & Barba-Espin, G. (2019). UV-B exposure of black carrot (*Daucus carota* ssp. *sativus* var. *atrorubens*) plants promotes growth, accumulation of anthocyanin, and phenolic compounds. *Agronomy-Basel*, 9, 323. <https://doi.org/10.3390/agronomy9060323>
  361. Del-Castillo-Alonso, M. Á., Monforte, L., Tomás-Las-Heras, R., Núñez-Olivera, E., & Martínez-Abaigar, J. (2020). A supplement of ultraviolet-B radiation under field conditions increases phenolic and volatile compounds of Tempranillo grape skins and the resulting wines. *European Journal of Agronomy*, 121, 126150. <https://doi.org/10.1016/j.eja.2020.126150>
  362. Del-Castillo-Alonso, M. A., Monforte, L., Tomas-Las-Heras, R., Ranieri, A., Castagna, A., Martinez-Abaigar, J., & Nunez-Olivera, E. (2021). Secondary metabolites and related genes in *Vitis vinifera* L. cv. Tempranillo grapes as influenced by ultraviolet radiation and berry development. *Physiologia Plantarum*. <https://doi.org/10.1111/ppl.13483>
  363. Santin, M., Lucini, L., Castagna, A., Rocchetti, G., Hauser, M. T., & Ranieri, A. (2019). Comparative “phenol-omics” and gene expression analyses in peach (*Prunus persica*) skin in response to different postharvest UV-B treatments. *Plant Physiology and Biochemistry*, 135, 511–519. <https://doi.org/10.1016/j.plaphy.2018.11.009>
  364. Ding, R. R., Che, X. K., Shen, Z., & Zhang, Y. H. (2021). Metabolome and transcriptome profiling provide insights into green apple peel reveals light- and UV-B-responsive pathway in anthocyanins accumulation. *BMC Plant Biology*, 21, 351. <https://doi.org/10.1186/s12870-021-03121-3>
  365. Li, T. S., Yamane, H., & Tao, R. (2021). Preharvest long-term exposure to UV-B radiation promotes fruit ripening and modifies stage-specific anthocyanin metabolism in highbush blueberry. *Horticulture Research*, 8, 67. <https://doi.org/10.1038/s41438-021-00503-4>
  366. Thomas, T. T. D., & Puthur, J. T. (2020). UV-B priming enhances specific secondary metabolites in *Oryza sativa* (L.) empowering to encounter diverse abiotic stresses. *Plant Growth Regulation*, 92(2), 169–180. <https://doi.org/10.1007/s10725-020-00628-x>
  367. Jiang, Z. F., Xu, M. F., Dong, J. F., Zhu, Y., Lou, P. P., Han, Y. D., Hao, J., Yang, Y. J., Ni, J., & Xu, M. J. (2022). UV-B pre-irradiation induces cold tolerance in tomato fruit by SIUVR8-mediated upregulation of superoxide dismutase and catalase. *Postharvest Biology and Technology*, 185, 111777. <https://doi.org/10.1016/j.postharvbio.2021.111777>
  368. Neugart, S., & Bumke-Vogt, C. (2021). Flavonoid glycosides in *Brassica* species respond to UV-B depending on exposure time and adaptation time. *Molecules*, 26, 494. <https://doi.org/10.3390/molecules26020494>
  369. Rady, M. R., Gierczik, K., Ibrahim, M. M., Matter, M. A., & Galiba, G. (2021). Anticancer compounds production in *Catharanthus roseus* by methyl jasmonate and UV-B elicitation. *South African Journal of Botany*, 142, 34–41. <https://doi.org/10.1016/j.sajb.2021.05.024>
  370. Santin, M., Giordani, T., Cavallini, A., Bernardi, R., Castagna, A., Hauser, M. T., & Ranieri, A. (2019). UV-B exposure reduces the activity of several cell wall-dismantling enzymes and affects the expression of their biosynthetic genes in peach fruit (*Prunus persica* L., cv. Fairtime, melting phenotype). *Photochemical and Photobiological Sciences*, 18, 1280–1289. <https://doi.org/10.1039/c8pp00505b>
  371. Yoon, H. I., Kim, D., & Son, J. E. (2020). Spatial and temporal bioactive compound contents and chlorophyll fluorescence of kale (*Brassica oleracea* L.) under UV-B exposure near harvest time in controlled environments. [Article; Early Access]. *Photochemistry and Photobiology*, 96, 845–852. <https://doi.org/10.1111/php.13237>
  372. Castillejo, N., Martinez-Zamora, L., & Artes-Hernandez, F. (2021). Periodical UV-B radiation hormesis in biosynthesis of kale sprouts nutraceuticals. *Plant Physiology and Biochemistry*, 165, 274–285. <https://doi.org/10.1016/j.plaphy.2021.05.022>
  373. Virsile, A., Samuoliene, G., Miliauskienė, J., & Duchovskis, P. (2019). Applications and advances in LEDs for horticulture and crop production. In: T. Koutchma (Ed.), *Ultraviolet LED Technology for Food Applications. From Farms to Kitchens* (pp 35–65). London, Elsevier. <https://doi.org/10.1016/B978-0-12-817794-5.00003-0>
  374. Khan, S., Dar, A. H., Shams, R., Aga, M. B., Siddiqui, M. W., Mir, S. A., & Rizvi, Q.-U.-e. H., Khan, S. A., & Altaf, A. (2021). Applications of ultraviolet light-emitting diode technology in horticultural produce: A systematic review and meta-analysis. *Food and Bioprocess Technology*, 15, 487–497. <https://doi.org/10.1007/s11947-021-02742-8>
  375. Yoon, H. I., Kim, H. Y., Kim, J., Oh, M.-M., & Son, J. E. (2021). Quantitative analysis of UV-B radiation interception in 3d plant structures and intraindividual distribution of phenolic contents. *International Journal of Molecular Sciences*, 12, 667456. <https://doi.org/10.3389/fpls.2021.667456>
  376. McLay, E. R., Pontaroli, A. C., & Wargent, J. J. (2020). UV-B induced flavonoids contribute to reduced biotrophic disease susceptibility in lettuce seedlings. *Frontiers in Plant Science*, 11, 594681. <https://doi.org/10.3389/fpls.2020.594681>
  377. Strauss, J., Schirrmeyer, L., Grosse, G., Fortier, D., Hugelius, G., Knoblauch, C., Romanovsky, V., Schädel, C., Schneider von Deimling, T., Schuur, E. A. G., Shmelev, D., Ulrich, M., & Veremeeva, A. (2017). Deep Yedoma permafrost: A synthesis



- of depositional characteristics and carbon vulnerability. *Earth-Science Reviews*, 172, 75–86. <https://doi.org/10.1016/j.earscirev.2017.07.007>
378. Hugelius, G., Strauss, J., Zubrzycki, S., Harden, J. W., Schuur, E. A. G., Ping, C. L., Schirrmeister, L., Grosse, G., Michaelson, G. J., Koven, C. D., O'Donnell, J. A., Elberling, B., Mishra, U., Camill, P., Yu, Z., Palmtag, J., & Kuhry, P. (2014). Estimated stocks of circumpolar permafrost carbon with quantified uncertainty ranges and identified data gaps. *Biogeosciences*, 11(23), 6573–6593. <https://doi.org/10.5194/bg-11-6573-2014>
  379. Biskaborn, B. K., Smith, S. L., Noetzli, J., Matthes, H., Vieira, G., Streletskiy, D. A., Schoeneich, P., Romanovsky, V. E., Lewkowicz, A. G., Abramov, A., Allard, M., Boike, J., Cable, W. L., Christiansen, H. H., Delaloye, R., Diekmann, B., Drozdov, D., Etzelmüller, B., Grosse, G., ... Lantuit, H. (2019). Permafrost is warming at a global scale. *Nature Communications*, 10(1), 264. <https://doi.org/10.1038/s41467-018-08240-4>
  380. IPCC (2019). IPCC Special Report on the Ocean and Cryosphere in a Changing Climate Summary for Policymakers. In: H.-O. Pörtner, D. C. Roberts, V. Masson-Delmotte, P. C. Zhai, M. Tignor, E. Poloczanska, et al. (Eds.). Cambridge, UK and New York, NY, USA. <https://doi.org/10.1017/9781009157964.001>
  381. Cory, R. M., Ward, C. P., Crump, B. C., & Kling, G. W. (2014). Sunlight controls water column processing of carbon in arctic fresh waters. *Science*, 345(6199), 925–928. <https://doi.org/10.1126/science.125311>
  382. Cory, R. M., Crump, B. C., Dobkowski, J. A., & Kling, G. W. (2013). Surface exposure to sunlight stimulates CO<sub>2</sub> release from permafrost soil carbon in the Arctic. *Proceedings of the National Academy of Sciences of the United States of America*, 110(9), 3429–3434. <https://doi.org/10.1073/pnas.1214104110>
  383. Turetsky, M. R., Abbott, B. W., Jones, M. C., Anthony, K. W., Olefeldt, D., Schuur, E. A. G., Koven, C. D., McGuire, A. D., Grosse, G., & Kuhry, P. (2019). Permafrost collapse is accelerating carbon release. *Nature*, 569, 32–34.
  384. Austin, A. T., & Ballaré, C. L. (2010). Dual role of lignin in plant litter decomposition in terrestrial ecosystems. *Proceedings of the National Academy of Sciences USA*, 107(10), 4618–4622. <https://doi.org/10.1073/pnas.0909396107>
  385. Day, T. A., & Bliss, M. S. (2020). Solar photochemical emission of CO<sub>2</sub> from leaf litter: Sources and significance to C loss. *Ecosystems*, 23, 1344–1361. <https://doi.org/10.1007/s10021-019-00473-8>
  386. King, J. Y., Brandt, L. A., & Adair, E. C. (2012). Shedding light on plant litter decomposition: Advances, implications and new directions in understanding the role of photodegradation. *Biogeochemistry*, 111(1–3), 57–81. <https://doi.org/10.1007/s10533-012-9737-9>
  387. Keiser, A. D., Warren, R., Filley, T., & Bradford, M. A. (2021). Signatures of an abiotic decomposition pathway in temperate forest leaf litter. *Biogeochemistry*, 153(2), 177–190. <https://doi.org/10.1007/s10533-021-00777-9>
  388. Day, T. A., Urbine, J. M., & Bliss, M. S. (2022). Supplemental precipitation accelerates decay but only in photodegraded litter and implications that sunlight promotes leaching loss. *Biogeochemistry*, 158(1), 113–129. <https://doi.org/10.1007/s10533-022-00888-x>
  389. Logan, J. R., Barnes, P., & Evans, S. E. (2022). Photodegradation of plant litter cuticles enhances microbial decomposition by increasing uptake of non-rainfall moisture. *Functional Ecology*, 36, 1727–1738. <https://doi.org/10.1111/1365-2435.14053>
  390. Barnes, P. W., Throop, H. L., Archer, S. R., Breshears, D. D., McCulley, R. L., & Tobler, M. A. (2015). Sunlight and soil-litter mixing: Drivers of litter decomposition in drylands. *Progress in Botany*, 76, 273–302. [https://doi.org/10.1007/978-3-319-08807-5\\_11](https://doi.org/10.1007/978-3-319-08807-5_11)
  391. Austin, A. T., Méndez, M. S., & Ballaré, C. L. (2016). Photodegradation alleviates the lignin bottleneck for carbon turnover in terrestrial ecosystems. *Proceedings of the National Academy of Sciences USA*, 113(16), 4392–4397. <https://doi.org/10.1073/pnas.1516157113>
  392. Pieristè, M., Forey, E., Lounès-Hadj Sahraoui, A., Megloul, H., Laruelle, F., Delporte, P., Robson, T. M., & Chauvat, M. (2020). Spectral composition of sunlight affects the microbial functional structure of Beech leaf litter during the initial phase of decomposition. *Plant and Soil*. <https://doi.org/10.1007/s11104-020-04557-6>
  393. Cuadra, P., Guajardo, J., Carrasco-Orellana, C., Stappung, Y., Fajardo, V., & Herrera, R. (2020). Differential expression after UV-B radiation and characterization of chalcone synthase from the Patagonian hairgrass *Deschampsia antarctica*. *Phytochemistry*, 169, 112179. <https://doi.org/10.1016/j.phytochem.2019.112179>
  394. Ruhland, C. T., & Niere, J. A. (2019). The effects of surface albedo and initial lignin concentration on photodegradation of two varieties of *Sorghum bicolor* litter. *Scientific Reports*, 9(1), 18748. <https://doi.org/10.1038/s41598-019-55272-x>
  395. Wang, Q.-W., Pieristè, M., Liu, C., Kenta, T., Robson, M. T., & Kurokawa, H. (2021). The contribution of photodegradation to litter decomposition in a temperate forest gap and understorey. *New Phytologist*, 229(5), 2625–2636. <https://doi.org/10.1111/nph.17022>
  396. Pieristè, M., Neimane, S., Solanki, T., Nybakken, L., Jones, A. G., Forey, E., Chauvat, M., Nečajeva, J., & Robson, T. M. (2020). Ultraviolet radiation accelerates photodegradation under controlled conditions but slows the decomposition of senescent leaves from forest stands in southern Finland. *Plant Physiology and Biochemistry*, 146, 42–54. <https://doi.org/10.1016/j.plaphy.2019.11.005>
  397. Araujo, P. I., Grasso, A. A., González-Arzac, A., Méndez, M. S., & Austin, A. T. (2022). Sunlight and soil biota accelerate decomposition of crop residues in the Argentine Pampas. *Agriculture Ecosystems & Environment*, 330, 107908. <https://doi.org/10.1016/j.agee.2022.107908>
  398. Liu, G. F., Wang, L., Jiang, L., Pan, X., Huang, Z. Y., Dong, M., & Cornelissen, J. H. C. (2018). Specific leaf area predicts dryland litter decomposition via two mechanisms. *Journal of Ecology*, 106(1), 218–229. <https://doi.org/10.1111/1365-2745.12868>
  399. Erdenebileg, E., Wang, C., Ye, X., Cui, Q., Du, J., Huang, Z., Liu, G., & Cornelissen, J. H. (2020). Multiple abiotic and biotic drivers of long-term wood decomposition within and among species in the semiarid inland dunes: A dual role for stem diameter. *Functional Ecology*. <https://doi.org/10.1111/1365-2435.13559>
  400. Esch, E. H., King, J. Y., & Cleland, E. E. (2019). Foliar litter chemistry mediates susceptibility to UV degradation in two dominant species from a semi-arid ecosystem. *Plant and Soil*, 440(1–2), 265–276. <https://doi.org/10.1007/s11104-019-04069-y>
  401. Berenstecher, P., Vivanco, L., Pérez, L. I., Ballaré, C. L., & Austin, A. (2020). Sunlight doubles aboveground carbon loss in a seasonally dry woodland in Patagonia. *Current Biology*. <https://doi.org/10.1016/j.cub.2020.06.005>
  402. Méndez, M. S., Martinez, M. L., Araujo, P. I., & Austin, A. T. (2019). Solar radiation exposure accelerates decomposition and biotic activity in surface litter but not soil in a semiarid woodland ecosystem in Patagonia Argentina. *Plant and Soil*. <https://doi.org/10.1007/s11104-019-04325-1>
  403. Asao, S., Parton, W. J., Chen, M. S., & Gao, W. (2018). Photodegradation accelerates ecosystem N cycling in a simulated California grassland. *Ecosphere*, 9(8), e02370. <https://doi.org/10.1002/ecs2.2370>
  404. Day, T. A., & Bliss, M. S. (2019). A spectral weighting function for abiotic photodegradation based on photochemical emission

- of CO<sub>2</sub> from leaf litter in sunlight. *Biogeochemistry*, 146(2), 173–190. <https://doi.org/10.1007/s10533-019-00616-y>
405. Logan, J. R., Jacobson, K. M., Jacobson, P. J., & Evans, S. E. (2021). Fungal communities on standing litter are structured by moisture type and constrain decomposition in a hyper-arid grassland. *Frontiers in Microbiology*, 12, 596517. <https://doi.org/10.3389/fmicb.2021.596517>
  406. Ball, B. A., Christman, M. P., & Hall, S. J. (2019). Nutrient dynamics during photodegradation of plant litter in the Sonoran desert. *Journal of Arid Environments*, 160, 1–10. <https://doi.org/10.1016/j.jaridenv.2018.09.004>
  407. Marinho, O. A., Martinelli, L. A., Duarte-Neto, P. J., Mazzi, E. A., & King, J. Y. (2020). Photodegradation influences litter decomposition rate in a humid tropical ecosystem Brazil. *Science of the Total Environment*, 715, 136601. <https://doi.org/10.1016/j.scitotenv.2020.136601>
  408. Wu, C. S., Wang, H. K., Mo, Q. F., Zhang, Z. J., Huang, G. X., Kong, F. Q., Liu, Y. Q., & Wang, G. G. (2019). Effects of elevated UV-B radiation and N deposition on the decomposition of coarse woody debris. *Science of the Total Environment*, 663, 170–176. <https://doi.org/10.1016/j.scitotenv.2019.01.271>
  409. Wei, B., Zhang, D., Kou, D., Yang, G., Liu, F., Peng, Y., & Yang, Y. (2022). Decreased ultraviolet radiation and decomposer biodiversity inhibit litter decomposition under continuous nitrogen inputs. *Functional Ecology*, 36, 998–1009. <https://doi.org/10.1111/1365-2435.14015>
  410. Shelton, S., Neale, P., Pinsonneault, A., & Tzortziou, M. (2022). Biodegradation and photodegradation of vegetation-derived dissolved organic matter in tidal marsh ecosystems. *Estuaries and Coasts*, 45, 1324–1342. <https://doi.org/10.1007/s12237-021-00982-7>
  411. Pieristè, M., Chauvat, M., Kotilainen, T. K., Jones, A. G., Aubert, M., Robson, M. T., & Forey, E. (2019). Solar UV-A radiation and blue light enhance tree leaf litter decomposition in a temperate forest. *Oecologia*, 191(1), 191–203. <https://doi.org/10.1007/s00442-019-04478-x>
  412. Austin, A. T., & Vivanco, L. (2006). Plant litter decomposition in a semi-arid ecosystem controlled by photodegradation. *Nature*, 442, 555–558. <https://doi.org/10.1038/nature05038>
  413. Yan, W., Shangguan, Z., & Zhong, Y. (2021). Responses of mass loss and nutrient release in litter decomposition to ultraviolet radiation. *Journal of Soils and Sediments*, 21(2), 698–704. <https://doi.org/10.1007/s11368-020-02810-0>
  414. Wang, Q.-W., Pieristè, M., Kotilainen, T., Forey, E., Chauvat, M., Kurokawa, H., Robson, M., & Jones, A. (2022) Meta-analysis of ecological studies attenuating solar radiation illustrates the importance of blue light over ultraviolet radiation in driving photodegradation of litter in terrestrial ecosystems. Doi:<https://doi.org/10.21203/rs.3.rs-1377521/v1>.
  415. Wang, Q.-W., Robson, T. M., Pieristè, M., Kenta, T., Zhou, W., & Kurokawa, H. (2022). Canopy structure and phenology modulate the impacts of solar radiation on C and N dynamics during litter decomposition in a temperate forest. *Science of the Total Environment*, 820, 153185. <https://doi.org/10.1016/j.scitotenv.2022.153185>
  416. Landuyt, D., Ampoorter, E., Bastias, C. C., Benavides, R., & Müller, S. (2020). Importance of overstorey attributes for understorey litter production and nutrient cycling in European forests. *Forest Ecosystems*, 7, 45. <https://doi.org/10.1186/s40663-020-00256-x>
  417. Berenstecher, P., Araujo, P. I., & Austin, A. T. (2021). Worlds apart: Location above-or below-ground determines plant litter decomposition in a semi-arid Patagonian steppe. *Journal of Ecology*, 109(8), 2885–2896. <https://doi.org/10.1111/1365-2745.13688>
  418. Predick, K. I., Archer, S. R., Aguillon, S. M., Keller, D. A., Throop, H. L., & Barnes, P. W. (2018). UV-B radiation and shrub canopy effects on surface litter decomposition in a shrub-invaded dry grassland. *Journal of Arid Environments*, 157, 13–21. <https://doi.org/10.1016/j.jaridenv.2018.06.007>
  419. Barnes, P. W., Throop, H. L., Hewins, D. B., Abbene, M. L., & Archer, S. R. (2012). Soil coverage reduces photodegradation and promotes the development of soil-microbial films on dryland leaf litter. *Ecosystems*, 15(2), 311–321. <https://doi.org/10.1007/s10021-011-9511-1>
  420. Lee, H.-S., Hur, J., Lee, M.-H., Brogi, S. R., Kim, T.-W., & Shin, H.-S. (2019). Photochemical release of dissolved organic matter from particulate organic matter: Spectroscopic characteristics and disinfection by-product formation potential. *Chemosphere*, 235, 586–595. <https://doi.org/10.1016/j.chemosphere.2019.06.127>
  421. Saunio, M., Stavert, A. R., Poulter, B., Bousquet, P., Canadell, J. G., Jackson, R. B., Raymond, P. A., Dlugokencky, E. J., Houweling, S., Patra, P. K., Ciais, P., Arora, V. K., Bastviken, D., Bergamaschi, P., Blake, D. R., Brailsford, G., Bruhwiler, L., Carlson, K. M., Carrol, M., ... Zhuang, Q. (2020). The global methane budget 2000–2017. *Earth System Science Data*, 12(3), 1561–1623. <https://doi.org/10.5194/essd-12-1561-2020>
  422. Jackson, R. B., Saunio, M., Bousquet, P., Canadell, J. G., Poulter, B., Stavert, A. R., Bergamaschi, P., Niwa, Y., Segers, A., & Tsuruta, A. (2020). Increasing anthropogenic methane emissions arise equally from agricultural and fossil fuel sources. *Environmental Research Letters*. <https://doi.org/10.1088/1748-9326/ab9ed2>
  423. Dean, J. F., Middelburg, J. J., Röckmann, T., Aerts, R., Blauw, L. G., Egger, M., Jetten, M. S. M., de Jong, A. E. E., Meisel, O. H., Rasigraf, O., Slomp, C. P., & in't Zandt, M. H., & Dolman, A. J. (2018). Methane feedbacks to the global climate system in a warmer world. *Reviews of Geophysics*, 56(1), 207–250. <https://doi.org/10.1002/2017RG000559>
  424. Covey, K. R., & Megonigal, J. P. (2019). Methane production and emissions in trees and forests. *New Phytologist*, 222(1), 35–51. <https://doi.org/10.1111/nph.15624>
  425. Pitz, S. L., Megonigal, J. P., Chang, C.-H., & Szlavecz, K. (2018). Methane fluxes from tree stems and soils along a habitat gradient. *Biogeochemistry*, 137(3), 307–320. <https://doi.org/10.1007/s10533-017-0400-3>
  426. Fraser, W. T., Blei, E., Fry, S. C., Newman, M. F., Reay, D. S., Smith, K. A., & McLeod, A. R. (2015). Emission of methane, carbon monoxide, carbon dioxide and short-chain hydrocarbons from vegetation foliage under ultraviolet irradiation. *Plant, Cell & Environment*, 38(5), 980–989. <https://doi.org/10.1111/pce.12489>
  427. Barba, J., Poyatos, R., & Vargas, R. (2019). Automated measurements of greenhouse gases fluxes from tree stems and soils: Magnitudes, patterns and drivers. *Scientific Reports*, 9(1), 4005. <https://doi.org/10.1038/s41598-019-39663-8>
  428. Tenhoviirta, S. A. M., Kohl, L., Koskinen, M., Patama, M., Lintunen, A., Zanetti, A., Lilja, R., & Pihlatie, M. (2022). Solar radiation drives methane emissions from the shoots of Scots pine. *New Phytologist*. <https://doi.org/10.1111/nph.18120>
  429. McLeod, A. R., Fry, S. C., Loake, G. J., Messinger, D. J., Reay, D. S., Smith, K. A., & Yun, B.-W. (2008). Ultraviolet radiation drives methane emissions from terrestrial plant pectins. *New Phytologist*, 180(1), 124–132. <https://doi.org/10.1111/j.1469-8137.2008.02571.x>
  430. Yip, D. Z., Veach, A. M., Yang, Z. K., Cregger, M. A., & Schadt, C. W. (2019). Methanogenic Archaea dominate mature heartwood habitats of eastern cottonwood (*Populus deltoides*). *New Phytologist*, 222(1), 115–121. <https://doi.org/10.1111/nph.15346>

431. Li, H.-L., Zhang, X.-M., Deng, F.-D., Han, X.-G., Xiao, C.-W., Han, S.-J., & Wang, Z.-P. (2019). Microbial methane production is affected by secondary metabolites in the heartwood of living trees in upland forests. *Trees*, 34(1), 243–254. <https://doi.org/10.1007/s00468-019-01914-6>
432. Feng, H., Guo, J., Han, M., Wang, W., Peng, C., Jin, J., Song, X., & Yu, S. (2020). A review of the mechanisms and controlling factors of methane dynamics in forest ecosystems. *Forest Ecology and Management*. <https://doi.org/10.1016/j.foreco.2019.117702>
433. Abatzoglou, J. T., Williams, A. P., Boschetti, L., Zubkova, M., & Kolden, C. A. (2018). Global patterns of interannual climate–fire relationships. *Global Change Biology*, 24(11), 5164–5175. <https://doi.org/10.1111/gcb.14405>
434. Andreae, M. O. (2019). Emission of trace gases and aerosols from biomass burning – an updated assessment. *Atmosphere Chemistry and Physics*, 19, 8523–8546. <https://doi.org/10.5194/acp-19-8523-2019>
435. Hart, S. J., Henkelman, J., McLoughlin, P. D., Nielsen, S. E., Truchon-Savard, A., & Johnstone, J. F. (2019). Examining forest resilience to changing fire frequency in a fire-prone region of boreal forest. *Global Change Biology*, 25(3), 869–884. <https://doi.org/10.1111/gcb.14550>
436. Jones, M. W., Santín, C., van der Werf, G. R., & Doerr, S. H. (2019). Global fire emissions buffered by the production of pyrogenic carbon. *Nature Geoscience*, 12(9), 742–747. <https://doi.org/10.1038/s41561-019-0403-x>
437. Rumpel, C. (2019). Soils linked to climate change. *Nature*, 572(7770), 442–443. <https://doi.org/10.1038/d41586-019-02450-6>
438. Walker, X. J., Baltzer, J. L., Cumming, S. G., Day, N. J., Ebert, C., Goetz, S., Johnstone, J. F., Potter, S., Rogers, B. M., & Schuur, E. A. G. (2019). Increasing wildfires threaten historic carbon sink of boreal forest soils. *Nature*, 572(7770), 520–523. <https://doi.org/10.1038/s41586-019-1474-y>
439. Williams, A. P., & Abatzoglou, J. T. (2016). Recent advances and remaining uncertainties in resolving past and future climate effects on global fire activity. *Current Climate Change Reports*, 2(1), 1–14. <https://doi.org/10.1007/s40641-016-0031-0>
440. Wagner, S., Jaffé, R., & Stubbins, A. (2018). Dissolved black carbon in aquatic ecosystems. *Limnology Oceanography Letters*, 3(3), 168–185. <https://doi.org/10.1002/lol2.10076>
441. Bostick, K. W., Zimmerman, A. R., Wozniak, A. S., Mitra, S., & Hatcher, P. G. (2018). Production and composition of pyrogenic dissolved organic matter from a logical series of laboratory-generated chars. *Frontiers in Earth Science*. <https://doi.org/10.3389/feart.2018.00043>
442. Hohner, A. K., Rhoades, C. C., Wilkerson, P., & Rosario-Ortiz, F. L. (2019). Wildfires alter forest watersheds and threaten drinking water quality. *Accounts of Chemical Research*, 52(5), 1234–1244. <https://doi.org/10.1021/acs.accounts.8b00670>
443. Goranov, A. I., Wozniak, A. S., Bostick, K. W., Zimmerman, A. R., Mitra, S., & Hatcher, P. G. (2022). Microbial labilization and diversification of pyrogenic dissolved organic matter. *Biogeosciences*, 19(5), 1491–1514. <https://doi.org/10.5194/bg-19-1491-2022>
444. Rust, A. J., Hogue, T. S., Saxe, S., & McCray, J. (2018). Post-fire water-quality response in the western United States. *International Journal of Wildland Fire*, 27(3), 203–216. <https://doi.org/10.1071/WF17115>
445. Wagner, S., Coppola, A. I., Stubbins, A., Dittmar, T., Niggemann, J., Drake, T. W., Seidel, M., Spencer, R. G. M., & Bao, H. (2021). Questions remain about the biolability of dissolved black carbon along the combustion continuum. *Nature Communications*, 12(1), 4281. <https://doi.org/10.1038/s41467-021-24477-y>
446. Bostick, K. W., Zimmerman, A. R., Goranov, A. I., Mitra, S., Hatcher, P. G., & Wozniak, A. S. (2021) Biolability of fresh and photodegraded pyrogenic dissolved organic matter from laboratory prepared chars. *Journal of Geophysical Research: Biogeosciences*. Doi: <https://doi.org/10.1029/2020JG005981>
447. Bostick, K. W., Zimmerman, A. R., Goranov, A. I., Mitra, S., Hatcher, P. G., & Wozniak, A. S. (2020). Photolability of pyrogenic dissolved organic matter from a thermal series of laboratory-prepared chars. *Science of the Total Environment*, 724, 138198. <https://doi.org/10.1016/j.scitotenv.2020.138198>
448. Tilmes, S., Kinnison, D. E., Garcia, R. R., Salawitch, R., Canty, T., Lee-Taylor, J., Madronich, S., & Chance, K. (2012). Impact of very short-lived halogens on stratospheric ozone abundance and UV radiation in a geo-engineered atmosphere. *Atmospheric Chemistry and Physics*, 12(22), 10945–10955. <https://doi.org/10.5194/acp-12-10945-2012>
449. Flint, S. D., Ryel, R. J., & Caldwell, M. M. (2003). Ecosystem UV-B experiments in terrestrial communities: A review of recent findings and methodologies. *Agricultural and Forest Meteorology*, 120(1–4), 177–189. <https://doi.org/10.1016/j.agrformet.2003.08.014>
450. Caldwell, M. M., & Flint, S. D. (2006). Use and evaluation of biological spectral UV weighting functions for the ozone reduction issue. In F. Ghatti, G. Checcucci, & J. F. Bornman (Eds.), *Environmental UV radiation: Impact on ecosystems and human health and predictive models* (pp 71–84, *Environmental UV Radiation: Impact on Ecosystems and Human Health and Predictive Models*). Dordrecht: Springer, Netherlands, Heidelberg. [https://doi.org/10.1007/1-4020-3697-3\\_6](https://doi.org/10.1007/1-4020-3697-3_6)
451. Carvalho, L. C., & Amâncio, S. (2019). Cutting the Gordian Knot of abiotic stress in grapevine: From the test tube to climate change adaptation. *Physiologia Plantarum*, 165(2), 330–342. <https://doi.org/10.1111/ppl.12857>
452. Day, T. A., Bliss, M. S., Tomes, A. R., Ruhland, C. T., & Gue-non, R. (2018). Desert leaf litter decay: Coupling of microbial respiration, water-soluble fractions and photodegradation. *Global Change Biology*, 24(11), 5454–5470. <https://doi.org/10.1111/gcb.14438>
453. Parton, W., Silver, W. L., Burke, I. C., Grassens, L., Harmon, M. E., Currie, W. S., King, J. Y., Adair, E. C., Brandt, L. A., Hart, S. C., & Fasth, B. (2007). Global-scale similarities in nitrogen release patterns during long-term decomposition. *Science*, 315, 361–364. <https://doi.org/10.1126/science.1134853>

## Authors and Affiliations

P. W. Barnes<sup>1</sup>  · T. M. Robson<sup>2,3</sup>  · R. G. Zepp<sup>4</sup>  · J. F. Bornman<sup>5</sup>  · M. A. K. Jansen<sup>6</sup>  · R. Ossola<sup>7</sup>  · Q.-W. Wang<sup>8</sup>  · S. A. Robinson<sup>9</sup>  · B. Foereid<sup>10</sup>  · A. R. Klekociuk<sup>11</sup>  · J. Martinez-Abaigar<sup>12</sup>  · W.-C. Hou<sup>13</sup>  · R. Mackenzie<sup>14,15</sup>  · N. D. Paul<sup>16</sup> 

✉ P. W. Barnes  
pwbarnes@loyno.edu

✉ T. M. Robson  
matthew.robson@helsinki.fi

- <sup>1</sup> Biological Sciences and Environment Program, Loyola University New Orleans, New Orleans, USA
- <sup>2</sup> Organismal & Evolutionary Biology (OEB), Faculty of Biological and Environmental Sciences, Viikki Plant Sciences Centre (ViPS), University of Helsinki, Helsinki, Finland
- <sup>3</sup> National School of Forestry, University of Cumbria, Ambleside, UK
- <sup>4</sup> ORD/CEMM, US Environmental Protection Agency, Athens, GA, USA
- <sup>5</sup> Food Futures Institute, Murdoch University, Perth, Australia
- <sup>6</sup> BEES, University College Cork, Cork, Ireland
- <sup>7</sup> Atmospheric Chemistry Observations and Modeling Laboratory, National Center for Atmospheric Research, Boulder, USA
- <sup>8</sup> Institute of Applied Ecology, Chinese Academy of Sciences (CAS), Shenyang, China
- <sup>9</sup> Global Challenges Program & School of Earth, Atmospheric and Life Sciences, Securing Antarctica's Environmental Future, University of Wollongong, Wollongong, Australia
- <sup>10</sup> Environment and Natural Resources, Norwegian Institute of Bioeconomy Research, Ås, Norway
- <sup>11</sup> Antarctic Climate Program, Australian Antarctic Division, Kingston, Australia
- <sup>12</sup> Faculty of Science and Technology, University of La Rioja, Logroño (La Rioja), Spain
- <sup>13</sup> Department of Environmental Engineering, National Cheng Kung University, Tainan City, Taiwan
- <sup>14</sup> Cape Horn International Center (CHIC), Puerto Williams, Chile
- <sup>15</sup> Millennium Institute Biodiversity of Antarctic and Subantarctic Ecosystems (BASE), Santiago, Chile
- <sup>16</sup> Lancaster Environment Centre, Lancaster University, Lancaster, UK